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(54) Title: **METHODS AND COMPOSITIONS FOR DETECTING THE ACTIVATION STATE OF THE MULTIPLE PROTEINS IN SINGLE CELLS**

(57) Abstract: The invention provides methods and compositions for simultaneously detecting the activation state of a plurality of proteins in single cells using flow cytometry. The invention further provides methods and compositions of the screening for bioactive agents capable of coordinately modulating the activity of a plurality of proteins in single cells. The methods and compositions can be used to determine the protein activation profile of a cell for predicting or diagnosing a disease state, and for monitoring treatment of a disease state.

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Methods and Compositions for Detecting the Activation State of Multiple Proteins in Single Cells

This application is a continuing application and claims the benefit of United States provisional
5 applications serial number 60/304,434, filed July 10, 2001, and serial number 60/310,141, filed
August 2, 2001.

The invention was made with United States Government support under Grant Numbers
P01-AI39646, AR44565, AI35304, N01-AR-6-2227, and A1/GF41520-01 awarded by the National
10 Institutes of Health. The United States Government has certain rights in the invention.

Field of The Invention

The invention relates generally to the field of protein detection using flow cytometry. More
specifically, the invention relates to simultaneously detecting the activation states of multiple
15 proteins in single cells using flow cytometry and, more particularly, using polychromatic flow
cytometry.

Background of the Invention

Proteins are the major components of cells. The spatiotemporal expression pattern and the
20 subcellular localization of proteins determines the shape, structure, and function of cells.
Proteins are assembled from 20 different amino acids, each with a distinct side chain and
chemical property. This provides for enormous variety in the chemical properties of proteins and
the activities they exhibit.

25 In addition, many proteins are dynamically regulated such that their activity is altered in response
to certain intrinsic or extrinsic cues. One form of regulation involves covalent modification of the
regulated protein. An example of such a covalent modification is the substitution of a phosphate
group for an hydroxyl group in the side chain of an amino acid (phosphorylation). Such
modification is often associated with an alteration in the activity of a protein. Protein kinases can
30 recognize specific protein substrates and catalyze the phosphorylation of serine, threonine, or
tyrosine residues on their protein substrates. Such substrate proteins capable of being
phosphorylated are referred to as phosphoproteins. Once phosphorylated, a substrate protein
may have its phosphorylated residue converted back to an hydroxylated one by the action of a
protein phosphatase which specifically recognizes the substrate protein. Protein phosphatases
35 catalyze the replacement of phosphate groups by hydroxyl groups on serine, threonine, or
tyrosine residues. Through the action of kinases and phosphatases a protein may be reversibly
phosphorylated on a multiplicity of residues and its activity may be regulated thereby.

Many kinases and phosphatases are known and play important roles in signal transduction. Many kinases and phosphatases are phosphoproteins, the kinase and phosphatase activities of which are regulated by phosphorylation. The phosphorylation of kinases and phosphatases is often regulated by other or same type kinases and phosphatases. In this way, the regulators of signal transduction are themselves regulated as part of a signaling cascade.

Many intracellular kinases and phosphatases transduce signals in response to extracellular signals that stimulate receptors at the cell surface. As a result, extracellular signals induce changes in the phosphorylation and activation states of many proteins in receptive cells. For example, many mitogens induce JNK and MAPK signaling activity which causes cell to proliferate. As another example, neurotrophic factors have been shown to induce AKT signaling activity which supports neuronal survival. In addition, many diseases, such as cancer, are associated with an alteration in the activation state of many signaling proteins. In many instances, the signaling activity observed in cancer cells is reminiscent of growth factor stimulated signaling activity in non-cancer cells.

Some lipids of the cell membrane are also substrates for kinases. For example, phosphatidylinositol 3-kinase (PI3K) specifically recognizes phosphatidylinositol 4, 5-diphosphate (PIP2) and catalyzes its phosphorylation at the 3' position of the inositol moiety. PIP2 is generated by the activity of another kinase, phosphatidylinositol 4-kinase which catalyzes phosphorylation of phosphatidylinositol at the 4' and 5' positions of the inositol moiety. PI3K regulates a variety of cellular functions and is activated by growth factors such as nerve growth factor (NGF), platelet derived growth factor (PDGF), insulin receptor substrate 1 (IRS-1) and CD28. Many of the proteins that activate PI3K also activate GTPases, predominantly the ras family, which in turn activate PI3K by binding to its p110 subunit. The direct pathway from tyrosine phosphorylated receptors to PI3K and activation of PI3K by GTPase proteins function synergistically. PI3K is a heterodimeric enzyme composed of a catalytic subunit (p110) and a regulatory subunit. Several genes encoding regulatory subunits of PI3K have been identified, including p85 α and p85 β . In addition, a number of splice variants of p85 α are known (including p55 α and p50 α).

It is understood that kinase signaling cascades play an important role in nearly every critical decision process in cells (for review see Hunter, Cell 100:113-127, 2000). Determining the role of specific signal transduction pathways in a given system has been aided by the advent of pharmacological inhibitors for specific kinases. However, monitoring activities for such kinases, for example, protein kinase B/AKT, cJun-N-terminal kinase (JNK), p38 mitogen activated protein

kinase (p38), p44/42 ERK1/2 (ERK), PKA, PKC or TYK2, is usually dependent upon in vitro kinase assays or more recently by immunoblotting to determine the phosphorylation state of these proteins using phospho-specific antibodies. To date it has not been possible to correlate rare subpopulations of cells with the activation state of kinases in important signaling pathways.

5

Another form of protein regulation involves proteolytic cleavage of a peptide bond. While random or misdirected proteolytic cleavage may be detrimental to the activity of a protein, many proteins are activated by the action of proteases that recognize and cleave specific peptide bonds. Many proteins derive from precursor proteins, or pro-proteins, which give rise to a mature isoform of the protein following proteolytic cleavage of specific peptide bonds. Many growth factors are synthesized and processed in this manner, with a mature isoform of the protein typically possessing a biological activity not exhibited by the precursor form. Many enzymes are also synthesized and processed in this manner, with a mature isoform of the protein typically being enzymatically active, and the precursor form of the protein being enzymatically inactive. This type of regulation is apparently not reversible. Thus, to inhibit the activity of a proteolytically activated protein, mechanisms other than *de-cleavage* must be used. It is the case that many proteolytically activated proteins are relatively short lived proteins.

Among the enzymes that are proteolytically activated are the caspases. The caspases are an important class of proteases that mediate programmed cell death (referred to in the art as "apoptosis"). Caspases are constitutively present in most cells, residing in the cytosol as a single chain proenzyme. These are activated to fully functional proteases by a first proteolytic cleavage to divide the chain into large and small caspase subunits and a second cleavage to remove the N-terminal domain. The subunits assemble into a tetramer with two active sites (Green, Cell 94:695-698, 1998).

Antibodies are particularly useful for the study of proteins. Antibodies are a large family of glycoproteins that specifically bind antigens, and do so with a high affinity. Antibodies comprise a constant domain and a variable domain. The variable domain of an antibody binds to antigen. Polyclonal antibodies comprise a multiplicity of variable region types. Monoclonal antibodies comprise a single type of variable region, and are thus more likely to specifically bind to fewer proteins. Monoclonal and polyclonal antibodies are routinely generated by means known in the art, and have been raised against many known proteins for use in both research and therapy. Antibodies may be used to determine the presence of antigens or proteins to which they specifically bind. Well known immunochemical methods that employ this approach include Western blotting, immunoprecipitation, and enzyme linked immunoassay (ELISA).

In particular, antibodies may be used to determine the presence of a specific isoform of a protein, especially, when the isoform possesses an antigen not present in other isoforms of the protein, or when the isoform lacks an antigen not present in other isoforms of the protein. Thus, the isoform can be antigenically distinct from other isoforms of the same protein. The antigenically distinct
5 isoform may uniquely possess or lack a covalently attached moiety, or may be in a conformation distinct from other isoforms and thereby present the same amino acid sequence in an antigenically distinguishable way.

For example, the determination of PI3K activity has typically involved immunoprecipitation of
10 PI3K and an in vitro enzymatic assay using phospholipids and labeled ATP. Recently, antibodies that specifically bind to phosphatidylinositol 4,5-diphosphate and phosphatidylinositol 3,4,5-triphosphate have been developed (Molecular Probes, A-21327 anti-phosphatidylinositol 4,5-diphosphate, mouse IgM monoclonal 2C11; A-21328 anti-phosphatidylinositol 3,4,5-triphosphate, mouse IgM monoclonal RC6F8). These antibodies serve as useful tools for the
15 detection of PI3K substrate (PIP2) and product (PIP3), and can be used to determine the activity of PI3K.

Biochemical investigations of protein expression and function have traditionally focused on one or a few proteins at a time. This has largely been due to limitations imposed by the available
20 investigative techniques. It is well understood that a cell's normal and abnormal physiology is the product of a large number of proteins participating in a large number of molecular interactions. For the purposes of designing treatments and cures, an understanding of the mechanisms underlying disease is desired. Such an understanding requires consideration of multiple proteins and their molecular interactions over time. In this regard, one important experimental hurdle is
25 how to analyze large numbers of proteins in a single experiment, for example to determine a protein expression profile for a single protein sample, or even for a single cell.

In addition to qualitative measurements of protein expression in cells, quantitative measures of protein expression are desirable. It is well known that many proteins can function differently at
30 different expression levels, and such differences in protein function can lead to differences in cell physiology.

It is highly desirable to determine expression levels of particular isoforms of proteins in a sample, given that different isoforms of a protein can have different activities. For example, proteins
35 involved in signal transduction can often exist in two or more forms, and it is usually a particular

form of the signaling protein that transmits a signal that mediates the effects of a signaling pathway. Conversion from one form to another is often used to turn a signal on or off through a change in protein activity.

- 5 In addition, in order to study protein expression and function, it is often necessary to immobilize proteins on a solid support. In Western blot analysis, proteins of interest are first separated by electrophoresis and then transferred and immobilized onto a nitrocellulose or a polyvinylidene difluoride (PVDF) membrane. In the phage display screening of a protein expression library, several hundred thousand proteins expressed by phages are immobilized on membranes. In
10 both Western blotting and phage display screening, proteins are immobilized non-covalently. The protein of interest is then selected by its unique property, i.e., interaction with an antibody. In some other applications such as immunoprecipitation and affinity purification, agents (e.g., antibodies, ligands) are covalently conjugated onto solid supports (e.g., agarose beads) through their primary amines, sulfhydryls or other reactive groups. In general, proteins retain their abilities
15 of interacting with other proteins or ligands after immobilization. However, even with the immobilization of a multiplicity of proteins from a sample, the problems of simultaneous detection of protein expression, protein form, and protein level for a multiplicity of proteins remains.

Thus, an object of the present invention is to overcome the problems described above.

- 20 Accordingly, the present invention provides an approach for the simultaneous determination of the activation states of a plurality of proteins in single cells. This approach permits the rapid detection of heterogeneity in a complex cell population based on protein activation states, and the identification of cellular subsets that exhibit correlated changes in protein activation within the cell population. Moreover, this approach allows the correlation of cellular activities or properties,
25 such as surface molecule expression or cell granularity, with protein activation at the single cell level.

Summary of the Invention

- In accordance with the above objects, the present invention provides methods and compositions
30 for simultaneously detecting the activation state of a plurality of activatable proteins in single cells using flow cytometry. The invention further provides methods and compositions of screening for bioactive agents capable of coordinately modulating the activity or activation state of a plurality of activatable proteins in single cells. The methods and compositions can be used to determine the protein activation profile of a cell for predicting or diagnosing a disease state, and for monitoring
35 treatment of a disease state. Further, the methods and compositions of the present invention can be used optionally to sequentially detect the activation state of a plurality of activatable proteins

in single cells. In addition, the methods and compositions of the present invention can be used optionally detect the activation state of a single protein or modulate the activity or activation state of a single protein.

- 5 The invention provides populations of cells, single cells, cell lysates, proteins, and samples comprising populations of cells, single cells, cell lysates, proteins useful in the methods of the present invention. In particular, the invention provides activatable proteins and activation state-specific antibodies that bind to a specific isoform of an activatable protein. In one aspect, the activation state-specific antibodies are conjugated to a label, preferably a fluorescent label, and
10 more preferably a FRET label.

- In one aspect, the invention provides methods of detecting the activation state of at least a first and a second activatable protein in single cells, the method comprising the steps of: a) providing a population of cells comprising the first and the second activatable proteins; b) contacting the
15 population of cells with a plurality of activation state-specific antibodies, wherein the plurality of activation state-specific antibodies comprise: i) at least one first activation state-specific antibody that is capable of binding to a corresponding isoform of the first activatable protein in the population of cells; and ii) at least one second activation state-specific antibody that is capable of binding to a corresponding isoform of the second activatable protein in the population of cells;
20 and c) using flow cytometry to detect the binding of the first and second activation state-specific antibodies in single cells of the population of cells, wherein the binding of the first activation state-specific antibody is indicative of a specific activation state of the first activatable protein, and the binding of the second activation state-specific antibody is indicative of a specific activation state of the second activatable protein.

25

In a further aspect, the first activatable protein is a kinase. Also in a further aspect, the first activatable protein is a caspase.

- In a further aspect, the first activatable protein is a first kinase and the second activatable protein
30 is a second kinase. Also in a further aspect, the isoform of the first kinase is a first phosphorylated kinase, and the isoform of the second kinase is a second phosphorylated kinase. In another aspect, the first activation site-specific antibody binds to the first phosphorylated kinase, and the second activation site-specific antibody binds the second phosphorylated kinase.

- 35 In a further aspect, the first activatable protein is a first caspase and the second activatable protein is a second caspase. Also in a further aspect, the isoform of the first caspase is a cleaved product of a first pro-caspase, and the isoform of the second caspase is a cleaved

product of a second pro-caspase. In another aspect, the plurality of activation site-specific antibodies comprise a first activation site-specific antibody that binds to the isoform of the first caspase, and a second activation site-specific antibody that binds to the isoform of the second caspase.

5

In another aspect, the invention provides methods of detecting the activation state of at least a first activatable protein in single cells, the method comprising the steps of: a) providing a population of cells comprising at least the first activatable protein; b) contacting the population of cells with a plurality of substrates; wherein the plurality of substrates comprise at least a first
10 substrate that is capable of being modified by a corresponding isoform of the first activatable protein in the population of cells; and c) using flow cytometry to detect the modification of the first substrate in single cells of the population of cells, wherein the modification is indicative of a specific activation state of the first activatable protein.

15 In a further aspect, the population of cells further comprises a second activatable protein; the plurality of substrates further comprise a second substrate that is capable of being modified by a corresponding isoform of the second activatable protein in the population of cells; and step c) further comprises using the flow cytometry to detect the modification of the second substrate in single cells of the population of cells, wherein the modification of the second substrate is
20 indicative of a specific activation state of the second activatable protein.

In another aspect, the invention provides methods of detecting a protein activation state profile of single cells based on the activation state of at least a first activatable protein in the cells, the method comprising the steps of: a) providing a population of cells comprising at least the first
25 activatable protein; b) contacting the population of cells with a plurality of substrates; wherein the plurality of substrates comprise at least a first substrate that is capable of being modified by a corresponding isoform of the first activatable protein in the population of cells; c) contacting the population of cells with a plurality of activation state-specific antibodies, wherein the activation state-specific antibodies comprise at least one first activation state-specific antibody that is
30 capable of binding to a corresponding isoform of the first activatable protein in the population of cells; and d) using flow cytometry to simultaneously detect: i) the binding of the first activation state-specific antibody in single cells of the population of cells, wherein the binding of the first activation state-specific antibody is indicative of a specific activation, state of the first activatable protein; and ii) the modification of the first substrate the single cells, wherein the modification is
35 indicative of the specific activation state of the first activatable protein.

In a further aspect, the population of cells further comprises a second activatable protein; the

plurality of substrates further comprises a second substrate that is capable of being modified by a corresponding isoform of the second activatable protein in the population of cells; and step d) further comprises using the flow cytometry to detect the modification of the second substrate in the single cells, wherein the modification is indicative of a specific activation state of the the
5 second activatable protein.

In a further aspect, the plurality of activation state-specific antibodies further comprises a second activation state-specific antibody that is capable of binding to a corresponding isoform of the second activatable protein in the population of cells, and step c) further comprises using the flow
10 cytometry to detect the binding of the second activation state-specific antibody the single cells of the population of cells, wherein the binding of the second activation state-specific antibody is indicative of the specific activation state of the second activatable protein.

In another aspect, the invention provides methods of screening for a bioactive agent capable of
15 modulating the activity of at least a first activatable protein in cells, the method comprising: a) providing a population of cells, each of the cells comprising at least the a first activable protein, a second activatable protein, and a third activatable protein, wherein the first activable protein can activate the second activatable protein thereby forming a specific isoform of the second activable protein (isoform-2), and wherein the second activable protein can activate the third activatable
20 protein thereby forming a specific isoform of the third activatable protein (isoform-3); b) contacting the cells with a second activation state-specific antibody, a third activation state-specific antibody, and a candidate bioactive agent, wherein the second activation state-specific antibody is capable of binding to the isoform-2, and wherein the third activation state-specific antibody is capable of binding to the isoform-3; c) using fluorescent activated cell sorting (FACS)
25 to sort single cells of the population of cells based on the presence of the isoform-2 and the isoform-3; and d) determining the ratio of the isoform-2 to the isoform-3 in the single cells in the presence and absence of the candidate bioactive agent, wherein a difference in the ratio of the isoform-2 to the isoform-3 in the presence and absence of the candidate bioactive agent is indicative of the ability of the candidate bioactive agent to modify the activity of the first activable
30 protein.

In a further aspect, the first activatable protein is activated by an activating agent.

In a further aspect, the first activatable protein is a caspase. In another aspect, the first
35 activatable protein is a kinase; in an additional aspect, the kinase is PI3K; and in a further aspect the PI3K is activated by a growth factor or by activation of a cell surface receptor.

In a further aspect, the kinase is PI3K; the second activatable protein is PIP2; the third activatable protein is PIP3; the isoform-2 is PIP 4,5 bisphosphate; and the isoform-3 is PIP 3,4,5.

5 In a further aspect, the first activatable protein is ICAM-2; the activity is apoptosis; the second activatable protein is PIP2; the third activatable protein is PIP3; the isoform-2 is PIP 4,5 bisphosphate; and the isoform-3 is PIP 3,4,5. In a further aspect step a) further comprises clustering the ICAM-2 with ICAM-3.

10 In a further aspect, the plurality of activation state-specific antibodies comprise at least one antibody selected from a group of antibodies consisting of: anti- AKT-phospho-Ser473, anti-AKT phospho-Thr308, anti-p44/42 MAPK phospho-Thr202/Tyr204, anti-TYK2 phospho-Tyr1054/1055, anti-p38 MAPK phospho-Thr180/Tyr182, anti-JNK/SAPK phospho-Thr183/Tyr185, anti-phospho-tyrosine, anti-phospho-threonine, anti-PIP2, and anti-PIP3.

15 In a further aspect, of the step a) further comprises contacting the population of cells with an agent that induces the activation of at least the first activatable protein.

In a further aspect, step a) further comprises contacting the population of cells with an agent that induces the activation of at least one of the first and the second activatable proteins.

20

The method according to any of claims 15-22, wherein step a) further comprises contacting the population of cells with an agent that induces the activation of at least one of the first, the second, and the third activatable proteins.

25 In a further aspect, the methods further comprises sorting the single cells based on the activation state of the first activatable protein, and the activation state of the second activatable protein.

30 In a further aspect, the first activation state-specific antibody comprises a first label, wherein the second activation state-specific antibody comprises a second fluorescent label and, wherein the sorting is by fluorescent activated cell sorting (FACS).

In a further aspect, the first activation state-specific antibody comprises a first FRET label; the second activation state-specific antibody comprises a second FRET label and the sorting is by fluorescent activated cell sorting (FACS).

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In a further aspect, the first activation state-specific antibody comprises a FRET label; the second activation state-specific antibody comprises a label; and the sorting is by fluorescent activated

cell sorting (FACS).

In a further aspect, step a) further comprises fixing the cells.

- 5 In a further aspect, the cells are mammalian cells.

Brief Description of the Drawings

- Figure 1** depicts the results of experiments demonstrating that phospho-specific antibodies correlate with kinase activity. A) Phospho specific antibodies for i) p44/42, ii) p38, iii) JNK/SAPK, iv) AKTser473/AKTthr308, v) TYK2, vi) PKA-substrate, and vii) PKC-PAN substrate were tested for phospho-specificity by specific kinase inducing and inhibitory conditions. The inducing conditions for kinase are shown as well as specific inhibitor(s) of the pathway. The ability of the kinase to have activity upon a known substrate is shown beneath each phospho-western set. Non-phospho specific antibodies for AKT, p3B, SAPK JNK, p44/42, and TYK2 reflect total kinase present. Kinase activity was determined under conditions found to elicit phosphorylation of respective kinase and correlated with the phosphorylated state of the kinase as detected by phospho-specific antibodies. Given the nature of the phospho-PKA substrate and phospho-PKC-PAN substrate antibodies, direct kinase activities could not be measured. Conditions used determined a general activation of PKC and PKA, since both antibodies recognize multiple PKC and PKA target proteins in the phosphorylated form of their respective kinase substrate recognition motif. Immunoblots/kinase assays for AKT, p44/42, p38, and JNK kinases were performed in NIH3T3 and data for TYK2, PKC, and PKA were performed in Jurkat cells (see Material and Methods for elaboration on treatment conditions).
- B) Jurkat cells were treated with a kinase inducing agent (induced), or kinase inhibitor (inhibited) before induction conditions as diagramed in Table 2. Cells were prepared for intracellular FACS staining and analyzed for kinase activities noted in the figures by log fluorescence of phospho-specific antibody-fluorophore conjugate. Plot overlays illustrate shifts respective of either increased or decreased kinase activity. The bottom right panel represents a dose response curve of EGF treatment as measured by phospho-ERK activity (0.001, 0.01, 0.1, 1.0, 10 μ g/ml). The inhibitors and activators used were the following: p44/42 (ERK1/2), was activated with epidermal growth factor (EGF) and inhibited with MEK1/2 inhibitor U0126²¹⁻²³; AKT activation was achieved by addition of growth media and platelet derived growth factor and inhibited by incubation with PI3K kinase inhibitor LY294002^{20*}; JNK was activated by anisomycin^{29*} and inhibited by staurosporine; p38 MAPK was activated by anisomycin^{29*} and inhibited with compound SB203580^{24,29*}; PKA was activated by phorbol 12 myristalated

13-acetate (PMA) and phospho detection was made possible by incubation with protein phosphatase inhibitor calyculin A²⁵; PKC was activated by ionomycin (IO) and inhibited using bisindolymaleimide II^{26,27}; TYK2 activity was induced by interferon γ (IFN γ) and inhibited using the general tyrosine kinase inhibitor genistein^{28,3*}. *See the corresponding reference number and cite in Example 1 under "References."

C) TYK2, PKA and PKC substrate probes are phospho specific. Treatment of stimulated samples (stated above) with lambda phosphatase prior to intracellular staining decreased detection by phospho-TYK2 and phospho-PKA substrate probes. Treatment of cells with calf intestinal phosphatase in addition to lambda phosphatase was required for decreased detection of phosphorylated PKC substrates.

D) Phosphorylation of p44/42 correlates with mean fluorescent intensity as measured by flow cytometry. Semiquantitative analysis of p44/42 phosphorylation as measured by both phospho specific detection and mean fluorescence intensity (MFI) by phospho-p44/42 kinase probe. Jurkat cells were subjected to a dose response curve of EGF at the indicated concentrations for 30 minutes, and assessed for p44/42 phosphorylation by both immunoblotting with phospho-specific antibodies and flow cytometry with developed p44/42 kinase probe. Y error bars are standard deviation for MFI values. X error bars are standard deviation for percent phosphorylated relative to unstimulated control. Values displayed are representative of 3 independent experiments. Error bars not visible are concealed by data point.

Figure 2 depicts the results of experiments demonstrating that multi-dimensional analysis provides insights into signaling networks as induced by differing stimuli.

25

A) Differential kinase activity within CD4+ and CD19+ lymphocyte subsets. PBMC were treated with the indicated stimulus for 1 hour and prepared for intracellular kinase phosphorylation status determination by flow cytometry. Both hyper and hypo responses can be observed with respect to p44/42 activity in CD4+ upon different stimuli.

30

B) T cell activation measured by CD69 presents differential p44/42 kinase activity to CD3/CD28 and PMA/ionomycin stimulation. The stimulation conditions are shown in the Figure.

C) Multiple kinase activities measured in artificial T cell stimulation. Jurkat cells in growth media and PMA/ionomycin (IO) treated were prepared for intracellular kinase phosphorylation status determination by flow cytometry. CD69 expression serves to indicate T cell activation. X-axis corresponds to log fluorescence values, Y-axis corresponds relative cell number.

D) Phosphorylation of p38 as a function of cell granularity. Simultaneous measurement of phosphorylated and non-phosphorylated p38 exemplifies a shift from non-phosphorylated to phosphorylated p38 as cell granularity increases (indicated by the shaded bar in the side scatter parameter).

5

E) 3-parameter analysis of p44/42 and p38 kinase activity as a function of JNK activity illustrate potential mapping of signaling crosstalk. PBMC were treated with IL-1 α . JNK, p44/42, and p38 activity was simultaneously measured by flow cytometry. Shaded bar in phospho-JNK/SAPK plot indicates intensity of JNK activation.

10

F) 5-parameter analysis of AKT, p38, PKA, and p44/42 activity as a function of TYK2 activity in IL-1 β treated PBMC. At initiation of TYK2 activity, all 4 kinase activities are not detectable. Synchronous activation of p38, p44/42, and AKT activity is observed at a greater level of TYK2 activity. PKA activity is observed at a greater level of p38 and p44/42 activity.

15

Figure 3 shows a flow diagram of resting T cell isolation, preparation, and analysis by polychromatic flow cytometry (11-color, 13 parameter).

Isolation of lymphocytes was achieved by a Ficoll-plaque gradient and depleted for adherent cells. Flow cytometry plots illustrate typical lymphocyte gate and presence of CD3⁻ populations. Magnetically activated cell sorting eliminated activated T cells, NK cells, remaining macrophages, and B cells, enriching for resting T cells. The enriched population was assessed prior to stimulation to ensure presence of CD4⁺ and CD8⁺ cells. Stimulation to IL-1 α , IL-1 β , or IL-2 was performed and cells were prepared for flow cytometry using stains depicted in Table 1.

Occasionally, downregulation of some surface markers was observed to certain stimuli and is presented by receptor downregulation of key molecules, namely CD3, CD4, CD8, (apparent when comparing flow cytometry plots of these markers prior to stimulus treatment). An example of memory and naive gating by 5 differentiation markers is depicted for both CD4⁺ and CD8⁺ T cells. Appropriate placement of CD45RA gate was done according to De Rosa et al and was carried out separately for CD4⁺ and CD8⁺ populations. After identification of naive and memory cells by multiple-gating according to differentiation markers, intracellular kinase activity was determined. Kinase profiles are plotted against CD11a markers, although they are gated for all naive and memory markers. Assessment of multiple kinase activities was made possible by combining two 11-color experiments.

35

Figure 4 depicts the results of experiments demonstrating that signaling profiles are readily discernible in particular lymphocyte subsets using polychromatic flow cytometry.

Naive and memory T cell kinase profiles are overlaid for direct comparison. Mechanisms involving threshold dependent kinase activation can be observed when kinase phosphorylation approaches but does not exceed Log 10 fluorescence intensity (value defined from inhibitor/inducer experiments Fig. 2) in CD8+ naive cells. Hyporesponses can be seen for IL-1 α CD4+ naive cells.

Figure 5 depicts the results of experiments monitoring intensities of both active and non-active kinases by flow cytometry

- 10 A) Phospho and non-phospho-specific probes for p44/42, p38, AKT, TYK2, and JNK detect respective kinase in its native state. Jurkat cells were treated to the following: AKT activity stimulated with PDGF, MAPK activity stimulated with EGF, p38 and JNK activity stimulated with anisomycin - for 10 minutes to induce initial activation of respective kinase with inducers as per the concentrations stated in Table 2. Phospho-p44/42, phospho-p38, phospho-AKTser473, and
- 15 phosphoAKTthr308, and phospho-JNK/SAPK detected distinct populations upon a given stimulus in respect to their non-phospho specific counterparts. AKT positive cells were gated and plotted against AKTser473- and AKTthr308 phosphorylation plot as denoted in panel. Unstimulated cells for all non-phospho specific kinases are appropriately displayed for comparison.
- 20 **Figure 6** depicts the results of experiments demonstrating that LFA-1 stimulus induces JAB1 and Cytohesin-1 release and phosphorylation in human naïve CD4+ T cells. A) Phospho-cJun immunoblot of nuclear extracts from cells stimulated with CD3, CD28 and sICAM-2 (5 μ g/ml, 30 min). B) Single cell analysis of phospho-cJun(S63) in α -JAB1, IgG transfected and untransfected cells. The median fluorescence intensity is plotted \pm SD. C) Confocal of JAB1 and cytohesin-1 unstimulated (panel A-C), LFA-1 stimulated (sICAM-2 5 μ g/ml) (panel D-F), and α -JAB1
- 25 transfected cells (G-I). Panels G-I were not stained with α -JAB1 antibody. D) Immunoprecipitates of JAB1 and Cytohesin-1 from unstimulated, sICAM-2 and CD3 (5 μ g/ml, 30 min) stimulated cells were immunostained for phospho-serine and phospho-threonine and subsequently stripped and stained for α -JAB1 and α -cytohesin-1 respectively. E) Flow cytometric
- 30 detection of phospho-p44/42(T202/Y204) in IgG, α -JAB1, α -cytohesin-1 N-terminal, α -cytohesin-1 C-terminal, and C-terminal cytohesin-1 peptide transfected cells stimulated with sICAM-2 (5 μ g/ml, 30 min).

Supplementary Figure 6 depicts the results of experiments demonstrating: A) Kinetic analyses of CD3, CD28, and LFA-1 stimulus of phospho-p38(T180/Y182), phospho-AKT(S473), phospho-Lck(S158), phospho-mek1/2(S217/221), phospho-elk(S38) by flow cytometry. We observed phospho-AKT, phospho-p38, and phospho-mek1/2 to be induced by CD28 stimulation to a

greater extent than CD3 or LFA-1 stimulus. These reports are consistent with recent findings suggesting the activation of these pathways by CD28 stimulation (Kane et al., 2001; Li et al., 2001). We noted that phospho-Ick was induced over time in the LFA-1 stimulus differentially with respect to CD3 and CD28 stimulation. Phosphorylation of elk-1 was imparted by CD3 and CD28 stimulation, and to a lesser degree by LFA-1 stimulus.

Figure 7 depicts the results of experiments demonstrating CD3/CD28/LFA-1 signaling in human naïve T cells by phospho-protein antibody array. A) Differential phospho-protein profile of CD3/CD28/LFA-1 vs. CD3/CD28 stimulated naïve CD4⁺ T cells. Ratios of the fluorescent phospho-protein array were calculated from the following equation where MFI equals mean fluorescent intensity: $\text{Log} [(MFI_{\text{CD3/CD28/LFA}})/(MFI_{\text{CD3/CD28}})]$, (see Materials and Methods). Fold-phosphorylation is depicted by the color intensity scale where green corresponds to high and red corresponds to low. B) Cell cycle analysis of CD4⁺ T cells stimulated with CD3, CD28, and sICAM-2. Cells in G0, G1, S+G2/M phases of the cell cycle were determined using double staining for expression of Ki-67 and DNA content. Low Ki-67- signals were considered cells in G0, Ki-67+ signals with 2n DNA were considered cells in G1 and Ki-67+ signals with 2n DNA were considered to be in S or G2/M.

Supplementary Figure 7 depicts the results of experiments, per continuation of Figure 8, demonstrating that the cells are naïve gated CD4⁺ T cells (as indicated by 5 differentiation markers) for individual stimulations of sICAM-2 and CD28, as well as the CD3/sICAM-2 combination. B) CD25 expression post 24 and 48 hr stimulus (as indicated, 10 µg/ml).

Figure 8 shows a flow diagram depicting multidimensional analysis of naïve CD4⁺ T cells. Magnetic cell sorting, 13-parameter flow cytometry, transcription factor profiling, and cytokine bead arrays were multiplexed to determine stimulations effect on immunophenotyped naïve human CD4⁺ T cells (CD3⁺CD4⁺CD8⁻CD45RA⁺CD62L⁺CD11a^{dim}CD27⁺CD28⁺). Intracellular levels of cytokines and phospho-proteins were correlated with surface phenotypes/activation markers as indicated in figure.

Supplementary Figure 8 depicts the results of experiments demonstrating the transcription factor profiling and cytometric bead array of CD3, CD28, and LFA-1 stimulation. A) Nuclear extracts of stimulated cells were prepared and hybridized to 96 well plates coated with consensus sequences to p65, p50, cFos, creb, atf2, and cRel. Bound protein was detected using standard ELISA techniques. CD3 stimulation presented low levels of the NF-κB subunits p65 and p50 and no detectable levels of cFos, cRel, CREB, effects of which were similar to LFA-1 stimulation. However, CD3 stimulation alone presented higher levels of atf2 than CD28 and LFA-1 stimulus. CD28 stimulation alone presented low levels of the NF-κB p65 and p50 subunits, with

elevated levels of CREB. LFA-1 stimulation presented slightly higher levels of NF- κ B p65 subunit and cRel. B) Cytometric bead array (CBA) for secreted IFN γ , IL-10, IL-2, IL-4, IL-5, IL-2, and TNF α to CD3/CD28 and CD3/CD28/LFA-1 stimulus (10 μ g/ml, 16 hr). Values were computed to standard curve using recombinant cytokines and were simultaneously measured from the
 5 supernatant of treated cells. Measurement of cytokine secretion after CD3 and LFA-1 stimulus alone did not report significant cytokine secretion CD28 stimulation gave elevated levels of IL-4 and IL-5, with low levels of IFN γ .

Figure 9 depicts the results of experiments demonstrating T cell activation as monitored by
 10 intracellular IL-2, and surface CD25 and CD69 markers. A) Intracellular detection of IL-2 of titration of CD3/CD28 vs. LFA-1 stimulus or CD3 vs. CD28 stimulus for 6 hours. Median Fluorescent intensity is plotted as a visual array. B) Bivariate analysis of CD69 and CD25 expression in unstimulated, CD3/CD28/LFA-1 (0.001 μ g/ml, 1:1:1) stimulated, and CD3/CD28 (10 μ g/ml) stimulated CD4 T cells. C) Bivariate plot of CD69 expression and intracellular detection of
 15 IL-2 post 12 hour stimulation of unstimulated, CD3/CD28, and CD3/CD28/LFA-1 stimulation.

Figure 10 depicts the results of experiments demonstrating the profiling of transcription factor nuclear localization and activity. A) Reporter gene assays of NFAT-luciferase, NF- κ B -luciferase, and AP-1-luciferase transfected Jurkat cells. CD3, CD3/CD28, and CD3/CD28/LFA-1 stimulus
 20 was performed (6 hr, 10 μ g/ml) and fold luciferase activity is plotted. B) Nuclear profile of transcription factors (as indicated in figure) to CD3, CD28, and LFA-1 stimulus (10 μ g/ml, 12 hr). Nuclear extracts (5 μ g/ml) were hybridized to transcription factor specific consensus sequence DNA coated plates, and detected by ELISA. Values were normalized to negative control and displayed as absorbance units of 655 nm. Nuclear profile of CD3/CD28 and CD3/CD28/sICAM-2
 25 stimulated cells (as described above). C) NFAT-luciferase reporter activity on CD3/CD28 and CD3/CD28/LFA-1 titrations (as indicated) as described above.

Figure 11 depicts the results of experiments demonstrating the intracellular production of IL-4 and IFN γ in human naïve CD4 T cells. Intracellular levels of IFN γ and IL-4 post 6 hr stimulus of
 30 titrations of CD3 vs. CD28 and CD3/CD28 vs. LFA-1 stimulus. Median fluorescence values were normalized to unstimulated cells, and log ratios of IL-4/IFN γ were computed and plotted as intensity contour plots. Intensity levels are depicted in range indicator (right).

Figure 12 depicts the results of experiments demonstrating cytometric bead arrays and
 35 Cytometric bead array (CBA) for secreted IFN γ , IL-10, IL-2, IL-4, IL-5, IL-5, and TNF α to

CD3/CD28 and CD3/CD28/LFA-1 stimulus (10 µg/ml, 16 hr). Values were computed to a standard curve using recombinant cytokines and were simultaneously measured from the supernatant of treated cells using CBA arrays from PharMingen.

5 **Figure 13** depicts the results of experiments demonstrating clustering of ectopic ICAM-2 in NIH3T3 activated the p44/42 MAPK cascade. A) ICAM-2 and ICAM2-AC retroviral expression vectors target to the cell surface. Plasma membrane targeting of ICAM-2 and ICAM2-AC transduced NIH3T3 was verified by flow cytometry using specific anti-human antibodies to respective termini. B) Western -blot verified ICAM-2 and ICAM2-AC expression to compare with
10 endogenous ICAM-2 from Jurkat cells. C) p44/42 MAPK cascade is initiated upon surface ICAM-2 clustering. Serum starved ICAM-2 cells were ICAM-2 crosslinked (10 µg/ml) for the indicated time by an anti-human ICAM-2 monoclonal antibody. Cell lysates were harvested and subjected to standard electrophoretic and immunoblotting protocols. Phospho specific antibodies for RAF, MEK, p44/42 MAPK (ERK1 /2), Elk-1, and Rsk1 were used to detect phosphorylated
15 proteins. Blots were then stripped and reprobed using a normalizing antibody (non-phospho specific antibody) to the each respective kinase. D) ICAM-2 clustering induces p44/42 MAPK activity as determine by p44/42 MAPK kinase activity assay.- Serum starved cells were crosslinked for ICAM-2 (10 µg/ml) for 30 minutes and assessed for p44/42 MAPK activity. Positive and negative denoted lanes indicate internal MAPK kinases controls and a control IgG
20 served as a negative control for induction condition.

Figure 14 depicts the results of experiments demonstrating that p44/42 MAPK activity mediates enhanced growth kinetics. A) Cell cycle re-entry in initiated upon ICAM-2 clustering. Cells were serum starved for 72 hours to synchronize cells in the absence of chemical agents and
25 subsequently ICAM-2 crosslinked (10 µg/ml, 18 hrs) and subjected to cell cycle analysis by flow cytometry. Errors bars denote standard deviation of triplicate experiments. Gating for specific cell cycle stages is presented (left). B) Contact inhibition induced cell cycle arrest as measured by the MTT cell viability assay. Indicated cell types were seeded in a 96 well plate at increasing cell density as indicated and cellular respiration was measured by incubating the cells with the
30 MTT reagent. Detection of product was done by reading absorbance at 580 nm 4 hrs post an initial 2-hour settling period. Error bars denote standard deviation of triplicate experiments. C) Growth kinetics monitored by cell density over time. Indicated cell types were seeded at 10⁴ cells and cell density was monitored by standard clonogenic assay over the indicated time period. Error bars denote standard deviation of triplicate experiments.

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Figure 15 depicts the results of experiments demonstrating that ICAM-2 binds surface β 1, β 2, and β 3 integrins. A) Affinity chromatography detects several ICAM-2 interacting surface proteins. An ICAM-2-FITC probe was generated by immuno-depletion of ICAM-2 from transduced NIH3T3, chemically conjugated to FITC (1st and 2nd panels), and tested by antiICAM2 immunoblot and fluorescent labeling. A farwestern approach utilizing ICAM-2-FITC as a labeled probe detects a 96 and 16 kDa interacting protein bands in ICAM-2 lysates (3rd panel). Surface proteins of ICAM-2 and vector control cells were biotinylated and subjected to affinity chromatography dual column system utilizing a constructed ICAM-2 coupled solid support and a streptavidin-agarose support. Several protein bands were identified that are both ICAM-2 interacting and surface expressed (biotinylation detected by streptavidin-FITC) (4th panel). B) ICAM-2 soluble protein binds to the surface of cells. The ICAM-2-FITC probe was used to label NIH3T3, Jurkat, and BaF3 cells and was detected by flow cytometry. Prior treatment of cells with trypsin attenuated the ICAM-2-FITC binding (right panel). C) ICAM-2 interacts with β 1, β 2, and β 3 integrins. Final elutions from the ICAM-2 solid support and subsequent streptavidin-agarose column passage were immunoblotted for β -integrins using monoclonal antibodies. Detection of β 1, β 2, and β 3 refined and verified the mass spectrometry reading of the MALDLMS analysis. D) ICAM-2 interaction with β 2 integrin induces p44/42 MAPK kinase activity. Beta-1, β 2, and β 3 integrin were blocked with monoclonal antibodies in serum starved cells prior to ICAM-2 crosslinking in ICAM-2, ICAM2-AC, and vector control cells. E) Integrin profile assessment by quantitative flow cytometry. Alpha-5, α L, β 1, β 2, β 3, β 4 monoclonal antibodies were conjugated to phycoerythrin (PE) and quantitated using QuantiBRiTE PE beads by flow cytometry (see materials and methods for elaboration). Linear regression analysis of fluorescent signal as a function of a standard antigen concentration curve allows quantitation of the number of surface molecules per cell. Error bars denote standard deviation of triplicate experiments. F) Soluble ICAM-2 induced p44/42 MAPK phosphorylation and activity is blocked by antibodies to α L integrin. ICAM-2 soluble protein was incubated for 30 minutes in serum starved Jurkat cells at the indicated concentrations in the presence or absence of monoclonal antibody pre-treatment to α L (similar to mAb blocking experiments). Immunoblot for phospho-p44/42 MAPK (left) and p44/42 MAPK activity assay (right).

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Figure 16 depicts the results of experiments demonstrating that ICAM-2/LFA-1 interaction is transmitted to Raf by PYK2 and SYK. Inhibitor screens using various pharmacological inhibitors to Src related kinases identify PYK2 and SYK to be necessary in relaying the ICAM-2 induced signal to Raf and p44/42 MAPK kinase. Serum starved cells were treated with 10 μ M of indicated compound 30 minutes prior to being ICAM-2 crosslinked for 30 minutes (10 μ g/ml) and assessed for phosphorylated Raf and p44/42 MAPK using phospho specific antibodies in standard immunoblot procedures. Control IgG dissolved in 0.1 % DMSO served as negative control. Blots

are representative of triplicate experiments. B) SYK and PYK interaction with LFA-1 subunits is intensified upon ICAM-2 interaction. Co-immunoprecipitation experiments were performed immunoprecipitating α L and β 2 integrin and immunoblotting for the presence of SYK and PYK as a function of a stimulus for 30 minutes as indicated (antibodies used at 10 μ g/ml, compounds used at 1 μ M and control 19 served as negative control). Reciprocal co-immunoprecipitations confirm PYK and SYK associations with β 2 integrin (bottom panel). C) PYK2 is phosphorylated in the presence of ICAM-2 or additional stimuli as determined by an antiphosphotyrosine immunoprecipitation and subsequent immunoblot for PYK2.

Figure 17 depicts result from experiments demonstrating that confocal microscopy reveals cellular the redistribution of PYK2 and SYK to the surface membrane in Jurkat cells after ICAM-2 interaction with LFA-1 and subsequent phosphorylation of PYK2. Jurkat cells were treated with soluble ICAM-2 protein (10 μ g/ml) or bovine serum albumin (Unstimulated, 10 μ g/ml) for 10 minutes and prepared for confocal microscopy (see material and methods). Cells were stained for PYK2, SYK2, and phosphorylated PYK2 at tyrosine 402 (denoted as PYK2 pTyr402). Panels A-F represent unstimulated cells and panels G-L represent ICAM-2 treated cells. Arrows serve to highlight marked differences in cellular distribution of PYK2 and SYK. Scale bar is denoted in lower left corner of panels (in micrometers). Panels G-L are at a higher magnification to see distribution differences clearly.

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Figure 18 depicts result from experiments demonstrating that cell-to-cell contact initiates p44/42 MAPK activity in LFA-1 expressing and AKT activity in ICAM-2 expressing cells as determined by flow cytometry based kineses assays. A) Jurkat cells and retrovirally transduced ICAM-2 cells were mixed for 30 minutes and subjected for flow cytometry by surface staining for CD3 and CD4 surface markers, and stained intracellularly for phospho-p44/42 MAPK and phospho-AKT using fluorescently conjugated phospho-specific monoclonal antibodies (see materials and methods for elaboration). Gating on high and low phosphorylated AKT and p44/42 MAPK populations, as determined for fluorescence log intensity, were analyzed according to the CD3 and CD4 surface markers. Plots depict CD3 vs. phospho-AKTser473 or phospho-p44/42 but are similar for CD4. B) Model of signaling pathways initiated upon intercellular ICAM-2/LFA-1 interaction. LFA-1, upon engagement of ICAM-2 activates the RAF/MEK/MAPK cascade, mediated through PYK2 and SYK. ICAM-2 expressing cells activate the AKT survival pathway as ICAM-2 is clustered on the cell surface.

Figure 19 depicts the results of experiments demonstrating that a retroviral library screen for anti-apoptotic molecules to staurosporine induced apoptosis identifies ICAM-2. A) cDNA library screening scheme. 10^7 NIH3T3 cells were infected with a Jurkat T cell retroviral cDNA library and treated with staurosporine ($1 \mu\text{M}$) for 24 hours to induce apoptosis (Stolzenberg et al., 2000).

5 Surviving clones were grown out for one week and subjected to a repeat selection three times. To determine which viral integrants encoded an antiapoptotic function, a phenotypic transfer assay was performed. An aliquot of the surviving cell population was infected with native Moloney murine leukemia virus (MMLV). This rescued the replication-defective viral vector proviruses, generating infectious particles. NIH3T3 cells were infected with the rescued viral

10 vectors and treated with staurosporine to confirm the anti-apoptotic phenotype. B) Complexity of populations surviving staurosporine induced apoptotic screen. RT-PCR was performed to assess the complexity of the surviving populations. Surviving population (1°) exhibited several discrete bands and the post-transfer population (2°) exhibited a single band. M- molecular weight marker. A negative control vector (LacZ), initially spiked into the library at 10% was

15 undetectable, while an independent infection of a Bcl-2-expressing vector was readily detected. C) Retroviral vector encoding ICAM-2 western blot analysis in infected NIH3T3. D) Retroviral vector encoding ICAM-2 ectopically expresses surface ICAM-2. Right Panel: BaF3 pro-B-cells infected with a retroviral vector construct encoding ICAM-2 or vector control and assayed for either endogenous and expressed ICAM-2 surface expression levels by flow cytometry. Left

20 Panel: HUVEC cells stained for endogenous ICAM-2 expression by flow cytometry. E) Dose response curves of ICAM-2 transduced NIH3T3 and Jurkat T-cells to treatment of staurosporine or anti-Fas antibody for 24 hours. Open squares: ICAM-2 expressing cells. Closed squares: control cells expressing vector alone. The proportion of apoptotic cells in each culture without staurosporine or anti-Fas mAb was less than 3%. The percentage of apoptotic cells is expressed

25 as the mean (bar) \pm SD of triplicate cultures.

Figure 20 depicts the results of experiments demonstrating that the overexpression of ICAM-2 results in a broadly acting anti-apoptotic phenotype. A) Effect of ICAM-2 on staurosporine induced apoptosis was assessed by annexin-V binding assay. ICAM-2 expressing NIH3T3 and

30 vector control NIH3T3 were subjected to an annexin-V binding assay in the presence or absence of staurosporine ($1 \mu\text{M}$, 24 hrs). DMSO vehicle served as the negative control. The proportion of apoptotic cells in each culture without staurosporine was 5% \pm 1.2. B) Effects of ICAM-2 overexpression in Jurkat, NIH3T3, 70Z/3, and BaF3 cells in the context of different apoptotic inducers. Infected cells were treated with either staurosporine ($1 \mu\text{M}$), anti-Fas antibody (5

35 $\mu\text{g/ml}$), or deprived of IL-3 for 24 hours as indicated and evaluated for apoptosis as described (Jacobson and Raff, 1995). Cells were infected with retroviral constructs encoding ICAM-2, ICAM2-AN, vector control, or Bcl2 and subjected to apoptosis as indicated (24 hrs, 70Z/3 cells:

0.2 μ M STP, 4hrs). The percentage of apoptotic cells is expressed as the mean (bar) \pm SD of triplicate cultures. C) The α -actinin binding site is required for survival. ICAM-2, wild-type sequence (red line); ICAM2AC, cytoplasmic domain deletion (green line); ICAM-2 C-scrambled, full length with scrambled α -actinin binding site (blue line) were FACS sorted by an
 5 antilCAM-2-FITC monoclonal antibody (IC2/2), and incubated with etoposide (100 μ M, 24hrs). Viable cells were scored after a 24 hr recovery period following etoposide treatment. The percentage of surviving cells is expressed as the mean (bar) \pm SD of triplicate cultures.

Figure 21 depicts the results of experiments demonstrating that PI3-kinase is necessary for
 10 ICAM-2's anti-apoptotic effect. A) Pharmacological inhibitors of PI3K abrogate ICAM-2's anti-apoptotic. ICAM-2 or vector transduced BaF3 cells were treated with staurosporine (1 μ M) with (unfilled) or without (filled) the addition of 100 nM wortmannin for 24 hours (left bar graph). Apoptosis was determined by nuclear staining and fluorescence microscopy (left bar graph) and by annexin-V binding assay (right bar graph). The proportion of apoptotic cells in each culture
 15 without staurosporine was 8% \pm 2. The percentage of apoptotic cells is expressed as the mean (bar) \pm SD of triplicate cultures. ICAM-2, ICAM2-AC, or vector transduced NIH3T3 were treated with staurosporine (1 μ M) in the presence or absence of 10 μ M LY294002 for 24 hours (right bar graph). DMSO vehicle served as negative control. The percentage of apoptotic cells is expressed as the mean (bar) \pm SD of triplicate cultures. B) Confocal images of PI3K (green) and AKT
 20 (red) localization in ICAM-2 (top) or vector control cells (bottom). C) Confocal images of ezrin (red) and the p85 subunit of PI3K (green) localization in ICAM-2 (top) or vector control cells (bottom).

Figure 22 depicts the results of experiments demonstrating that ICAM-2 signal results in
 25 phosphatidylinositol 3,4,5 triphosphate production, PDK-1 and AKT activation, and subsequent phosphorylation of AKT downstream effectors. A) ICAM-2 interacts with α -actinin and ezrin. ICAM-2, ICAM2-AC or control cells were immunoprecipitated for ICAM-2 and immunoblotted for either α -actinin or ezrin. Control lysates were not subjected to immunoprecipitation and reflect positive controls for detected interacting protein. B) ICAM-2 cells exhibit high levels of
 30 phosphatidylinositol 3,4,5 triphosphate. ICAM-2 and control cells were serum starved (48 hrs), treated with LY294002 (10 μ M, 24 hrs), or cultured in growth media, and prepared for intracellular detection of phosphatidylinositol 3,4,5 triphosphate detection by flow cytometry. C) PDK-1 kinase levels are elevated in ICAM-2 cells and are partially inhibited by Y-27632. PDK-1 kinase was immunoprecipitated from cells incubated with the indicated compounds (10
 35 μ M, 1hr) and subjected to an immuno-kinase reaction with inactive SGK enzyme and peptide substrate. Phosphotransfer of [γ 32P]ATP to peptide substrate was detected by scintillation

- counting and presented as counts per minute CPM. D) Immunoblot for AKT in membrane/cytosol fractions of ICAM2 and controls cells and AKT kinase activity. E) AKT kinase level and dual phosphorylation at serine 473 and threonine 308 are sustained in ICAM-2 cells in the presence of staurosporine. F) The AKT kinase substrates BAD and GSK3 remain phosphorylated in
- 5 ICAM-2-expressing cells following staurosporine treatment. Extracts from ICAM-2, ICAM-2AC and control vector cells treated with DMSO-vehicle (-) or 1 μ M staurosporine (+) were assayed for BAD and GSK3 phosphorylation by immunoblotting as indicated.

- Figure 23** depicts the results of experiments demonstrating that ligation of endogenous ICAM-2
- 10 induces tyrosine phosphorylation of ezrin and recruits PI3K to the membrane, resulting in PDK-1 and AKT kinase activation. A) Expression levels of ICAM-2 vary across cell lines of different origins. Quantitative flow cytometry analysis was performed to assess the number of ICAM-2 surface molecules across transformed and untransformed fibroblasts, lymphocytes of T-cell and different B-cell differentiation states and myeloid lineages (see Material and Methods for
- 15 elaboration). The number of surface molecules/cell is expressed as the mean (bar) \pm SD of triplicate experiments. B) Crosslinking of endogenous ICAM-2 induces ezrin tyrosine phosphorylation and enhanced ICAM-2 interaction. Jurkat cells were crosslinked for ICAM-2 using monoclonal antibody for the indicated times and subjected to immunoprecipitation/immunoblotting as indicated. C) Inhibition of ezrin phosphotyrosine by
- 20 chemical inhibitors of src and Rho-dependent kinases. D) Ezrin phosphorylation and enhanced interaction with the p85 subunit of PI3K as a function of ICAM-2. Determined as described above. E) PDK-1 activity as a function of ICAM-2 ligation, as described above. F) PI3K and AKT localize to the membrane as a function of ICAM-2 ligation. Membrane/cytosolic fractionation was performed on ICAM-2 ligated Jurkat cells as a function of time and subjected to immunoblotting
- 25 for PI3K and AKT.

- Figure 24** depicts the results of experiments demonstrating that ligation of endogenous ICAM-2 induced AKT activation in lymphoid cells conferring cell survival. A) ICAM-2 crosslinking induced
- AKT activity in Jurkat T cells and subsequent phosphorylation of GSK3, FKHR, and AFX as a
- 30 function of time. Serum starved Jurkat T cells were incubated with either isotype control or anti-human ICAM-2 (10 μ g/ml) for the indicated times. AKT kinase activity was determined as described. Immunoblot analysis verified AKT dual phosphorylation, FKHR and GSK3 phosphorylation, and BAD phosphorylation as a function of ICAM-2 crosslinking. B) ICAM-2 crosslinking induces AKT activity in BaF3 pre-B cells. C) ICAM-2 clustering on Jurkat T cells
- 35 treated with staurosporine (1 μ M for 6 hours) shows a decrease in apoptosis as measured by flow cytometry annexin-V binding assay. ICAM-2 was crosslinked on 106Jurkat Tcells using an

ICAM-2 mAb (10 µg/ml) 30 minutes prior to treatment with staurosporine (+) or DMSO vehicle (-). Apoptosis was determined by flow cytometry using annexin-V binding assay. D) ICAM-2 survival signal is abrogated in the presence of GTP~S, Psi-tectorigenin, Y-27632 and herbimycin A. Jurkat cells were incubated with indicated inhibitor (10 µM, 1 hour) prior to being crosslinked for ICAM-2 (10 µg/ml, one hour) and treated with anti-Fas (50 ng/ml, 18 hours). Cells were assessed for apoptosis by annexin-V binding assay. E) ICAM-2 crosslinking induces ezrin association with PI3K. ICAM-2, vector control, Jurkat and Jurkat cells crosslinked for ICAM-2 (10 µg/ml, 15 minutes) were subjected to immunoprecipitation and subsequent immunoblotting as indicated.

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Figure 25 depicts the results of experiments demonstrating that ligation of ICAM-2 induces AKT activity and protects primary B cells from apoptosis. A) Human peripheral blood monocytes were crosslinked for ICAM-2 (10 µg/ml 45 minutes) prior to being subjected to apoptosis by treatment with TNFa200 ng/ml plus 1 µg/ml cycloheximide) and anti-Fas antibody (10 ng/ml CH11) for 12 hours. Multi-parameter FACS analysis illustrates ICAM-2 induced AKT activity in CD4+ and CD19+ populations protects from apoptosis. (1x10⁶ events collected to enumerate a minimum of 80,000 CD19+ cells). B) PBMC were crosslinked for ICAM-1, -2, -3, CD43, or CD44 (10 µg/ml 45 minutes) and subjected to phosphatidylinositol detection of phosphatidylinositol 3,4,5 triphosphate, phosphatidylinositol 4,5 bisphosphate, or phospho-AKTser473 detection by flow cytometry. Cell subsets are gated and displayed as indicated. C) PBMC were crosslinked for ICAM-1, -2, -3, CD43, or CD44 (10 µg/ml 45 minutes) prior to being induced to apoptose as described above. Apoptosis was measured by annexin-V binding detected by flow cytometry, and gated for CD19+ cells. D) PBMC was crosslinked for ICAM-2 prior to being induced to apoptose as described above in the presence or absence of inhibitory compounds Psi-tectorigenin and Y-27632 (10 µM, 30 minutes). Apoptosis was measured by annexin-V binding assay. E) LFA-1 overlay assay. LFA-1+ Jurkat cells were overlaid on either vector control or ICAM-2 expressing cells after being treated with LY294002 (2 µM, one hour). Some Jurkat samples were treated with monoclonal antibodies to LFA-1 or Mac-1 (5 µg/ml, 30 minutes) prior to being overlaid on ICAM-2 or vector control. Cells were mixed for 30 minutes at 37°C, and then subjected for flow cytometry. Jurkat cells were gated and removed from displayed analysis of phospho-AktSer473 ICAM-2 cells F) Model for ICAM-2 mediated cell survival signal. ICAM-2 oligomerization recruits and phosphorylates ezrin through a Src/Rho dependent mechanism. This results in recruitment of PI3K to the membrane through ezrin's interaction with the p85 regulatory subunit of PI3K. Membrane translocation of PI3K generates phosphatidylinositol 3,4,5 triphosphates that activate PDK-1 and recruit AKT to the plasma membrane by binding to its PH domain. Translocation of cytosolic AKT to the membrane allows PDK1/2 to phosphorylate AKT at ser473 and thr308, activating AKT. AKT subsequently

phosphorylates downstream effectors GSK3, BAD, FKHR (and potentially others depending on the cell type). Phosphorylation of these effectors can block apoptosis alone or synergistically to generate a survival signal.

5 **Figure 26** depicts the results of experiments demonstrating the retroviral library screen for anti-apoptotic molecules to staurosporine induced apoptosis. A) cDNA library screening scheme. 10^7 NIH3T3 cells were infected with a Jurkat T cell retroviral cDNA library and treated with staurosporine ($1\mu\text{M}$) for 24 hours to induce apoptosis (Stolzenberg et al., 2000). Surviving clones were grown out for one week and subjected to a repeat selection three times. To determine
10 which viral integrants encoded an anti-apoptotic function, a phenotypic transfer assay was performed. An aliquot of the surviving cell population was infected with native Moloney murine leukemia virus (MMLV). This rescued the replication-defective viral vector proviruses, generating infectious particles. NIH3T3 cells were infected with the rescued viral vectors and treated with staurosporine to confirm the anti-apoptotic phenotype. B) Complexity of populations surviving
15 staurosporine induced apoptotic screen. RT-PCR was performed to assess the complexity of the surviving populations. Surviving population (1°) exhibited several discrete bands and the post-transfer population (2°) exhibited a single band. M- molecular weight marker. A negative control vector (LacZJ, initially spiked into the library at 10% was undetectable, while an independent infection of a Bcl2-expressing vector was readily detected.

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Figure 27 depicts the results of experiments demonstrating that overexpression of ICAM-2 inhibits staurosporine induced apoptosis. A) Retroviral vector encoding ICAM-2 western blot analysis in infected NIH3T3. B) Retroviral vector encoding ICAM-2 ectopically expresses surface ICAM-2. Right Panel: BaF3 pro-B-cells infected with a retroviral vector construct encoding
25 ICAM-2 or vector control and assayed for either endogenous and expressed ICAM-2 surface expression levels by flow cytometry. Left Panel: HUVEC cells stained for endogenous ICAM-2 expression by flow cytometry. C) Dose response curves of ICAM-2 transduced NIH3T3 and Jurkat T-cells to treatment of staurosporine or anti-Fas antibody for 24 hours. Open squares: ICAM-2 expressing cells. Closed squares: control cells expressing vector alone. The proportion
30 of apoptotic cells in each culture without staurosporine or anti-Fas mAb was less than 3%. The percentage of apoptotic cells is expressed as the mean (bar) \pm SD of triplicate cultures. D) Pyknotic nuclei of negative control NIH3T3 vector control (-STP), positive control NIH3T3 vector control (+ STP), and ICAM-2 expressing NIH3T3. Nuclei were stained with Hoescht ($5\mu\text{g/ml}$) and acridine orange ($25\mu\text{M}$) and ethidium bromide ($25\mu\text{M}$). Inserts indicate degree of cell death
35 (orange) or cell viability (green). E) Effect of ICAM-2 on staurosporine induced apoptosis was assessed by annexin-V binding assay ICAM-2 expressing NIH3T3 and vector control NIH3T3

were subjected to an annexin-V binding assay in the presence or absence of staurosporine (1 μ M, 24 hr). DMSO vehicle served as the negative control. The proportion of apoptotic cells in each culture without staurosporine was 5% \pm 1.2.

- 5 **Figure 28** depicts the results of experiments demonstrating that ICAM-2 mediates an anti-apoptotic effect. A) Effects of ICAM-2 overexpression in Jurkat, NIH3T3, 70Z/3, and BaF3 cells in the context of different apoptotic inducers. Infected cells were treated with either staurosporine (1 μ M), anti-Fas antibody (5 μ g/ml), or deprived of IL-3 for 24 hours as indicated and evaluated for apoptosis as described (Jacobson and Raff, 1995). Jurkat T cells were
- 10 infected with retroviral constructs encoding ICAM-2 or vector control. Mouse pre-B 70Z/3 cells were infected with a retroviral vector construct encoding ICAM-2 or vector control and treated with staurosporine (0.2 μ M) for 4 hours and evaluated for apoptosis. IL-3-dependent BaF3 cells were infected with retroviral vector constructs encoding ICAM-2, ICAM2- Δ N, or Bcl2. To induce apoptosis, cells were maintained in IL-3-deficient medium for 24 hours. BaF3- cells expressing
- 15 retroviral vector encoded ICAM-2 or an in-frame deletion of the extracellular domain were treated with anti-mouse Fas for 24 hours and evaluated for cell death. The percentage of apoptotic cells is expressed as the mean (bar) \pm SD of triplicate cultures. B) The α -actinin binding site is required for survival. NIH3T3 cells were transduced with ICAM-2 encoding retroviral constructs: ICAM-2, wild-type sequence (red line); ICAM-2 Δ C, cytoplasmic domain deletion (green line);
- 20 ICAM-2 C-scrambled, full length with scrambled α -actinin binding site (blue line). Transduced cells were labeled with a fluorescently conjugated anti-ICAM-2 monoclonal antibody that recognized only the native protein (IC2/2). Cells were then purified by FACS (MoFlo, Cytomation, Ft. Collins, CO). Sorted cells were cultured and then cells were challenged with 100 μ M etoposide for 24 hours. The surviving cells were allowed to recover for 24 hours following
- 25 removal of the drug and the number of viable cells was scored. The percentage of surviving cells is expressed as the mean (bar) \pm SD of triplicate cultures. C) α -actinin and ezrin both co-immunoprecipitate with ICAM-2 and interact with the C-terminus of ICAM-2. ICAM-2 was immunoprecipitated from ICAM-2, ICAM2- Δ C, and vector control transduced NIH3T3. Immunoprecipitates were resolved by SDS-PAGE and subjected to immunoblotting for α -actinin,
- 30 ezrin, and ICAM-2

- Figure 29** depicts the results of experiments demonstrating that pharmacological inhibitors of PI3K abrogate ICAM-2's anti-apoptotic effect. A) The anti-apoptotic effect of ICAM-2 is both wortmannin and LY294002 sensitive. ICAM-2 or vector transduced BaF3 cells were treated with
- 35 staurosporine (1 μ M) with (unfilled) or without (filled) the addition of 100 nM wortmannin for 24 hours. Apoptosis was determined by nuclear staining and fluorescence microscopy (left bar

graph) and by annexin-V binding assay (right bar graph). The proportion of apoptotic cells in each culture without staurosporine was $8\% \pm 2$. The percentage of apoptotic cells is expressed as the mean (bar) \pm SD of triplicate cultures. ICAM-2, ICAM2- Δ C, or vector transduced NIH3T3 were treated with staurosporine (1 μ M) in the presence or absence of 10 μ M LY294002 for 24 hours. DMSO vehicle served as negative control. The percentage of apoptotic cells is expressed as the mean (bar) \pm SD of triplicate cultures. B) ICAM-2 expressing NIH3T3 cells are sensitive to apoptosis in the presence of PI3K inhibitor LY294002. BrdU TUNEL assay was performed in staurosporine (1 μ M) or LY294002 (10 μ M) treated ICAM-2, ICAM2- Δ C, and control vector transduced NIH3T3 cells. The percentage of apoptotic cells is expressed as the mean (bar) \pm SD of triplicate cultures.

Figure 30 depicts the results of experiments demonstrating that ICAM-2 induces AKT kinase activity and phosphorylation of BAD and GSK. A) AKT kinase levels and dual phosphorylation at serine 473 and threonine 308 are elevated in ICAM-2 expressing cells and sustained in the presence of staurosporine. AKT kinase was immunoprecipitated from ICAM-2, ICAM-2 Δ C and control vector (pBMN-Z-IN-GFP) expressing NIH3T3 fibroblasts in the presence or absence of staurosporine (with 1 μ M staurosporine (+) or DMSO-vehicle (-) and incubated with a GSK protein substrate. Phosphorylated GSK was identified by anti-phospho-GSK specific immunoblotting. B) The AKT kinase substrates BAD, FKHR, and GSK3 remain phosphorylated in ICAM-2-expressing cells following staurosporine treatment. Extracts from ICAM-2, ICAM-2 Δ C and control vector expressing NIH3T3 fibroblasts treated with DMS-vehicle (-) or 1 μ M staurosporine (+) were assayed for BAD and GSK3 phosphorylation and by immunoprecipitating BAD or GSK3 and immunoblotting with phospho-BAD, or phospho-GSK3 specific antibodies. C) ICAM-2's anti-apoptotic effect is independent of Bcl-2 or Bcl-X_L as determined by immunoblotting. ICAM-2 or ICAM2- Δ C transduced NIH3T3 were treated with DMSO vehicle (-) or 1 μ M staurosporine (+) for 24 hours and assayed for Bcl2 and Bcl-X_L expression by immunoblotting. D) Expression levels of ICAM-2 vary across cell lines of different origins. Quantitative flow cytometry analysis was performed to assess the number of ICAM-2 surface molecules across transformed and untransformed fibroblasts, lymphocytes of T-cell and different B-cell differentiation states, and myeloid lineages. Geometric mean fluorescence values were calibrated to standard curve derived from geometric mean fluorescence values of beads labeled with known amounts of phycoerythrin molecules (see experimental procedures for elaboration). The number of surface molecules/cell is expressed as the mean (bar) \pm SD of triplicate experiments.

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Figure 31 depicts the results of experiments demonstrating that cross-linking of endogenous ICAM-2 induces AKT activation in BaF3 B-cells and protects Jurkat T cells from apoptosis. A) Cross-linking ICAM-2 with monoclonal anti-ICAM2 antibody induced AKT activity. Serum starved BaF3 preB cells were incubated with either normal rat IgG2 (isotype control) or anti-mouse ICAM-2 at 10 µg/ml for the indicated times. AKT kinase activity was determined as described. immunoblot analysis was performed to verify AKT dual phosphorylation, FKHR phosphorylation, and BAD phosphorylation as a function of ICAM-2 crosslinking. Isotype control did not induce AKT phosphorylation or phosphorylation of FKHR or BAD (data not shown). B) ICAM-2 clustering on Jurkat T cells treated with staurosporine (1 µM for 6 hours) shows a decrease in apoptosis as measured by flow cytometry annexin-V binding assay. ICAM-2 was crosslinked on 10⁶ Jurkat T-cells using a monoclonal anti-ICAM2 antibody (10 µg/ml) 30 minutes prior to treatment with staurosporine (+) or DMSO vehicle (-). Apoptosis was determined by flow cytometry using annexin-V binding assay. (100,000 events collected). C) ICAM-2 crosslinking on Jurkat T cells decreases percentage of death in staurosporine treatment. The percentage of dead cells is expressed as the mean (bar) ± SD of triplicate experiments. D) ICAM-2 cross-linking induces AKT activity in Jurkat T cells and subsequent phosphorylation of GSK3, FKHR, AFX as a function of time. E) ICAM-2 crosslinking can overcome staurosporine induce apoptosis. 10⁶ Jurkat T-cells were treated with staurosporine (1 µM) for one hour as indicated by (+) to induce apoptosis. After one hour of staurosporine induced apoptosis, ICAM-2 was crosslinked using a monoclonal anti-ICAM2 antibody (10 µg/ml) for the indicated times. Attenuation of cleaved caspases was not observed in isotype control (10 µg/ml) (data not shown). F) ICAM-2 cross-linking induces AKT activity, subsequent phosphorylation of BAD, and is sufficient to override apoptotic induction by staurosporine. Jurkat cells were treated as mentioned above. AKT activity was determined by AKT kinase assay and detection of dually phosphorylation AKT by immunoblotting with phospho-specific antibodies against ser473 and thr308. BAD phosphorylation was detected by immunoprecipitation for BAD, followed by immunoblotting with phospho-specific antibodies ser112 and ser136. Isotype control (10 µg/ml) did not generate AKT activation (data not shown).

Figure 32 depicts the results of experiments demonstrating that resistance to TNFα and Fas induced apoptosis in primary human CD4+ and CD19+ cells is mediated by active AKT following ICAM-2 crosslinking. A) Human peripheral blood monocytes were assayed for AKT activity following ICAM-2 clustering using monoclonal antibodies (middle panel) or anti-ICAM-2 labeled antibodies (bottom panel) and subsequently gated for CD4 or CD19 markers (50,000 events collected). B) Human peripheral blood monocytes were crosslinked for ICAM-2 (10 µg/ml 45 minutes) prior to being subjected to apoptosis by treatment with TNFα 200 ng/ml plus 1 µg/ml cycloheximide) and anti-Fas antibody (10 ng/ml CH11) for 12 hours. Multi-parameter FACS

analysis illustrates ICAM-2 induced AKT activity in CD4+ and CD19+ populations protects from apoptosis. (1×10^6 events collected to enumerate a minimum of 80,000 CD19+ cells). C) Model for ICAM-2 mediated anti-apoptotic signaling. ICAM-2 binds intracellular linker proteins α -actinin and ezrin, which recruit PI3K to the plasma membrane. PI3K generates inositol phosphates that

5 recruit AKT to the plasma membrane by binding to its PH domain. Translocation of cytosolic AKT to the membrane allows PDK1/2 to phosphorylate AKT at ser473 and thr308, activating AKT. AKT subsequently phosphorylates downstream effectors GSK3, BAD, FKHR (and potentially others depending on the cell type). Phosphorylation of these effectors can block apoptosis alone or synergistically to generate a survival signal.

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Figure 33 depicts results demonstrating that soluble ICAM-2 binding induces LFA-1 clustering and cytoskeletal polarization. A) Purified human ICAM-2 from retrovirally transduced NIH3T3 (see materials and methods) was tested for purity, size, and aggregate formation by native gel electrophoresis and gel filtration. Analysis of purified ICAM-2 from two different preps and

15 ICAM2-FC from NSO cells. Molecular weight was calculated using relative distance migration from a molecular weight standard. Concentration was determined using serial dilutions of BSA as a standard. Molecular weight and concentration are displayed in the figure. B) ICAM-2 ligand binding to cell surface as a function of time. 5×10^6 unstimulated and PMA stimulated ($1 \mu\text{g/ml}$, 30 min) Jurkat T cells were incubated with ICAM-2-FITC or LFA-1-FITC antibody ($1 \mu\text{g/ml}$) and

20 subjected to time dependent flow cytometry. Mean fluorescence intensity was plotted per time of a gated homogenous cell population. Flow rate was maintained at 200-300 cells/second, with fluorescence intensity values acquired at 10 millisecond intervals for 1200 seconds. C) Flow cytometric analysis of ICAM-2-FITC and α -LFA-1-FITC surface binding on cytochalasin D treated ($10 \mu\text{M}$, 30 min) Jurkat cells at 37°C and 4°C . D) Curve fit analysis of ICAM-2-FITC binding per

25 10^4 cells. Jurkat cells were incubated with ICAM-2-FITC at 37°C for 30 min at indicated concentrations in $50 \mu\text{L}$ volume. Cells were washed 1X, and analyzed for mean fluorescent intensity (MFI) of 10^4 cells. Percent ICAM-2-FITC bound was calculated from the following equation: $100 \times [(MFI_{[x]} - MFI_{\text{unstained}}) / (MFI_{\text{saturated}} - MFI_{\text{unstained}})]$ where $[x] = \text{ICAM-2-FITC}$ concentration and $[\text{saturated}] = \text{concentration that saturated binding}$. Values were fit to the

30 equation $Y = m1 \cdot M0 / (m2 + M0)$ (see materials and methods).

Figure 34 depicts the results of experiments demonstrating the simultaneous detection of ICAM-2 induced LFA-1 clustering, activation, and actin/microtubule reorganization. A) Confocal microscopy of actin and microtubule architecture upon ICAM-2 stimulus ($10 \mu\text{g/ml}$, 30 min) in

35 Jurkat cells. Actin was stained by phalloidin-alexa633 and tubulin by taxol-alexa546. Inserts display enlarged cell image of representative treatment B) Flow cytometric staining for actin and

microtubules as described above. C) Left: Fluorescence topography analysis of ICAM-2-FITC (1 μ g/ml, time as indicated) surface distribution. Intensity color gradient depicts high (red) and low (blue) fluorescent intensity values. Right: Confocal microscopy of ICAM-2-FITC surface binding (10 μ g/ml, 30 min) and anti- β 2-alexa568 (clone CTB104). D) Staining for ICAM-2-FITC surface binding (fluorescence intensity) and surface distribution (fluorescence pulse) in cytochalasin D treated Jurkat cells at 37°C and 4°C by flow cytometry. E) Values for ICAM-2-FITC fluorescence intensity per time-of-flight per cell as described in text. F) Flow cytometric staining for LFA-1 activation by mAb24-alexa633. Stimuli as indicated (10 μ M, 30 min), PMA (1 μ g/ml) prior to ICAM-2 stimulation (10 μ g/ml, 30 min). Staining was performed at 37°C. G) Mean fluorescence intensity (MFI) values of mAb24-633 and ICAM-2 cluster values, computed as described above, as a function of time.

Figure 35 depicts the results of experiments demonstrating ICAM-2 induced p44/42 MAPK activation via LFA-1 interaction. A) Mean fluorescent intensity values (MFI) of ICAM-2-FITC (adhesion) and mAb24-Alexa633 (LFA-1 activation) in treated Jurkat cells (as above). Control unstimulated and/or compound pretreated values were subtracted. B) Top panel: Intracellular phospho-p44/42 MAPK detection as a function of ICAM-2 dose in Jurkat cells by flow cytometry (see Material and Methods). Mean fluorescent intensity values (MFI) were plotted \pm standard deviation (SD). Bottom panel: ICAM-2 treated Jurkat cells were stained for intracellular phospho-p44/42 MAPK as described above. mAbs to β 2 and α_L integrins (10 μ g/ml, 10 minutes) were titrated prior to ICAM-2 treatment as indicated and plotted for MFI \pm SD. C) ICAM-2 induced p44/42 MAPK activity is blocked by β 2 and α_L integrin antibodies as determined by a p44/42 MAPK kinase assay. Conditions for induction and inhibition are as described above. Recombinant active p44/42 MAPK served as an internal positive control (denoted by "+"). D) Inhibition and activation profile for intracellular phospho-p44/42 by flow cytometry in the presences of chemical agents (10 μ M), EDTA (1 mM), or PMA, ionomycin and ICAM-2 stimulus (as indicated above). MFI values \pm SD are plotted.

Figure 36 depicts the results of experiments demonstrating ICAM-2 induced phosphorylation of Pyk2 and Syk, and β 2 integrin association. A) Phospho-raf and phospho-p44/42 immunoblot inhibition profile by tyrosine kinase inhibitors. 1×10^5 cells were treated with indicated compound (10 μ M, 30 min) and then stimulated with ICAM-2 (10 μ g/ml, 30 min). Cell lysates were immunoblotted for phospho-raf and phospho-p44/42. Compound alone did not induce detectable phosphorylation. B) Pyk2 and Syk are phosphorylated and co-immunoprecipitate with β 2 integrin upon ICAM-2 stimulus. Phospho-specificity was determined by phospho-PykY402 and phospho-syk(Tyr525/526) antibodies. C) Kinetic analyses of the phosphorylation state of

PKC α/β , Pyk2, and Syk as a function of ICAM-2 stimulus per time. Cells were treated and processed as above. Phospho-specific PKC α/β _{II}(Thr638) and the following antibodies were used; Pyk2 and Syk were first immunoprecipitated, probed with anti-phosphotyrosine antibody (PY20), stripped and subsequently probed with indicated non-phospho specific antibody.

5 Immunoblots are representative of triplicate experiments.

Figure 37 depicts the results of experiments demonstrating that ICAM-2 induces cytotoxic lymphocyte activity in IL-2 activated human PBMC. A) PBMC were either treated with ICAM-2 (10 μ g/ml) in the presence of IL-2 for 12 hr and then incubated with CFSE labeled target HL60 cells at a 50:1 E:T ratio for 4 hrs. Remaining HL60 cells were quantified by flow cytometry. B) PBMC were treated with IL-2 (100 U/ml, 12 hrs) and treated with ICAM-1, -2, or 3 FC proteins (10 μ g/ml) and used in a cytotoxicity assay as described above. Results are representative of 4 independent experiments.

Figure 38 depicts the results of experiments demonstrating ICAM-2 exhibits differences to ICAM-1 and ICAM-3 in mediating perforin and granzyme release from CD56⁺CD8⁺ cytotoxic lymphocytes subsets. IL-2 activated PBMC were mock-treated (IgG), ICAM-1, ICAM-2, or ICAM-3 treated (10 μ g/ml of FC fusion protein) for 12 hrs prior to incubation with target HL60 cells at a 50:1 E:T ratio for 4 hrs. Cells were then prepared for flow cytometry with CD8-CY5PE, CD56-PE surface stains, and perforin-CY5 and granzyme-A-FITC intracellular stains. Cells were gated for CD56⁺CD8^{low}, CD56⁺CD8^{med}, CD56⁺CD8^{high}, CD56⁺CD8⁺, CD56⁺CD8^{high} populations as shown in panel I and population frequencies within appropriate gate. Panels II-V are subset gated populations displayed for perforin and granzyme-A log fluorescent intensities. Results are representative of 3 independent experiments and were similar at 25:1 and 12.5:1 E:T ratios (data not shown). Manual calibration was performed. B) CD56CD8 population subsets were gated (as indicated) and displayed for intracellular perforin for ICAM-1, -2, -3 stimulated cells. Perforin percentage was calculated from the following equation where MFI equals mean fluorescent intensity: $100 \times [(MFI_{\text{experimental}} - MFI_{\text{isotype mAb}}) / (MFI_{\text{control}} - MFI_{\text{isotype mAb}})]$. Unstimulated cells were used as control. C) Intracellular granzyme-A values were calculated and displayed as described above.

Figure 39 depicts the results of experiments demonstrating that ICAM-2 induced LFA-1 mediated p44/42 MAPK correlates with LFA-1 activation in human CD56⁺CD8⁺ cells. A) Conjugate formation of CFSE labeled HL60 cells and CD56⁺CD8⁺ cells. Conjugate flow cytometric based assay was performed on PBMC treated with indicated chemicals (10 μ M, 30 min) prior to

treatment with ICAM-2 (10 µg/ml, 30 min). CFSE labeled HL60 cells were incubated at 25:1 E:T ratio for 5 min, and fixed with 1% paraformaldehyde. Cells were then immunolabeled with CD8 and CD56 antibodies, gated for CD8⁺CD56⁺ cell populations and percent HL60 fluorescence was made relative to total HL60 cells. B) IL-2 activated PBMC were either treated with ICAM-2 (10 µg/ml, 30 min) or mock-treated (IgG) and stained for active-LFA-1 (mAb24-Alexa633), phospho-p44/42-Ax488, CD8-CY5PE, and CD56-PE. CD56⁺CD8⁺ cell populations are gated and displayed for active LFA-1 vs. phospho-p44/42. The mean fluorescent intensities (MFI) of mAb24-Ax633 and phospho-44/42-Ax488 were computed and displayed over time as described above.

10

Detailed Description of the Invention

Intracellular assays of signaling systems have been limited by an inability to correlate functional subsets of cells in complex populations based on the activity of signaling agents, for example, the activity of kinases. Such correlations are important for distinguishing changes in signaling status that arise in rare cell subsets during signaling or in disease manifestations. The present invention solves these problems by providing methods and compositions for simultaneously detecting the activation state of a plurality of activatable proteins in single cells using flow cytometry. The invention further provides methods and compositions of screening for bioactive agents capable of coordinately modulating the activity or activation state of a plurality of activatable proteins in single cells. The methods and compositions can be used to determine the protein activation profile of a cell for predicting or diagnosing a disease state, and for monitoring treatment of a disease state. Further, the methods and compositions of the present invention can be used optionally to sequentially detect the activation state of a plurality of activatable proteins in single cells. In addition, the methods and compositions of the present invention can be used optionally to detect the activation state of a single protein or modulate the activity or activation state of a single protein in single cells.

The methods and compositions of the present invention may be used to detect any particular protein isoform in a sample that is antigenically detectable and antigenically distinguishable from other isoforms of the protein which are present in the sample. For example, as demonstrated (see, e.g., the Examples) and described herein, the activation state-specific antibodies of the present invention can be used in the present methods to identify distinct signaling cascades of a subset or subpopulation of complex cell populations; and the ordering of protein activation (e.g., kinase activation) in potential signaling hierarchies. Further, in the methods of the present invention, the use of flow cytometry, particularly polychromatic flow cytometry, permits the multi-dimensional analysis and functional assessment of the signaling pathway in single cells.

As used herein, the term "activation state-specific antibody" or "activation state antibody" or grammatical equivalents thereof, refer to an antibody that specifically binds to a corresponding and specific antigen. Preferably, the corresponding and specific antigen is a specific isoform of an activable protein. Also preferably, the binding of the activation state-specific antibody is
5 indicative of a specific activation state of a specific activatable protein. Thus, in preferred embodiments, the binding of an activation state-specific antibody to a corresponding isoform of an activable protein is indicative of the identity of the activatable protein and of the activation state of the activatable protein.

10 In a preferred embodiment, the activation state-specific antibody is a peptide comprising a recognition structure that binds to a target structure on an activatable protein. A variety of recognition structures are well known in the art and can be made using methods known in the art, including by phage display libraries (see e.g., Gururaja et al. Chem. Biol. (2000) 7:515-27; Houimel et al., Eur. J. Immunol. (2001) 31:3535-45; Cochran et al. J. Am. Chem. Soc. (2001)
15 123:625-32; Houimel et al. Int. J. Cancer (2001) 92:748-55, each incorporated herein by reference). In a preferred embodiment, the activation state-specific antibody comprises the following recognition structure: SKVILFE—random peptide loop— SKVILFE. Antibodies having such recognition structures can bind with high affinity to specific target structures. Further, fluorophores can be attached to such antibodies for use in the methods of the present invention.

20

A variety of recognitions structures are known in the art (e.g., Cochran et al., J. Am. Chem. Soc. (2001) 123:625-32; Boer et al., Blood (2002) 100:467-73, each expressly incorporated herein by reference)) and can be produced using methods known in the art (see e.g., Boer et al., Blood (2002) 100:467-73; Gualillo et al., Mol. Cell Endocrinol. (2002) 190:83-9, each expressly
25 incorporated herein by reference)), including for example combi chem methods for producing recognition structures such as polymers with affinity for a target structure on an activable protein (see e.g., Barn et al., J. Comb. Chem. (2001) 3:534-41; Ju et al., Biotechnol. (1999) 64:232-9, each expressly incorporated herein by reference). In another preferred embodiment, the activation state-specific antibody is a protein that only binds to an isoform of a specific activatable
30 protein that is phosphorylated and does not bind to the isoform of this activable protein when it is not phosphorylated or nonphosphorylated. In another preferred embodiment the activation state-specific antibody is a protein that only binds to an isoform of an activatable protein that is intracellular and not extracellular, or vice versa.

In a preferred embodiment, the recognition structure is an anti-laminin single-chain antibody fragment (scFv) (see e.g., Sanz et al., Gene Therapy (2002) 9:1049-53; Tse et al., J. Mol. Biol. (2002) 317:85-94, each expressly incorporated herein by reference).

5 As used herein, the terms "polypeptide" and "protein" may be used interchangeably and mean at least two covalently attached amino acids, which includes proteins, polypeptides, oligopeptides and peptides. The protein may be made up of naturally occurring amino acids and peptide bonds, or synthetic peptidomimetic structures. Thus "amino acid", or "peptide residue", as used herein means both naturally occurring and synthetic amino acids. For example, homo-
10 phenylalanine, citrulline and noreleucine are considered amino acids for the purposes of the invention. "Amino acid" also includes imino acid residues such as proline and hydroxyproline. The side chains may be in either the (R) or the (S) configuration. In the preferred embodiment, the amino acids are in the (S) or L-configuration. If non-naturally occurring side chains are used, non-amino acid substituents may be used, for example to prevent or retard *in vivo* degradation.

15

As used herein, an "activatable protein" or "substrate" or "substrate protein" or "protein substrate" grammatical equivalents thereof, refers to a protein that has at least one isoform (and in some cases two or more isoforms) that corresponds to a specific form of the protein having a particular biological, biochemical, or physical property, e.g., an enzymatic activity, a modification (e.g.,
20 post-translational modification), or a conformation. The activatable protein can be activated or nonactivated with respect to a particular biological activity, modification, or conformation. Specifically, the "activated" or "active" form of the activatable protein has the particular biological activity, modification, or conformation, whereas the "non-activated" or "non-active" form of the activatable protein does not have (or has a lesser or diminished level of) the particular biological
25 activity, modification, or conformation, respectively. In some embodiments, there may be more than one isoform associated with activity or activation state; for example, there may be an isoform associated with an "open" conformation available for substrate binding, a second "transition state" isoform, and an isoform devoid of activity (e.g., where the activity is inhibited). Examples of activatable proteins include, but are not limited to, phospho-proteins and phospho-
30 lipids. Further examples of activatable proteins include, but are not limited to, kinases, phosphatases, PIP2, PIP3, and proteases such as cysteine and serine proteases, including, but not limited to caspases, cathepsins and a variety of well known serine proteases.

In a preferred embodiment, the biological, biochemical, or physical property (e.g. enzymatic
35 activity, modification, or conformation) of the activatable protein is inducible or "activatable" by an

activating agent or by cell signaling events. Examples of activating agents include, but are not limited to, kinases, phosphatases, proteases (e.g., caspases), and hormones. Examples of cell signaling events include, but are not limited to, receptor clustering or binding of a cognate molecule or ligand.

5

As used herein, an "isoform" or grammatical equivalents thereof, refers to a form of an activatable protein having a specific, and preferably detectable, biological activity, modification, or conformation. The isoform can be an activated (or active) form, or non-activated (or not active) form of an activatable protein. As mentioned, in preferred embodiments, the binding of an
10 activation state-specific antibody to a corresponding isoform of an activable protein is indicative of the identity of the activatable protein and of the activation state of the activatable protein. In a preferred embodiment, the invention provides methods for determining a protein isoform profile which comprise determining the presence of an isoform of an activatable protein that is activated (or activated isoform).

15

In a preferred embodiment, the activated isoform or activated state of an activable protein is a form of the activable protein having a particular or specific biological, biochemical, or physical property that is not possessed by at least one other isoform of activatable protein. Examples of such properties include, but are not limited to, enzymatic activity (e.g., kinase activity and
20 protease activity), and protein binding activity. Thus, such particular or specific properties or activities are associated with an activated isoform of an activatable protein. Such properties or activities are sometimes referred to herein as activation state activities.

An example of activation state activity is kinase activity for an activated protein kinase. As used
25 herein, a protein kinase may refer to a protein that when activated is capable of catalyzing the phosphorylation of amino acids, or derivatives thereof, which possess an hydroxyl group. Preferred kinases are those which are capable of catalyzing the phosphorylation of serine, threonine, and tyrosine residues. Kinase activity may be determined by supplying a substrate for phosphorylation by kinase, a source of phosphate usable by kinase, and determining the
30 phosphorylation of substrate in the presence of kinase.

Another example of activation state activity is protease activity for an activated protease. As used herein, a protease may refer to a protein that when activated is capable of hydrolyzing a peptide bond within a polypeptide comprising an amino acid sequence. Preferred proteases are
35 the family of cysteine proteases known as caspases. Caspase activity may be determined by

providing a substrate of a caspase and determining the cleavage of substrate in the presence of caspase. Other cysteine proteases having activations states that may be monitored using the methods of the invention include, but are not limited to, cathepsins. Similarly, an additional class of proteases that may be monitored using the methods of the invention include serine proteases,
5 of which a wide variety of suitable classes are known.

The antigenicity of an activated isoform of an activatable protein is distinguishable from the antigenicity of non-activated isoform of an activatable protein or from the antigenicity of an isoform of a different activation state. In a preferred embodiment, an activated isoform of a
10 protein possesses an epitope that is absent in a non-activated isoform of a protein, or vice versa. In another preferred embodiment, this difference is due to covalent addition of moieties to a protein, such as phosphate moieties, or due to a structural change in a protein, as through protein cleavage, or due to an otherwise induced conformational change in a protein which causes the protein to present the same sequence in an antigenically distinguishable way. In
15 another preferred embodiment, such a conformational change causes an activated isoform of a protein to present at least one epitope that is not present in a non-activated isoform, or to not present at least one epitope that is presented by a non-activated isoform of the protein. In some embodiments, the epitopes for the distinguishing antibodies are centered around the active site of the enzyme, although as is known in the art, conformational changes in one area of a protein
20 may cause alterations in different areas of the protein as well.

Among proteins that are capable of existing in a non-activated isoform and an activated isoform, which isoforms are antigenically distinguishable, are kinases. Kinases are enzymes that catalyze protein phosphorylation on serine, threonine, or tyrosine residues. The majority of kinases are
25 phosphoproteins that are reversibly phosphorylated as a form of activity regulation. In the phosphorylated form (phosphorylated isoform), protein kinases are typically enzymatically active kinases. In the un-phosphorylated form, protein kinases are typically enzymatically inactive.

Many antibodies, many of which are commercially available (for example, see Cell Signaling
30 Technology, www.cellsignal.com, the contents which are incorporated herein by reference). have been produced which specifically bind to the phosphorylated isoform of a protein but do not specifically bind to a non-phosphorylated isoform of a protein. Many such antibodies have been produced for the study of signal transducing proteins which are reversibly phosphorylated. Particularly, many such antibodies have been produced which specifically bind to
35 phosphorylated, activated isoforms of protein kinases and are sometimes referred to herein as

kinase activation state antibodies or grammatical equivalents thereof. Particularly preferred antibodies for use in the present invention include: phospho-AKT Ser473 monoclonal anti-4E2, phospho-p44/42 MAP kinase (Thr202/Tyr204) monoclonal antibody, phospho-TYK2 (Tyr1054/1055) antibody, phospho-p38 MAP kinase (Thr180/Tyr182) monoclonal antibody
5 28B10, phospho-PKC-PAN substrate antibody, phospho-PKA-substrate, phospho-SAPK/JNK (Thr183/Tyr185) G9 monoclonal antibody, phospho-tyrosine monoclonal antibody (P-tyr-100), p44/42 MAPK, p38 MAPK, JNK/SAPK, and phospho-AKT-Thr308.

The present invention provides methods for the determination of a kinase activation state profile
10 for a sample which comprise simultaneously determining the presence of activated isoforms of a multiplicity of kinases using a multiplicity of antibodies that specifically bind to active, phosphorylated isoforms of the multiplicity of kinases.

Another example of proteins that are capable of existing in a non-activated isoform and an
15 activated isoform, which isoforms are antigenically distinguishable, are caspases. Caspases are proteases that catalyze the hydrolysis of peptide bonds within proteins. Caspases are synthesized as pro-caspases which are proteolytically inactive precursors of caspases. A pro-caspase is specifically cleaved by protease to yield an active caspase. As is known in the art, this is known to be true for a variety of other proteases, including both cysteine and serine
20 proteases such as cathepsins, tissue plasminogen activators, etc., that are known to exist as pro-proteases, as is well known in the art.

The present invention provides methods for the determination of a caspase activation state profile for a sample which comprise simultaneously determining the presence of activated
25 isoforms of a multiplicity of caspases using a multiplicity of antibodies that specifically bind to a multiplicity of caspases but not to the correlating pro-caspases from which they may be derived. For example, antibodies which specifically bind to caspases but do not specifically bind to pro-caspases from which caspases are derived are useful in the methods of the invention. Many such antibodies are commercially available (for example, see Cell Signaling Technology,
30 www.cellsignal.com, the contents of which are incorporated herein by reference).

Additional means for determining kinase activation are provided by the present invention. Substrates that are specifically recognized by protein kinases and phosphorylated thereby are known. Antibodies that specifically bind to such phosphorylated substrates but do not bind to
35 such non-phosphorylated substrates (phospho-substrate antibodies) may be used to determine

the presence of activated kinase in a sample.

In a preferred embodiment, the present invention provides methods for determining a kinase activation state profile for a sample, comprising providing a population of cells comprising a plurality of activatable proteins and contacting the cells with at least one non-phosphorylated protein kinase substrate under conditions which provide for phosphorylation of the substrate by activated kinase if present, and determining the presence of phosphorylated protein kinase substrate using a phospho-substrate antibody that binds specifically to the phosphorylated substrate. In a preferred embodiment, the present invention provides methods for determining a kinase activation state profile for a sample which comprises the use of a multiplicity of protein kinase substrates, a multiplicity of phospho-substrate antibodies, and a multiplicity of activation state antibodies to simultaneously determine the activation state of a multiplicity of kinases.

In another embodiment, the present invention provides methods for determining the activation state of one or more protein kinases and one or more caspases in a sample by combining the methods described for determining the activation state profile of each provided herein. Preferably, the sample comprises a population of cells, a cell, cell lysate, or proteins.

Similarly, the invention provides methods utilizing any and all other combinations of kinases and proteases (e.g., kinases and caspases, kinases and cathepsins, kinases and serine proteases, caspases and cathepsins, etc.).

In a further embodiment, a protein activation state profile is determined using a multiplicity of activation state antibodies that are immobilized. Antibodies may be non-diffusibly bound to an insoluble support having isolated sample receiving areas (e.g. a microtiter plate, an array, etc.). The insoluble supports may be made of any composition to which the compositions can be bound, is readily separated from soluble material, and is otherwise compatible with the overall method of screening. The surface of such supports may be solid or porous and of any convenient shape. Examples of suitable insoluble supports include microtiter plates, arrays, membranes, and beads. These are typically made of glass, plastic (e.g., polystyrene), polysaccharides, nylon or nitrocellulose, teflon™, etc. Microtiter plates and arrays are especially convenient because a large number of assays can be carried out simultaneously, using small amounts of reagents and samples. In some cases magnetic beads and the like are included.

- The particular manner of binding of the composition is not crucial so long as it is compatible with the reagents and overall methods of the invention, maintains the activity of the composition and is nondiffusable. Preferred methods of binding include the use of antibodies (which do not sterically block either the ligand binding site or activation sequence when the protein is bound to the support), direct binding to "sticky" or ionic supports, chemical crosslinking, the synthesis of the antibody on the surface, etc. Following binding of the antibody, excess unbound material is removed by washing. The sample receiving areas may then be blocked through incubation with bovine serum albumin (BSA), casein or other innocuous protein or other moiety.
- 5
- 10 In a preferred embodiment, an epitope-recognizing fragment of an activation state antibody rather than the whole antibody is used. In another preferred embodiment, the epitope-recognizing fragment is immobilized. In another preferred embodiment, the antibody light chain which recognizes an epitope is used. A recombinant nucleic acid encoding a light chain gene product which recognizes an epitope may be used to produce such an antibody fragment by recombinant
- 15 means well known in the art.

- A chip analogous to a DNA chip can be used in the methods of the present invention. Arrayers and methods for spotting nucleic acid to a chip in a prefigured array are known. In addition, protein chips and methods for synthesis are known. These methods and materials may be adapted for the purpose of affixing activation state antibodies to a chip in a prefigured array.
- 20

- In a preferred embodiment, such a chip comprises a multiplicity of kinase activation state antibodies, and is used to determine a kinase activation state profile for a sample. In a preferred embodiment, such a sample is a cell extract. In such a method, detection of activated kinase is by "sandwich assay" as known in the art. Briefly, a sample, preferably a cell extract, is passed over a the chip under conditions which allow the multiplicity of immobilized kinase activation state antibodies to simultaneously bind to a multiplicity of activated kinases if present in the sample. The immobilized antibody-kinase complex is optionally washed and contacted with a second plurality of antibodies comprising non-activation state antibodies that are capable of specifically binding to activated kinases while the kinases are specifically bound to kinase activation state specific antibodies. Such non-activation state specific antibodies specifically bind to activated kinases via an epitope that is not recognized by the kinase activation state specific antibody. Binding of the non-activation state specific antibodies to the activation state antibody-activated kinase complex is determined and reveals the presence of activated kinase in sample. As will be appreciated, the determination of binding of second antibody in the sandwich assay can be
- 25
- 30
- 35

accomplished in many different ways. Preferably, the multiplicity of non-activation state specific antibodies are uniquely labeled to facilitate detection.

5 In an alternative embodiment, a chip comprises a multiplicity of non-activation state specific antibodies. Such a chip is contacted with sample, preferably cell extract, and a second multiplicity of antibodies comprising kinase activation state specific antibodies is used in the sandwich assay to simultaneously determine the presence of a multiplicity of activated kinases in sample. Preferably, the multiplicity of activation state specific antibodies are uniquely labeled to facilitate detection.

10

In a preferred embodiment, one or more components of the methods of the present invention comprise a tag. By "tag" is meant an attached molecule or molecules useful for the identification or isolation of the attached molecule(s), which are preferably substrate molecules. For example, a tag can be an attachment tag or a label tag. Components having a tag are referred to as "tag-
15 X", wherein X is the component. Preferably, the tag is covalently bound to the attached component. When more than one component of a combination has a tag, the tags will be numbered for identification, for example, "tag1-antibody", "tag1-protein", and "tag1-substrate". Components may comprise more than one tag, in which case each tag will be numbered, for example "tag 1,2-antibody", "tag1,2-protein", and "tag1,2-substrate. Preferred tags include, but
20 are not limited to, a label, a partner of a binding pair, and a surface substrate binding molecule (or attachment tag). As will be evident to the skilled artisan, many molecules may find use as more than one type of tag, depending upon how the tag is used.

By "label" is meant a molecule that can be directly (*i.e.*, a primary label) or indirectly (*i.e.*, a
25 secondary label) detected; for example a label can be visualized and/or measured or otherwise identified so that its presence or absence can be known. A compound can be directly or indirectly conjugated to a label which provides a detectable signal, e.g. radioisotope, fluorescers, enzyme, antibodies, particles such as magnetic particles, chemiluminescers, or specific binding molecules, etc. Specific binding molecules include pairs, such as biotin and streptavidin, digoxin
30 and antidigoxin etc. Preferred labels include, but are not limited to, fluorescent labels, label enzymes and radioisotopes.

By "fluorescent label" is meant any molecule that may be detected via its inherent fluorescent properties. Suitable fluorescent labels include, but are not limited to, fluorescein, rhodamine,
35 tetramethylrhodamine, eosin, erythrosin, coumarin, methyl-coumarins, pyrene, Malacite green,

- stilbene, Lucifer Yellow, Cascade Blue™, Texas Red, IAEDANS, EDANS, BODIPY FL, LC Red 640, Cy 5, Cy 5.5, LC Red 705 and Oregon green. Suitable optical dyes are described in the 1996 Molecular Probes Handbook by Richard P. Haugland, hereby expressly incorporated by reference. Suitable fluorescent labels also include, but are not limited to, green fluorescent protein (GFP; Chalfie, *et al.*, *Science* 263(5148):802-805 (Feb 11, 1994); and EGFP; Clontech - Genbank Accession Number U55762), blue fluorescent protein (BFP; 1. Quantum Biotechnologies, Inc. 1801 de Maisonneuve Blvd. West, 8th Floor, Montreal (Quebec) Canada H3H 1J9; 2. Stauber, R. H. *Biotechniques* 24(3):462-471 (1998); 3. Heim, R. and Tsien, R. Y. *Curr. Biol.* 6:178-182 (1996)), enhanced yellow fluorescent protein (EYFP; 1. Clontech Laboratories, Inc., 1020 East Meadow Circle, Palo Alto, CA 94303), luciferase (Ichiki, *et al.*, *J. Immunol.* 150(12):5408-5417 (1993)), -galactosidase (Nolan, *et al.*, *Proc Natl Acad Sci USA* 85(8):2603-2607 (Apr 1988)) and Renilla WO 92/15673; WO 95/07463; WO 98/14605; WO 98/26277; WO 99/49019; U.S. patent 5,292,658; U.S. patent 5,418,155; U.S. patent 5,683,888; U.S. patent 5,741,668; U.S. patent 5,777,079; U.S. patent 5,804,387; U.S. patent 5,874,304; U.S. patent 5,876,995; and U.S. patent 5,925,558). All of the above-cited references are expressly incorporated herein by reference.

- Particularly, preferred labels for use in the present invention include: Alexa-Fluor dyes (Alexa Fluor 350, Alexa Fluor 430, Alexa Fluor 488, Alexa Fluor 546, Alexa Fluor 568, Alexa Fluor 594, Alexa Fluor 633, Alexa Fluor 660, Alexa Fluor 680), Cascade Blue, Cascade Yellow and R-phycoerythrin (PE) (Molecular Probes) (Eugene, Oregon), FITC, Rhodamine, and Texas Red (Pierce, Rockford, Illinois), Cy5, Cy5.5, Cy7 (Amersham Life Science, Pittsburgh, Pennsylvania). Tandem conjugate protocols for Cy5PE, Cy5.5PE, Cy7PE, Cy5.5APC, Cy7APC can be found at www.dmr.com/abcon. Quantitation of fluorescent probe conjugation may be assessed to determine degree of labeling and protocols including dye spectral properties can be found at www.metazoa.com/UPL3419.

In another preferred embodiment, the fluorescent label is a GFP and, more preferably, a *renilla*, *ptilosarcus*, or *aequorea* species of GFP.

In preferred embodiments, multiple fluorescent labels are employed in the methods and compositions of the present invention. In a preferred embodiment, at least two fluorescent labels are used which are members of a fluorescence resonance energy transfer (FRET) pair.

FRET is phenomenon known in the art wherein excitation of one fluorescent dye is transferred to another without emission of a photon. A FRET pair consists of a donor fluorophore and an acceptor fluorophore. The fluorescence emission spectrum of the donor and the fluorescence absorption spectrum of the acceptor must overlap, and the two molecules must be in close proximity. The distance between donor and acceptor at which 50% of donors are deactivated (transfer energy to the acceptor) is defined by the Förster radius (R_0), which is typically 10-100 Å. Changes in the fluorescence emission spectrum comprising FRET pairs can be detected, indicating changes in the number of that are in close proximity (*i.e.*, within 100 Å of each other). This will typically result from the binding or dissociation of two molecules, one of which is labeled with a FRET donor and the other of which is labeled with a FRET acceptor, wherein such binding brings the FRET pair in close proximity. Binding of such molecules will result in an increased fluorescence emission of the acceptor and/or quenching of the fluorescence emission of the donor.

FRET pairs (donor/acceptor) useful in the invention include, but are not limited to, EDANS/fluorescein, IAEDANS/fluorescein, fluorescein/tetramethylrhodamine, fluorescein/LC Red 640, fluorescein/Cy 5, fluorescein/Cy 5.5 and fluorescein/LC Red 705.

In another aspect of FRET, a fluorescent donor molecule and a nonfluorescent acceptor molecule ("quencher") may be employed. In this application, fluorescent emission of the donor will increase when quencher is displaced from close proximity to the donor and fluorescent emission will decrease when the quencher is brought into close proximity to the donor. Useful quenchers include, but are not limited to, TAMRA, DABCYL, QSY 7 and QSY 33. Useful fluorescent donor/quencher pairs include, but are not limited to EDANS/DABCYL, Texas Red/DABCYL, BODIPY/DABCYL, Lucifer yellow/DABCYL, coumarin/DABCYL and fluorescein/QSY 7 dye.

The skilled artisan will appreciate that FRET and fluorescence quenching allow for monitoring of binding of labeled molecules over time, providing continuous information regarding the time course of binding reactions.

By "label enzyme" is meant an enzyme which may be reacted in the presence of a label enzyme substrate which produces a detectable product. Suitable label enzymes for use in the present invention include but are not limited to, horseradish peroxidase, alkaline phosphatase and glucose oxidase. Methods for the use of such substrates are well known in the art. The

presence of the label enzyme is generally revealed through the enzyme's catalysis of a reaction with a label enzyme substrate, producing an identifiable product. Such products may be opaque, such as the reaction of horseradish peroxidase with tetramethyl benzedine, and may have a variety of colors. Other label enzyme substrates, such as Luminol (available from Pierce
5 Chemical Co.), have been developed that produce fluorescent reaction products. Methods for identifying label enzymes with label enzyme substrates are well known in the art and many commercial kits are available. Examples and methods for the use of various label enzymes are described in Savage *et al.*, *Previews* 247:6-9 (1998), Young, *J. Virol. Methods* 24:227-236 (1989), which are each hereby incorporated by reference in their entirety.

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By "radioisotope" is meant any radioactive molecule. Suitable radioisotopes for use in the invention include, but are not limited to ^{14}C , ^3H , ^{32}P , ^{33}P , ^{35}S , ^{125}I , and ^{131}I . The use of radioisotopes as labels is well known in the art.

15 As mentioned, labels may be indirectly detected, that is, the tag is a partner of a binding pair. By "partner of a binding pair" is meant one of a first and a second moiety, wherein the first and the second moiety have a specific binding affinity for each other. Suitable binding pairs for use in the invention include, but are not limited to, antigens/antibodies (for example, digoxigenin/anti-digoxigenin, dinitrophenyl (DNP)/anti-DNP, dansyl-X-anti-dansyl, Fluorescein/anti-fluorescein,
20 lucifer yellow/anti-lucifer yellow, and rhodamine anti-rhodamine), biotin/avid (or biotin/streptavidin) and calmodulin binding protein (CBP)/calmodulin. Other suitable binding pairs include polypeptides such as the FLAG-peptide [Hopp *et al.*, *BioTechnology*, 6:1204-1210 (1988)]; the KT3 epitope peptide [Martin *et al.*, *Science*, 255:192-194 (1992)]; tubulin epitope peptide [Skinner *et al.*, *J. Biol. Chem.*, 266:15163-15166 (1991)]; and the T7 gene 10 protein
25 peptide tag [Lutz-Freyermuth *et al.*, *Proc. Natl. Acad. Sci. USA*, 87:6393-6397 (1990)] and the antibodies each thereto. As will be appreciated by those in the art, binding pair partners may be used in applications other than for labeling, as is described herein.

As will be appreciated by those in the art, a partner of one binding pair may also be a partner of
30 another binding pair. For example, an antigen (first moiety) may bind to a first antibody (second moiety) which may, in turn, be an antigen for a second antibody (third moiety). It will be further appreciated that such a circumstance allows indirect binding of a first moiety and a third moiety via an intermediary second moiety that is a binding pair partner to each.

35 As will be appreciated by those in the art, a partner of a binding pair may comprise a label, as

described above. It will further be appreciated that this allows for a tag to be indirectly labeled upon the binding of a binding partner comprising a label. Attaching a label to a tag which is a partner of a binding pair, as just described, is referred to herein as "indirect labeling".

- 5 By "surface substrate binding molecule" or "attachment tag" and grammatical equivalents thereof is meant a molecule have binding affinity for a specific surface substrate, which substrate is generally a member of a binding pair applied, incorporated or otherwise attached to a surface. Suitable surface substrate binding molecules and their surface substrates include, but are not limited to poly-histidine (poly-his) or poly-histidine-glycine (poly-his-gly) tags and Nickel
- 10 substrate; the Glutathione-S Transferase tag and its antibody substrate (available from Pierce Chemical); the flu HA tag polypeptide and its antibody 12CA5 substrate [Field *et al.*, Mol. Cell. Biol., 8:2159-2165 (1988)]; the c-myc tag and the 8F9, 3C7, 6E10, G4, B7 and 9E10 antibody substrates thereto [Evan *et al.*, Molecular and Cellular Biology, 5:3610-3616 (1985)]; and the
- 15 Herpes Simplex virus glycoprotein D (gD) tag and its antibody substrate [Paborsky *et al.*, Protein Engineering, 3(6):547-553 (1990)]. In general, surface binding substrate molecules useful in the present invention include, but are not limited to, polyhistidine structures (His-tags) that bind to nickel substrates, antigens that bind to surface substrates comprising antibody, haptens that bind to avidin substrate (e.g., biotin) and CBP that binds to surface substrate comprising calmodulin.
- 20 Production of antibody-embedded substrates is well known; see Slinkin *et al.*, Bioconj. Chem. 2:342-348 (1991); Torchilin *et al.*, *supra*; Trubetskoy *et al.*, Bioconj. Chem. 3:323-327 (1992); King *et al.*, Cancer Res. 54:6176-6185 (1994); and Wilbur *et al.*, Bioconjugate Chem. 5:220-235 (1994) (all of which are hereby expressly incorporated by reference), and attachment of or production of proteins with antigens is described above. Calmodulin-embedded substrates are
- 25 commercially available, and production of proteins with CBP is described in Simcox *et al.*, Strategies 8:40-43 (1995), which is hereby incorporated by reference in its entirety.

As will be appreciated by those in the art, tag-components of the invention can be made in various ways, depending largely upon the form of the tag. Components of the invention and tags

30 are preferably attached by a covalent bond.

The production of tag-polypeptides by recombinant means when the tag is also a polypeptide is described below. Production of tag-labeled proteins is well known in the art and kits for such production are commercially available (for example, from Kodak and Sigma). Examples of tag

35 labeled proteins include, but are not limited to, a Flag-polypeptide and His-polypeptide. Methods

for the production and use of tag-labeled proteins are found, for example, in Winston *et al.*, *Genes and Devel.* 13:270-283 (1999), incorporated herein in its entirety, as well as product handbooks provided with the above-mentioned kits.

- 5 Biotinylation of target molecules and substrates is well known, for example, a large number of biotinylation agents are known, including amine-reactive and thiol-reactive agents, for the biotinylation of proteins, nucleic acids, carbohydrates, carboxylic acids; see chapter 4, Molecular Probes Catalog, Haugland, 6th Ed. 1996, hereby incorporated by reference. A biotinylated substrate can be attached to a biotinylated component via avidin or streptavidin. Similarly, a
10 large number of haptenylation reagents are also known (Id.).

Methods for labeling of proteins with radioisotopes are known in the art. For example, such methods are found in Ohta *et al.*, *Molec. Cell* 3:535-541 (1999), which is hereby incorporated by reference in its entirety.

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Production of proteins having tags by recombinant means is well known, and kits for producing such proteins are commercially available. For example, such a kit and its use is described in the QIAexpress Handbook from Qiagen by Joanne Crowe *et al.*, hereby expressly incorporated by reference.

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- The functionalization of labels with chemically reactive groups such as thiols, amines, carboxyls, etc. is generally known in the art. In a preferred embodiment, the tag is functionalized to facilitate covalent attachment. The covalent attachment of the tag may be either direct or via a linker. In one embodiment, the linker is a relatively short coupling moiety, that is used to attach the
25 molecules. A coupling moiety may be synthesized directly onto a component of the invention and contains at least one functional group to facilitate attachment of the tag. Alternatively, the coupling moiety may have at least two functional groups, which are used to attach a functionalized component to a functionalized tag, for example. In an additional embodiment, the linker is a polymer. In this embodiment, covalent attachment is accomplished either directly, or
30 through the use of coupling moieties from the component or tag to the polymer. In a preferred embodiment, the covalent attachment is direct, that is, no linker is used. In this embodiment, the component preferably contains a functional group such as a carboxylic acid which is used for direct attachment to the functionalized tag. It should be understood that the component and tag
35 the tag is attached to the amino or carboxyl terminus of the polypeptide. As will be appreciated by

those in the art, the above description of the covalent attachment of a label applies to the attachment of virtually any two molecules of the present disclosure.

In a preferred embodiment, the tag is functionalized to facilitate covalent attachment, as is generally outlined above. Thus, a wide variety of tags are commercially available which contain functional groups, including, but not limited to, isothiocyanate groups, amino groups, haloacetyl groups, maleimides, succinimidyl esters, and sulfonyl halides, all of which may be used to covalently attach the tag to a second molecule, as is described herein. The choice of the functional group of the tag will depend on the site of attachment to either a linker, as outlined above or a component of the invention. Thus, for example, for direct linkage to a carboxylic acid group of a protein, amino modified or hydrazine modified tags will be used for coupling via carbodiimide chemistry, for example using 1-ethyl-3-(3-dimethylaminopropyl)-carbodiimide (EDAC) as is known in the art (see Set 9 and Set 11 of the Molecular Probes Catalog, *supra*; see also the Pierce 1994 Catalog and Handbook, pages T-155 to T-200, both of which are hereby incorporated by reference). In one embodiment, the carbodiimide is first attached to the tag, such as is commercially available for many of the tags described herein.

Antibody conjugation may be performed using standard procedures (<http://drmr.com.abcon>) or by using protein-protein/protein-dye crosslinking kits from Molecular Probes (Eugene, Oregon).

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Using the example of two activation state specific antibodies, by "uniquely labeled" is meant that a first activation state antibody recognizing a first activated kinase comprises a first label, and second activation state antibody recognizing a second activated kinase comprises a second label, wherein the first and second label are detectable and distinguishable, making the first antibody and the second antibody uniquely labeled.

Non-activation state antibodies may also be used in the present invention. In a preferred embodiment, non-activation state antibodies bind to epitopes in both activated and non-activated forms of a protein. Such antibodies may be used to determine the amount of non-activated plus activated protein in a sample. In another preferred embodiment, non-activation state antibodies bind to epitopes present in non-activated forms of a protein but absent in activated forms of a protein. Such antibodies may be used to determine the amount of non-activated protein in a sample. Both types of non-activation state antibodies may be used to determine if a change in the amount of activation state protein, for example from samples before and after treatment with a candidate bioactive agent as described herein, coincide with changes in the amount of non-

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activation state protein. For example, such antibodies can be used to determine whether an increase in activated protein is due to activation of non-activation state protein, or due to increased expression of protein, or both.

- 5 In another preferred embodiment, antibodies are immobilized using beads analogous to those known and used for standardization in flow cytometry. Attachment of a multiplicity of activation state specific antibodies to beads may be done by methods known in the art and/or described herein. Such conjugated beads may be contacted with sample, preferably cell extract, under conditions which allow for a multiplicity of activated kinases, if present, to bind to the multiplicity
10 of immobilized antibodies. A second multiplicity of antibodies comprising non-activation state antibodies which are uniquely labeled may be added to the immobilized activation state specific antibody-activated kinase complex and the beads may be sorted by FACS on the basis of the presence of each label, wherein the presence of label indicates binding of corresponding second antibody and the presence of corresponding activated kinase.

15

In a preferred embodiment, the present invention provides methods for determining a protein activation state profile for a single cell. The methods comprise sorting cells by FACS on the basis of the activation state of at least two proteins. Activation state-specific antibodies are used to sort cells on the basis of protein activation state.

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- When using fluorescent labeled components in the methods and compositions of the present invention, it will be recognized that different types of fluorescent monitoring systems, e.g., FACS systems, can be used to practice the invention. Preferably, FACS systems are used or systems dedicated to high throughput screening, e.g. 96 well or greater microtiter plates. Methods of
25 performing assays on fluorescent materials are well known in the art and are described in, e.g., Lakowicz, J. R., Principles of Fluorescence Spectroscopy, New York: Plenum Press (1983); Herman, B., Resonance energy transfer microscopy, in: Fluorescence Microscopy of Living Cells in Culture, Part B, Methods in Cell Biology, vol. 30, ed. Taylor, D. L. & Wang, Y.-L., San Diego: Academic Press (1989), pp. 219-243; Turro, N.J., Modern Molecular Photochemistry, Menlo
30 Park: Benjamin/Cummings Publishing Co., Inc. (1978), pp. 296-361.

- Fluorescence in a sample can be measured using a fluorimeter. In general, excitation radiation, from an excitation source having a first wavelength, passes through excitation optics. The excitation optics cause the excitation radiation to excite the sample. In response, fluorescent
35 proteins in the sample emit radiation which has a wavelength that is different from the excitation

wavelength. Collection optics then collect the emission from the sample. The device can include a temperature controller to maintain the sample at a specific temperature while it is being scanned. According to one embodiment, a multi-axis translation stage moves a microtiter plate holding a plurality of samples in order to position different wells to be exposed. The multi-axis
5 translation stage, temperature controller, auto-focusing feature, and electronics associated with imaging and data collection can be managed by an appropriately programmed digital computer. The computer also can transform the data collected during the assay into another format for presentation.

10 In a preferred embodiment, flow cytometry is used to detect fluorescence. Other methods of detecting fluorescence may also be used, e.g., Quantum dot methods (see, e.g., Goldman et al., J. Am. Chem. Soc. (2002) 124:6378-82; Pathak et al. J. Am. Chem. Soc. (2001) 123:4103-4; and Remacle et al., Proc. Natl. Sci. USA (2000) 18:553-8, each expressly incorporated herein by reference).

15 In a preferred embodiment, FRET is used as a way of monitoring the activation state of proteins inside a cell. The degree of FRET can be determined by any spectral or fluorescence lifetime characteristic of the excited construct, for example, by determining the intensity of the fluorescent signal from the donor, the intensity of fluorescent signal from the acceptor, the ratio of the
20 fluorescence amplitudes near the acceptor's emission maxima to the fluorescence amplitudes near the donor's emission maximum, or the excited state lifetime of the donor. For example, cleavage of the linker can increase the intensity of fluorescence from the donor, decreases the intensity of fluorescence from the acceptor, decreases the ratio of fluorescence amplitudes from the acceptor to that from the donor, and increases the excited state lifetime of the donor.

25 Preferably, changes in the degree of FRET are determined as a function of the change in the ratio of the amount of fluorescence from the donor and acceptor moieties, a process referred to as "ratioing." Changes in the absolute amount of substrate, excitation intensity, and turbidity or other background absorbances in the sample at the excitation wavelength affect the intensities of
30 fluorescence from both the donor and acceptor approximately in parallel. Therefore the ratio of the two emission intensities is a more robust and preferred measure of cleavage than either intensity alone.

The ratio-metric fluorescent reporter system described herein has significant advantages over
35 existing reporters for protein integration analysis, as it allows sensitive detection and isolation of both expressing and non-expressing single living cells. In a preferred embodiment, the assay

system uses a non-toxic, non-polar fluorescent substrate which is easily loaded and then trapped intracellularly. Modification of the fluorescent substrate by a cognate protein yields a fluorescent emission shift as substrate is converted to product. Because the reporter readout is ratiometric it is unique among reporter protein assays in that it controls for variables such as the amount of
5 substrate loaded into individual cells. The stable, easily detected, intracellular readout eliminates the need for establishing clonal cell lines prior to expression analysis. This system and other analogous flow sorting systems can be used to isolate cells having a particular protein activation profile from pools of millions of viable cells.

- 10 The detecting, sorting, or isolating step of the methods of the present invention can entail fluorescence-activated cell sorting (FACS) techniques, where FACS is used to select cells from the population containing a particular surface marker, or the selection step can entail the use of magnetically responsive particles as retrievable supports for target cell capture and/or
background removal. A variety FACS systems are known in the art and can be used in the
15 methods of the invention (see e.g., WO99/54494, filed April 16, 1999; U.S.A.N. 20010006787, filed July 5, 2001, each expressly incorporated herein by reference).

In a preferred embodiment, a FACS cell sorter (e.g. a FACSVantage.TM. Cell Sorter, Becton Dickinson Immunocytometry Systems, San Jose, Calif.) is used to sort and collect cells based on
20 their protein activation profile (positive cells). The cells are first contacted with fluorescent-labeled activation state-specific antibodies directed against specific isoforms of specific activatable proteins. In one embodiment, the amount of bound antibody on each cell can be measured by passing droplets containing the cells through the cell sorter. By imparting an electromagnetic charge to droplets containing the positive cells, the cells can be separated from
25 other cells. The positively selected cells can then be harvested in sterile collection vessels. These cell sorting procedures are described in detail, for example, in the FACSVantage.TM. Training Manual, with particular reference to sections 3-11 to 3-28 and 10-1 to 10-17.

In another embodiment, positive cells can be sorted using magnetic separation of cells based on
30 the presence of an isoform of an activatable protein. In such separation techniques, cells to be positively selected are first contacted with specific binding agent (e.g., an antibody or reagent that binds an isoform of an activatable protein). The cells are then contacted with retrievable particles (e.g., magnetically responsive particles) which are coupled with a reagent that binds the specific binding agent (that has bound to the positive cells). The cell-binding agent-particle
35 complex can then be physically separated from non-positive or non-labeled cells, for example, using a magnetic field. When using magnetically responsive particles, the positive or labeled cells can be retained in a container using a magnetic field while the negative cells are removed.

These and similar separation procedures are described, for example, in the Baxter Immunotherapy Isolex training manual.

In a preferred embodiment, methods for the determination of a kinase activation state profile for a single cell are provided. The methods comprise providing a population of cells and sorting the population of cells by FACS. Preferably, cells are separated on the basis of the activation state of at least two kinases. In a preferred embodiment, a multiplicity of kinase activation state antibodies (sometimes referred to herein as kinase activation state specific antibodies) are used to simultaneously determine the activation state of a multiplicity of kinases. Particularly preferred is the use of at least two, preferably more than two, preferably all of the following kinase activation state antibodies: anti-AKT-phospho-Ser473, anti-AKT phospho-Thr308, anti-p44/42 MAPK phospho-Thr202/Tyr204, anti-TYK2 phospho-Tyr1054/1055, anti-p38 MAPK phospho-Thr180/Tyr182, anti-JNK/SAPK phospho-Thr183/Tyr185, anti-phospho-tyrosine, anti-phospho-threonine.

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In addition to the direct detection of phosphorylated kinases in single cells, the methods provide for the detection of activated kinases in single cells using specific substrates for kinases and phospho-substrate antibodies as described herein. In a preferred embodiment, cells are contacted with at least two such substrates, contacted with phospho-substrate antibodies, and then sorted on the basis of kinase activation using a FACS machine. Particularly preferred are phospho-substrate antibodies that specifically bind to PKC substrate PAN phospho-substrate (which is a substrate for α , β I, β II, and δ isoforms of PKC only when phosphorylated at a carboxy-terminal residue homologous to Ser660 of PKC β II), and phospho-substrate antibodies that specifically bind to PKA phospho-substrate (consensus phosphorylation site of PKA phosphorylation of threonine with arginine at -3 position).

25

In another preferred embodiment, a combination of kinase activation state antibodies that specifically bind to phosphorylated kinases, specific substrates for kinases, and phospho-substrate antibodies are used to determine a kinase activation state profile for a single cell. The methods comprise sorting cells on the basis of the activation state of at least two kinases using a FACS machine. Activation of at least two kinases is determined using at least one kinase activation state antibody and at least one specific substrate for a kinase and at least one phospho-substrate antibody for the measurement of the at least one phosphorylated substrate. Preferably, a kinase activation state antibody is selected from the group consisting of anti-AKT-phospho-Ser473, anti-AKT phospho-Thr308, anti-p44/42 MAPK phospho-Thr202/Tyr204, anti-TYK2 phospho-Tyr1054/1055, anti-p38 MAPK phospho-Thr180/Tyr182, anti-JNK/SAPK

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phospho-Thr183/Tyr185, anti-phospho-tyrosine, anti-phospho-threonine. Preferably, a phospho-substrate antibody is selected from an antibody that specifically binds to PKC substrate PAN phospho-substrate (which is a substrate for α , β I, β II, and δ isoforms of PKC only when phosphorylated at a carboxy terminal residue homologous to Ser660 of PKC β II), and an antibody
5 that specifically binds to PKA phospho-substrate (consensus phosphorylation site of PKA phosphorylation of threonine with arginine at -3 position).

In a preferred embodiment, cell sorting by FACS on the basis of the activation state of at least two proteins, preferably kinases and/or caspases, is combined with a determination of other
10 FACS readable outputs, such as the presence of surface markers, granularity and cell size to provide a correlation between the activation state of a multiplicity of proteins and other cell qualities measurable by FACS for single cells.

As will be appreciated, the present invention also provides for the ordering of protein activation
15 events in signal transduction. Particularly, the present invention allows the artisan to construct a kinase activation hierarchy based on the correlation of levels of activation of a multiplicity of kinases within single cells.

The present invention may also be used to determine the presence of cellular subsets, based on
20 correlated protein activation, preferably kinase activation, within complex cellular mixtures such as peripheral blood mononuclear cells. These subsets may represent different differentiation or activation states or different cell lineages or sublineages.

It will also be recognized that a homogeneous cell population is desirable for studying signal
25 transduction in order that variances in signaling between cells not qualitatively and quantitatively mask signal transduction events. The ultimate homogeneous system is the single cell. The present invention provides methods for the analysis of signal transduction in single cells, where the activated state of the signal transducing proteins involved is antigenically distinguishable from a non-activated state.

30

As will be appreciated, these methods provide for the identification of distinct signaling cascades for both artificial and stimulatory conditions in complex cell populations, such a peripheral blood mononuclear cells, or naive and memory lymphocytes.

The methods provided herein may also involve the use of specific inhibitors of particular kinases. The methods provided herein may also involve the use of specific inhibitors of caspases. The methods provided herein may also involve the use of other pharmacological inhibitors of signaling pathways. These inhibitors may be used as controls to ensure that antibodies

- 5 specifically bind to activated isoforms of proteins. For example, an inhibitor of a kinase known to phosphorylate and activate a second kinase may be used to inhibit phosphorylation of the second kinase and examine whether an antibody specifically recognizes a phosphorylated isoform of the second kinase. Alternatively, the inhibitors may be used to further probe signaling pathways and correlations in protein activity, particularly in single cells.

10

In a preferred embodiment, methods for the determination of a caspase activation state profile for a single cell are provided. The methods comprise providing a population of cells and sorting the population of cells by FACS. Preferably, cells are separated on the basis of the activation state of at least one caspase. In a preferred embodiment, caspase cleavage product antibodies as

- 15 described herein are used to determine the activation state of at least one caspase.

In a preferred embodiment, a method for screening for a bioactive agent capable of modulating kinase activity is provided which comprises contacting a cell with a candidate bioactive agent and determining kinase activation in said cell by cell sorting said cell by FACS.

20

In a preferred embodiment, the method comprises contacting a plurality of cells with a plurality of candidate bioactive agents and sorting the cells by FACS on the basis of the activation of at least one kinase.

- 25 In a preferred embodiment, a method for screening for a bioactive agent capable of modulating caspase activity is provided which comprises contacting a cell with a candidate bioactive agent and determining caspase activation in said cell by cell sorting said cell by FACS.

- 30 In a preferred embodiment, the method comprises contacting a plurality of cells with a plurality of candidate bioactive agents and sorting the cells by FACS on the basis of the activation of at least one caspase.

By "candidate bioactive agent", "candidate agent", "candidate modulator", "candidate modulating

agent", or "exogeneous compound" or grammatical equivalents herein is meant any molecule, e.g., protein, small organic molecule, carbohydrates (including polysaccharides), polynucleotide, lipids, etc. Generally a plurality of assay mixtures can be run in parallel with different agent concentrations to obtain a differential response to the various concentrations. Typically, one of these concentrations can serve as a negative control, *i.e.*, at zero concentration or below the level of detection. In addition, positive controls can be used.

Candidate agents encompass numerous chemical classes. In a preferred embodiment, the candidate agents are small molecules. In another preferred embodiment, the candidate agents are organic molecules, particularly small organic molecules, comprising functional groups necessary for structural interaction with proteins, particularly hydrogen bonding, and typically include at least an amine, carbonyl, hydroxyl or carboxyl group, preferably at least two of the functional chemical groups. The candidate agents often comprise cyclical carbon or heterocyclic structures and/or aromatic or polyaromatic structures substituted with one or more chemical functional groups.

Candidate agents are obtained from a wide variety of sources, as will be appreciated by those in the art, including libraries of synthetic or natural compounds. As will be appreciated by those in the art, the present invention provides a rapid and easy method for screening any library of candidate modulators, including the wide variety of known combinatorial chemistry-type libraries.

In a preferred embodiment, candidate agents are synthetic compounds. Any number of techniques are available for the random and directed synthesis of a wide variety of organic compounds and biomolecules, including expression of randomized oligonucleotides. See for example WO 94/24314, hereby expressly incorporated by reference, which discusses methods for generating new compounds, including random chemistry methods as well as enzymatic methods. As described in WO 94/24314, one of the advantages of the present method is that it is not necessary to characterize the candidate agent prior to the assay. Using the methods of the present invention, any candidate agents can be screened for the ability to modulate (e.g., increase or decrease) the activity of an activatable protein. In addition, as is known in the art, coding tags using split synthesis reactions may be used to essentially identify the chemical moieties tested.

Alternatively, a preferred embodiment utilizes libraries of natural compounds, as candidate agents, in the form of bacterial, fungal, plant and animal extracts that are available or readily

produced.

Additionally, natural or synthetically produced libraries and compounds are readily modified through conventional chemical, physical and biochemical means. Known pharmacological
5 agents may be subjected to directed or random chemical modifications, including enzymatic modifications, to produce structural analogs.

In a preferred embodiment, candidate agents include proteins, nucleic acids, and chemical moieties.

10

In a preferred embodiment, the candidate agents are proteins, as defined above. In a preferred embodiment, the candidate agents are naturally occurring proteins or fragments of naturally occurring proteins. Thus, for example, cellular extracts containing proteins, or random or directed digests of proteinaceous cellular extracts, may be tested, as is more fully described
15 below. In this way libraries of prokaryotic and eukaryotic proteins may be made for screening against any number of candidate agents. Particularly preferred in this embodiment are libraries of bacterial, fungal, viral, and mammalian proteins, with the latter being preferred, and human proteins being especially preferred.

20 In a preferred embodiment, the candidate agents are peptides of from about 2 to about 50 amino acids, with from about 5 to about 30 amino acids being preferred, and from about 8 to about 20 being particularly preferred. The peptides may be digests of naturally occurring proteins as is outlined above, random peptides, or "biased" random peptides. By "randomized" or grammatical equivalents herein is meant that each nucleic acid and peptide consists of essentially random
25 nucleotides and amino acids, respectively. Since generally these random peptides (or nucleic acids, discussed below) are chemically synthesized, they may incorporate any nucleotide or amino acid at any position. The synthetic process can be designed to generate randomized proteins or nucleic acids, to allow the formation of all or most of the possible combinations over the length of the sequence, thus forming a library of randomized candidate bioactive
30 proteinaceous agents.

The library should provide a sufficiently structurally diverse population of randomized agents to effect a probabilistically sufficient range of diversity to allow interaction with a particular activatable protein. Accordingly, an interaction library must be large enough so that at least one

- of its members will have a structure that interacts with an activatable protein or other specific components of the signal transduction pathway involving the activatable protein. Although it is difficult to gauge the required absolute size of an interaction library, nature provides a hint with the immune response: a diversity of 10^7 - 10^8 different antibodies provides at least one
- 5 combination with sufficient affinity to interact with most potential antigens faced by an organism. Published *in vitro* selection techniques have also shown that a library size of 10^7 to 10^8 is sufficient to find structures with affinity for a target. A library of all combinations of a peptide 7 to 20 amino acids in length, such as generally proposed herein, has the potential to code for 20^7 (10^5) to 20^{20} . Thus, with libraries of 10^7 to 10^8 different molecules the present methods allow a
- 10 "working" subset of a theoretically complete interaction library for 7 amino acids, and a subset of shapes for the 20^{20} library. Thus, in a preferred embodiment, at least 10^6 , preferably at least 10^7 , more preferably at least 10^8 and most preferably at least 10^9 different sequences are simultaneously analyzed in the subject methods. Preferred methods maximize library size and diversity.
- 15
- In one embodiment, the library is fully randomized, with no sequence preferences or constants at any position. In a preferred embodiment, the library is biased. That is, some positions within the sequence are either held constant, or are selected from a limited number of possibilities. For example, in a preferred embodiment, the nucleotides or amino acid residues are randomized
- 20 within a defined class, for example, of hydrophobic amino acids, hydrophilic residues, sterically biased (either small or large) residues, towards the creation of cysteines, for cross-linking, prolines for SH-3 domains, serines, threonines, tyrosines or histidines for phosphorylation sites, etc., or to purines, etc.
- 25 In a preferred embodiment, the bias is towards peptides or nucleic acids that interact with known classes of molecules. For example, when the candidate agent is a peptide, it is known that much of intracellular signaling is carried out via short regions of polypeptides interacting with other polypeptides through small peptide domains. For instance, a short region from the HIV-1 envelope cytoplasmic domain has been previously shown to block the action of cellular
- 30 calmodulin. Regions of the Fas cytoplasmic domain, which shows homology to the mastoparan toxin from Wasps, can be limited to a short peptide region with death-inducing apoptotic or G protein inducing functions. Magainin, a natural peptide derived from *Xenopus*, can have potent anti-tumor and anti-microbial activity. Short peptide fragments of a protein kinase C isozyme (β PKC), have been shown to block nuclear translocation of β PKC in *Xenopus* oocytes following
- 35 stimulation. And, short SH-3 target peptides have been used as pseudosubstrates for specific binding to SH-3 proteins. This is of course a short list of available peptides with biological

activity, as the literature is dense in this area. Thus, there is much precedent for the potential of small peptides to have activity on intracellular signaling cascades. In addition, agonists and antagonists of any number of molecules may be used as the basis of biased randomization of candidate modulators as well.

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Thus, a number of molecules or protein domains are suitable as starting points for the generation of biased randomized candidate modulators. A large number of small molecule domains are known, that confer a common function, structure or affinity. In addition, as is appreciated in the art, areas of weak amino acid homology may have strong structural homology. A number of these molecules, domains, and/or corresponding consensus sequences, are known, including, but are not limited to, SH-2 domains, SH-3 domains, Pleckstrin, death domains, protease cleavage/recognition sites, enzyme inhibitors, enzyme substrates, and Traf.

In a preferred embodiment, the candidate modulating agent is a polypeptide. In another preferred embodiment, the polypeptide is a cyclic peptide having at least 4 to 20 amino acids. Also in another preferred embodiment, the polypeptide is a catalytically inactive polypeptide. Examples of catalytically inactive polypeptides include, but are not limited to, catalytically inactive activable proteins and, more specifically a catalytically inactive kinases (e.g., PI3K) or caspases. In a further aspect, the candidate modulating agent is peptide fragment of an activatable protein, wherein the peptide fragment comprises an amino acid sequence that is a subsequence of the full-length amino acid sequence of the activable protein.

In a preferred embodiment, the candidate agents are nucleic acids. With reference to candidate agents, by "nucleic acid" or "oligonucleotide" or grammatical equivalents herein means at least two nucleotides covalently linked together. A nucleic acid of the present invention will generally contain phosphodiester bonds, although in some cases, as outlined below, nucleic acid analogs are included that may have alternate backbones, comprising, for example, phosphoramidate (Beaucage *et al.*, Tetrahedron 49(10):1925 (1993) and references therein; Letsinger, J. Org. Chem. 35:3800 (1970); Sprinzl *et al.*, Eur. J. Biochem. 81:579 (1977); Letsinger *et al.*, Nucl. Acids Res. 14:3487 (1986); Sawai *et al.*, Chem. Lett. 805 (1984), Letsinger *et al.*, J. Am. Chem. Soc. 110:4470 (1988); and Pauwels *et al.*, Chemica Scripta 26:141 (1986)), phosphorothioate (Mag *et al.*, Nucleic Acids Res. 19:1437 (1991); and U.S. Patent No. 5,644,048), phosphorodithioate (Briu *et al.*, J. Am. Chem. Soc. 111:2321 (1989), O-methylphosphoroamidite linkages (see Eckstein, Oligonucleotides and Analogues: A Practical Approach, Oxford University Press), and peptide nucleic acid backbones and linkages (see Egholm, J. Am. Chem. Soc.

114:1895 (1992); Meier *et al.*, Chem. Int. Ed. Engl. 31:1008 (1992); Nielsen, Nature, 365:566 (1993); Carlsson *et al.*, Nature 380:207 (1996), all of which are incorporated by reference). Other analog nucleic acids include those with positive backbones (Denpcy *et al.*, Proc. Natl. Acad. Sci. USA 92:6097 (1995); non-ionic backbones (U.S. Patent Nos. 5,386,023, 5,637,684, 5,602,240, 5,216,141 and 4,469,863; Kiedrowshi *et al.*, Angew. Chem. Intl. Ed. English 30:423 (1991); Letsinger *et al.*, J. Am. Chem. Soc. 110:4470 (1988); Letsinger *et al.*, Nucleoside & Nucleotide 13:1597 (1994); Chapters 2 and 3, ASC Symposium Series 580, "Carbohydrate Modifications in Antisense Research", Ed. Y.S. Sanghui and P. Dan Cook; Mesmaeker *et al.*, Bioorganic & Medicinal Chem. Lett. 4:395 (1994); Jeffs *et al.*, J. Biomolecular NMR 34:17 (1994); Tetrahedron Lett. 37:743 (1996)) and non-ribose backbones, including those described in U.S. Patent Nos. 5,235,033 and 5,034,506, and Chapters 6 and 7, ASC Symposium Series 580, "Carbohydrate Modifications in Antisense Research", Ed. Y.S. Sanghui and P. Dan Cook. Nucleic acids containing one or more carbocyclic sugars are also included within the definition of nucleic acids (see Jenkins *et al.*, Chem. Soc. Rev. (1995) pp169-176). Several nucleic acid analogs are described in Rawls, C & E News June 2, 1997 page 35. All of these references are hereby expressly incorporated by reference. These modifications of the ribose-phosphate backbone may be done to facilitate the addition of additional moieties such as labels, or to increase the stability and half-life of such molecules in physiological environments.

As will be appreciated by those in the art, all of these nucleic acid analogs may find use in the present invention. In addition, mixtures of naturally occurring nucleic acids and analogs can be made. Alternatively, mixtures of different nucleic acid analogs, and mixtures of naturally occurring nucleic acids and analogs may be made. Particularly preferred are peptide nucleic acids (PNA) which includes peptide nucleic acid analogs. These backbones are substantially non-ionic under neutral conditions, in contrast to the highly charged phosphodiester backbone of naturally occurring nucleic acids.

The nucleic acids may be single stranded or double stranded, as specified, or contain portions of both double stranded or single stranded sequence. The nucleic acid may be DNA, both genomic and cDNA, RNA or a hybrid, where the nucleic acid contains any combination of deoxyribo- and ribo-nucleotides, and any combination of bases, including uracil, adenine, thymine, cytosine, guanine, inosine, xanthine hypoxanthine, isocytosine, isoguanine, etc. As used herein, the term "nucleoside" includes nucleotides and nucleoside and nucleotide analogs, and modified nucleosides such as amino modified nucleosides. In addition, "nucleoside" includes non-naturally occurring analog structures. Thus for example the individual units of a peptide nucleic acid, each containing a base, are referred to herein as a nucleoside.

- As described above generally for proteins, nucleic acid candidate agent may be naturally occurring nucleic acids, random nucleic acids, or "biased" random nucleic acids. For example, digests of prokaryotic or eukaryotic genomes may be used as is outlined above for proteins. Where the ultimate expression product is a nucleic acid, at least 10, preferably at least 12, more preferably at least 15, most preferably at least 21 nucleotide positions need to be randomized, with more preferable if the randomization is less than perfect. Similarly, at least 5, preferably at least 6, more preferably at least 7 amino acid positions need to be randomized; again, more are preferable if the randomization is less than perfect.
- 5
- 10 In a preferred embodiment, the candidate modulating agent is a mutant cDNA encoding a catalytically inactive polypeptide. Examples of such catalytically inactive polypeptides include, but are not limited to, catalytically inactive activatable proteins and, more specifically, catalytically inactive kinases (e.g., PI3K) or caspases.
- 15 In a preferred embodiment, the candidate modulating agent is an RNA, for example an antisense RNA or siRNA (small inhibitory RNA). In another preferred embodiment, the siRNA cleaves RNA encoding an activatable protein. The siRNAs can be prepared using the methods described herein and known in the art.
- 20 In a preferred embodiment, the candidate agents are organic moieties. In this embodiment, as is generally described in WO 94/24314, candidate agents are synthesized from a series of substrates that can be chemically modified. "Chemically modified" herein includes traditional chemical reactions as well as enzymatic reactions. These substrates generally include, but are not limited to, alkyl groups (including alkanes, alkenes, alkynes and heteroalkyl), aryl groups
- 25 (including arenes and heteroaryl), alcohols, ethers, amines, aldehydes, ketones, acids, esters, amides, cyclic compounds, heterocyclic compounds (including purines, pyrimidines, benzodiazepines, beta-lactams, tetracyclines, cephalosporins, and carbohydrates), steroids (including estrogens, androgens, cortisone, ecodysone, etc.), alkaloids (including ergots, vinca, curare, pyrrolizidine, and mitomycines), organometallic compounds, hetero-atom bearing
- 30 compounds, amino acids, and nucleosides. Chemical (including enzymatic) reactions may be done on the moieties to form new substrates or candidate agents which can then be tested using the present invention.

- As will be appreciated by those in the art, it is possible to screen more than one type of candidate agent at a time, e.g., by combining the candidate agents in the methods of the present invention.
- 35

Thus, the library of candidate agents used may include only one type of agent (*i.e.* peptides), or multiple types (peptides and organic agents).

5 By "combining" is meant the combining of the various components in a reaction mixture in vitro or in a cell in vivo under conditions which promote an activity that is detectable using known methods or using the methods of the present invention (e.g., the binding of an antibody to a corresponding antigen or isoform of an activatable protein, or activation state of an activatable protein).

10 It is understood by the skilled artisan that the steps of the assays provided herein can vary in order. It is also understood, however, that while various options (of compounds, properties selected or order of steps) are provided herein, the options are also each provided individually, and can each be individually segregated from the other options provided herein. Moreover, steps which are obvious and known in the art that will increase the sensitivity of the assay are intended
15 to be within the scope of this invention. For example, there may be additionally washing steps, blocking steps, etc.

In a preferred embodiment, the reaction mixture or cells are contained in a well of a 96 well plate or other commercially available multiwell plate. In an alternate preferred embodiment, the
20 reaction mixture or cells are in a FACS machine. Other multiwell plates useful in the present invention include, but are not limited to 384 well plates and 1536 well plates. Still other vessels for containing the reaction mixture or cells and useful in the present invention will be apparent to the skilled artisan.

25 The addition of the components of the assay for detecting the activation state or activity of an activatable protein, or modulation of such activation state or activity, may be sequential or in a predetermined order or grouping under conditions appropriate for the activity that is assayed for. Such conditions are described here and known in the art. Moreover, further guidance is provided below (see, e.g., in the Examples).

30

In a preferred embodiment, the methods of the invention include the use of liquid handling components. The liquid handling systems can include robotic systems comprising any number of components. In addition, any or all of the steps outlined herein may be automated; thus, for example, the systems may be completely or partially automated.

As will be appreciated by those in the art, there are a wide variety of components which can be used, including, but not limited to, one or more robotic arms; plate handlers for the positioning of microplates; automated lid or cap handlers to remove and replace lids for wells on non-cross contamination plates; tip assemblies for sample distribution with disposable tips; washable tip assemblies for sample distribution; 96 well loading blocks; cooled reagent racks; microtiter plate pipette positions (optionally cooled); stacking towers for plates and tips; and computer systems.

Fully robotic or microfluidic systems include automated liquid-, particle-, cell- and organism-handling including high throughput pipetting to perform all steps of screening applications. This includes liquid, particle, cell, and organism manipulations such as aspiration, dispensing, mixing, diluting, washing, accurate volumetric transfers; retrieving, and discarding of pipet tips; and repetitive pipetting of identical volumes for multiple deliveries from a single sample aspiration. These manipulations are cross-contamination-free liquid, particle, cell, and organism transfers. This instrument performs automated replication of microplate samples to filters, membranes, and/or daughter plates, high-density transfers, full-plate serial dilutions, and high capacity operation.

In a preferred embodiment, chemically derivatized particles, plates, cartridges, tubes, magnetic particles, or other solid phase matrix with specificity to the assay components are used. The binding surfaces of microplates, tubes or any solid phase matrices include non-polar surfaces, highly polar surfaces, modified dextran coating to promote covalent binding, antibody coating, affinity media to bind fusion proteins or peptides, surface-fixed proteins such as recombinant protein A or G, nucleotide resins or coatings, and other affinity matrix are useful in this invention.

In a preferred embodiment, platforms for multi-well plates, multi-tubes, holders, cartridges, minitubes, deep-well plates, microfuge tubes, cryovials, square well plates, filters, chips, optic fibers, beads, and other solid-phase matrices or platform with various volumes are accommodated on an upgradable modular platform for additional capacity. This modular platform includes a variable speed orbital shaker, and multi-position work decks for source samples, sample and reagent dilution, assay plates, sample and reagent reservoirs, pipette tips, and an active wash station.

In a preferred embodiment, thermocycler and thermoregulating systems are used for stabilizing the temperature of heat exchangers such as controlled blocks or platforms to provide accurate temperature control of incubating samples from 0°C to 100°C.

In a preferred embodiment, interchangeable pipet heads (single or multi-channel) with single or multiple magnetic probes, affinity probes, or pipettors robotically manipulate the liquid, particles, cells, and organisms. Multi-well or multi-tube magnetic separators or platforms manipulate liquid, particles, cells, and organisms in single or multiple sample formats.

5

In some embodiments, the instrumentation will include a detector, which can be a wide variety of different detectors, depending on the labels and assay. In a preferred embodiment, useful detectors include a microscope(s) with multiple channels of fluorescence; plate readers to provide fluorescent, ultraviolet and visible spectrophotometric detection with single and dual
10 wavelength endpoint and kinetics capability, fluorescence resonance energy transfer (FRET), luminescence, quenching, two-photon excitation, and intensity redistribution; CCD cameras to capture and transform data and images into quantifiable formats; and a computer workstation.

In a preferred embodiment, the detecting is by FACS. In another aspect, the detecting is by high
15 pressure liquid chromatography (HPLC), for example, reverse phase HPLC, and in a further aspect, the detecting is by mass spectrometry.

These instruments can fit in a sterile laminar flow or fume hood, or are enclosed, self-contained systems, for cell culture growth and transformation in multi-well plates or tubes and for hazardous
20 operations. The living cells may be grown under controlled growth conditions, with controls for temperature, humidity, and gas for time series of the live cell assays. Automated transformation of cells and automated colony pickers may facilitate rapid screening of desired cells.

Flow cytometry or capillary electrophoresis formats can be used for individual capture of
25 magnetic and other beads, particles, cells, and organisms.

The flexible hardware and software allow instrument adaptability for multiple applications. The software program modules allow creation, modification, and running of methods. The system diagnostic modules allow instrument alignment, correct connections, and motor operations. The
30 customized tools, labware, and liquid, particle, cell and organism transfer patterns allow different applications to be performed. The database allows method and parameter storage. Robotic and computer interfaces allow communication between instruments.

In a preferred embodiment, the robotic apparatus includes a central processing unit which communicates with a memory and a set of input/output devices (e.g., keyboard, mouse, monitor, printer, etc.) through a bus. Again, as outlined below, this may be in addition to or in place of the CPU for the multiplexing devices of the invention. The general interaction between a central
5 processing unit, a memory, input/output devices, and a bus is known in the art. Thus, a variety of different procedures, depending on the experiments to be run, are stored in the CPU memory.

These robotic fluid handling systems can utilize any number of different reagents, including buffers, reagents, samples, washes, assay components such as label probes, etc.

10

The following examples serve to more fully describe the manner of using the above-described invention, as well as to set forth the best modes contemplated for carrying out various aspects of the invention. It is understood that these examples in no way serve to limit the true scope of this invention, but rather are presented for illustrative purposes. All references cited herein are
15 expressly incorporated by reference in their entirety.

EXAMPLES

Example 1

20 **Functional Signaling Analysis in Single Cells: Simultaneous Measurement of Multiple Kinase Activities Using Polychromatic Flow Cytometry**

Intracellular assays of signaling systems has been limited by an inability to correlate functional subsets of cells in complex populations based on kinase activity. Such correlations could be important to distinguish changes in signaling status that arise in rare cell subsets during signaling or in disease manifestation. In this Example, using the methods and compositions of the present
25 invention, the present inventors (also referred to herein as "we") demonstrate the ability to simultaneously detect the kinase activities of members of the mitogen activated protein kinases family (p38 MAPK, p44/42 MAPK, JNK/SAPK), members of cell survival pathways (PKA, AKT/PKB), and members of T-cell activation pathways (TYK2, PKC) in subpopulations of complex cell populations by multi-parameter flow cytometric analysis. Further, in this Example
30 the present inventors demonstrate the utility of these probes in identifying distinct signaling cascades for: 1) both artificial and physiological stimulatory conditions of peripheral blood mononuclear cells (PBMC); 2) cytokine stimulation in human memory and naive lymphocyte subsets as identified by 5 differentiation markers; and 3) ordering of kinase activation in potential signaling hierarchies. Polychromatic flow cytometric kinase activity measurements (PFC-KA)

demonstrate that multi-dimensional analysis of signaling pathway interplay can provide functional signaling pathway assessment on a single cell level.

Introduction

- 5 The study of multiple activated signaling pathways in complex populations of cells, such as peripheral blood, at the single cell level has not been previously possible. Multiparameter flow cytometric analysis allows for small subpopulations -- representing different cellular subsets, differentiation or activation states -- to be discerned using cell surface markers. Downstream activation of signaling pathways can be read out by reporter systems that include intracellular
- 10 detection of enzyme activity for pgalactoside^{1,2}, and β -glucuronidase³ to study transcription from defined promoter(s) and green fluorescent proteins (GFP). Interactions between proteins can be determined by both Fluorescence Resonance Energy Transfer and Bioluminescence Resonance Energy Transfer^{4,5,6,7}. In either of these latter cases, the activation of the signaling system is determined through overexpression of reporter proteins-as such the endogenous
- 15 kinase activation states are not accessible.

- It is understood that kinase signaling cascades play an important role in nearly every critical decision process in cells⁸. Determining the role of specific signal transduction pathways in a given system has been aided by the advent of pharmacological inhibitors for specific kinases.
- 20 However, monitoring activities for such kinases as Protein Kinase B/AKT, c-Jun-N-terminal Kinase (JNK), p38 mitogen activated protein kinase (p38), p44/42 ERK1/2 (ERK) activity, PKA activity, PKC activity, or TYK2 activity is usually dependent upon in vitro kinase assays or, more recently, by immunoblotting to determine their phosphorylation state using phospho-specific antibodies. To date it has not been possible to correlate rare subpopulations with the activation
- 25 state of kinases in important signaling paths. With the recent advent of 11-color flow cytometry⁹ we sought to dissect biologically relevant subsets of cells based on defined kinase activation states.

- Phospho-specific monoclonal, phospho-substrate specific polyclonal, and cognate
- 30 (nonphospho-specific) kinase antibodies were conjugated to fluorophores (see Table 1 herein; and Table 1 legends below) as probes for specific kinase activities. Each antibody was tested for phospho-specificity prior to conjugation by treating cells with appropriate kinase inducing agents or specific kinase inhibitors (see Table 2 herein; and Table 2 legends below) and subjected to traditional western blot analysis and kinase activity assays (Fig. 1A). Antibodies
- 35 exhibiting specificity for the phosphorylated state of the kinase were: AKT phospho-Ser473, AKT

- phospho-Thr308, p44/42 MAPK phosphoThr202 Tyr204, TYK2 phospho-Tyr1054/1055, p38 MAPK phospho-Thr180 Tyr182, PKC-PAN phospho-substrate (detects PKC α , β I, β 11, and δ isoforms only when phosphorylated at a carboxy terminal residue homologous to ser660 of PKC β 11), PKA phospho-substrate (consensus phosphorylation site of PKA phosphorylation of
- 5 threonine with arginine at -3 position), JNK/SAPK phospho-Thr183/Tyr185, phosphotyrosine (p-tyr-100) and phospho-threonine. All kinase activation states were verified using specific inhibitors for the upstream kinase known to phosphorylate the cognate kinase (Fig. 1A). The inhibitors applied are listed in Table 2.
- 10 FACS based detection of kinase activity, after 24-hour serum-starvation, was tested as a function of a given stimulus specific to kinase activation and inhibition by kinase selective inhibitors. Table 1 provides staining combinations for the flow cytometry experiments used in this report. Kinase selective activators and inhibitors used are listed in Table 2 and FACS based kinase activity plots are illustrated in Figure 1B. Kinase probes for p44/42, AKT, JNK, TYK2, p38, and
- 15 phospho-tyrosine containing proteins exhibited the strongest resolution of peaks between uninduced and induced conditions (Fig. 1B). Kinase probes for PKC and PKA exhibited the lowest apparent induction from basal fluorescence levels as determined by FACS; this could have been dependent on the conditions we had established for induction and staining or the substrate motif recognition might not be optimal for these probes. We therefore determined that
- 20 the fluorescence observed was due to a phosphorylation of a target protein by treating the sample prior to staining with one or more phosphatases to support the phospho-PKC-PAN, phospho-PKA substrate, and phospho-TYK2's probe phospho-specificity (Fig. 1 C). Figure 1C highlights the phospho-PKC-PAN, phospho | PKA-substrate and phospho-TYK2 probes are dependent on their respective phosphorylated substrate for recognition as phosphatase
- 25 treatment diminishes their ability to recognize the stimulus induced phosphorylation product(s).

To exemplify that these probes can differentiate amongst varying levels of kinase activity a FACS dose response curve demonstrates there is excellent p44/42 activity to fluorescence correlation (Fig. 1B, bottom right panel). Furthermore, the mean fluorescence intensity values of the

30 phospho-p44/42 probe had a correlation coefficient of 0.96 with the densitometry values of phosphorylated p44/42 in a dose response curve, validating the kinase probes as being able to measure small differences in kinase activity (Fig. 1 D).

To demonstrate the ability of these probes to represent biologically relevant signaling events we

35 activated peripheral blood mononuclear cells (PBMC), a population of cells that is likely to be

heterogeneous in terms of potential kinase activities, to see if subsets of cells demonstrated differing kinase activation kinetics. Total PBMC in mixed culture were stimulated with CD3 and CD28 monoclonal antibodies to mimic physiological T cell activation of surface receptors. As another form of stimulus we activated these cells with the artificial T cell activation agents phorbol myristyl acetate (PMA) and ionomycin. Cell subsets were gated for CD4+ (a T cell subset marker) or CD19+ (a B cell marker). The T cell and B cell subsets were compared to discern differential kinase activity profiles between these two cell types upon a given stimulus. Figure 2A illustrates that primary B and T cells have inherent differences with respect to kinase activity of p38 MAPK and p44/42 MAPK in an unstimulated state, and respond differently to the various stimulatory conditions applied. Dramatic differences for T cells in both p38 and p44/42 activity can be seen when comparing PMA/ionomycin and CD3/CD28 stimulation, suggesting that CD3/CD28 does not engage full activation of CD4+ T cells along the p38 or p44/42 pathway. Some activation of the CD19+ B cells is observed using combinations of CD3 and CD28 that we presume is due to T cell activation and their production of stimulatory cytokines: for B cells. Alternatively other T cell helper interactions with B cells could lead to the B cell activations observed with CD3 and CD28.

We also determined in a cell line differential responses to cytokines. Using CD69, an early T cell activator marker^{10,11} as a surface antigen to distinguish between activated and non-activated Jurkat cells, CD69 expression was plotted as a function of p44/42 activity to a variety of T cell stimulatory agents (Fig. 2B). The differences observed in kinase activity and CD69 expression when comparing CD3/CD28 stimulation to that of PMA/ionomycin (artificial) would suggest that other co-stimulatory molecules exist and are necessary for optimal T cell activation with respect to p44/42 MAPK. In addition, experiments involving PMA/ionomycin stimulated Jurkat cells demonstrated decreased activation of TYK2 activity in CD69+ cells (data not shown) This would suggest that T cell activation could induce intrinsic regulatory mechanisms that cause cells to become hyporesponsive to further stimuli and point out that the kinase profiling probes can potentially be used to study anergic responses.

As an example of the type of information yielded by kinase-activity gating analysis, both traditional histogram analysis and kinase-activity gating analysis was undertaken and is presented as follows. Jurkat cells in either growth media or stimulated with PMA/ionomycin were assessed for multiple intracellular kinase activity and CD69 expression (Fig. 2C). Basal activity of PKC, p38 and p44/42 is observed, however, kinase activity for JNK, p38, p44/42, PKC, and PKA is dramatically altered for PMA/ionomycin stimulated cells (Fig. 2C). This is congruent with the observations for the CD69+ PBMC data (Fig. 2B). Thus, monitoring phosphorylation events in

intact cells makes it possible to correlate activity with other parameters such as surface molecule expression, cell size, or cell granularity approaches not possible with conventional lysate analysis of kinase phosphorylation states.

- 5 To further explore subset analysis of cells we anisomycin-treated Jurkat cells and probed them for total p38 MAPK (using an antibody against a presumed nonphosphorylated region of p38) and with phospho-p38 MAPK probe. This would allow us to follow not only the changes to p38 phosphorylation but also to simultaneously measure steady state levels. Since FACS based kinase assays allow alternative analytical correlations to be made with cellular size (forward
- 10 scatter) and shape of the cell (side scatter), we determined the status of the phosphorylation as a function of the side scatter parameter (a measurement of granularity). Cell granularity is a function of surface molecule distribution, cell structure and conformation, and can be influenced by internal cellular signals that result in outward cell surface modulation (events that are common for instance in the field of integrin "inside-out" signaling^{12,13,14,15}). Contour plots depict a shift from
- 15 non-phosphorylated to phosphorylated p38 as the granularity of the cell increases (Fig. 2D). Similarly, a direct correlation between elevated p44/42 kinase activity and increased cell size (forward scatter parameter) was observed when kinase activity was monitored as a function of cell size (data not shown). We note that the phospho-p38 fluorescence staining occurs at the expense of the apparent p38 nonphospho staining. We believe this might be due to occlusion of
- 20 binding of the nonphospho-specific probe in the activated state by a cellular protein (see below).

- We illustrate a three-kinase gating analysis by correlating the phosphorylated states of p38 and p44/42 to phospho-JNK for total PBMC stimulated with IL-1a (Fig. 2E). Excerpts from regions representing the lower end of phospho-JNK levels, up to higher levels, demonstrate an initial
- 25 phosphorylation of p44/42 is followed by a phosphorylation in p38 as the intensity of JNK phosphorylation increases. Thus in the cell, higher levels of JNK activation correlate to an apparent passing of signals from p44/42 to p38. Although the phosphorylation events are not normalized according to time, the phosphorylation shifts observed in p38 and p44 as a function of JNK phosphorylation suggest pathway interplay, and could represent the result of one signaling
- 30 cascade activating another (further experiments are underway to determine the cell subsets responsible for these shifts). Simultaneous monitoring of AKT, PKA, p44/42, and p38 kinase activity as a function of TYK2 activity in IL-1 p stimulated PBMC exemplifies that kinase cascades can be ordered with respect to their activation status when analyzed in a multi-parameter fashion (Fig. 2F). In regards to IL-1 β stimulation, concomitant activation of p38
- 35 and p44/42 is observed when activation of TYK2 begins to be detected. However, a distinct profile is observed with respect to AKT and PKA activation, as the shift in detected

phosphorylated AKT is correlated to lower levels of phospho-TYK2 and PKA phosphorylation of PKA substrates. The utility of these probes to spatially resolve as many as five simultaneous kinase activations allows global assessment of signaling pathways. In addition, population subsets can be distinguished based upon intracellular kinase activity and be correlated for phenotype by using multiple surface markers- (alternative data analysis not presented here). To demonstrate the ability of these kinase probes to screen for activation of individual signaling pathways to other specific stimuli, we treated human peripheral blood monocytes (PBMC) with several different cytokines. We used the phospho-kinase probes to correlate phospho-kinase profiles in the responding cells using polychromatic flow cytometry (11-color, 13 parameters).

10 In initial experiments functional characterization of T cells occasionally varied from donor to donor. Apparent donor variability was reduced when magnetically activated cell sorting (MACS) was used to isolate resting T cells by negative isolation prior to stimulation and analysis. This is most likely due to the depletion of pre-existing activated T cells and other cell types that might be producing cytokines. Figure 3 illustrates a flow diagram of the protocol used to isolate, characterize, and analyze truly naive ($CD3^+ CD4^+ CD8^- CD62L^+ CD45RA^+ CD11a^{dim} CD28^- CD27^+$) and memory ($CD3^+ CD4^+ CD8^- CD62L^- CD45RA^+ CD11a^{br} CD28^- CD27^-$) T cell intracellular kinase activities. We observed clear constitutive activation, for instance, of AKT in the CD4, and to a lesser degree CD8, positive subsets of naive T cells, as opposed to memory T cells, even in non-activated cell types (Fig. 3, bottom panels). Other differences were apparent as well in the non-stimulated cells, including differences in p38 phosphorylation. Figure 4 demonstrates the power of polychromatic flow cytometry based kinase assays to identify functional differences in naive and memory T cells in their response to IL-1 α , IL-1 β , and IL-2 as measured by the phospho-kinase profiles. Interestingly, stimulation with any of these cytokines led to a drop in p44/42 phosphorylation in CD4+ naive cells relative to CD8+ cells. A less dramatic drop also occurs in JNK activity in these same cell subsets. Other kinase profiles are not effected.

Differences in kinase expression as consequences of stimuli are typically controlled for in standard western blot/kinase analysis by protein content normalization or by running a comparative blot with a normalizing antibody that recognizes the kinase irrespective of phosphorylation. We determined whether we could simultaneously assess kinases in their active and inactive state using cognate non-phospho specific fluorescent probes for AKT, p44/42 MAPK, p38 MAPK, and JNK/SAPK (normalizing antibodies). In unstimulated cells, we observed strong staining using the normalization antibody to p44/42. After EGF stimulation, all cells showed phosphorylated p44/42, but the normalization antibody revealed a negative population.

This result could be due to EGF-induced binding by an unknown protein to the site recognized by the p44/42 antibody probe, or a change in the structure of the p44/42 molecule that occludes the antibody binding site. For p38 we observed a strong increase after anisomycin stimulation in the levels of apparent p38 (or loss of a bound protein that occludes the normalization antibody recognition site) as well as a subset of cells that show increased phosphorylation of p38. For JNK, three distinct populations were observed before stimulation and four distinct populations could be visualized after stimulation.

One possibility is that the plots could depict two phosphorylation events of the p46 and p54 isoforms of JNK/SAPK, phosphorylation of JNK coupled by a rapid onset of JNK or p54 isoform expression, or if phosphorylation alters the capacity for interacting proteins to occlude/expose the antibody recognition site(s). For AKT, two distinct cellular populations that differed from the unstimulated state appeared after PDGF stimulation of Jurkat cells. The population of cells that had a higher AKT fluorescence intensity existed in both a mono and bi-phosphorylated state (Fig. 5A lower panel). The apparent AKT negative population of PDGF stimulated cells presented minimal phosphorylation as detected by the phospho-thr308 and the phospho-ser473 probes. It is unlikely that total AKT levels rapidly decrease within the 10 minutes of PDGF stimulation since this was not observed in standard western and kinase assays (see Fig. 1A) and thus we infer a new protein binds there or a signal induced change in structure prevent binding of the antibody.

These latter results lead us to believe that the changes in subpopulations observed represent changes in the binding of cellular proteins to the kinases. Thus, there are potential pitfalls in the simultaneous use of antibodies to normalize intracellular staining on the basis of a normalizing antibody- this warrants caution in the interpretation of such data. However, the results suggest that a series of antibodies, that recognize different epitopes on the same protein, could both infer the existence of cellular binding partners, map where they interact on the target protein structure, and derive specific functional correlations associated with such binding. We know of no other technique that allows protein interactions to be mapped on endogenous proteins in a cell-by cell analysis of subsets of cells in complex populations.

Summary

Utilizing multi-parameter flow cytometry, we demonstrate that lymphocyte subsets can be distinguished based upon levels of as many as five intracellular kinase activities. The power of the approach can be appreciated not only in dissection of intracellular signaling pathways, but presents for the first time the ability to perform multiple kinase activity measurements in a single cell by flow cytometry. Functional signaling dissection on a single cell level could be applicable

to rare cell subpopulations such as stem cells or stem cell progenitors and can be extended to many systems where the rarity of certain cellular subsets preclude isolation of cells for in vitro functional characterization. The application of these kinase specific probes is the following: (1) To obtain a unique signaling profile for a stimulus to understand the biology of various cellular processes (2) To monitor signaling mechanisms in rare cell populations in the development, differentiation, or maintenance of cellular fates, (3) The potential to utilize these kinase probes (and others) in high-throughput FACS based screens for pharmacological inhibitors of specific kinase activity within mammalian cells, (4) the potential to temporally and spatially resolve kinase activation cascades on a global scale, and (5) the possibility that binding partners might be inferred before and after stimulation with appropriate antibody sets. Development of other intracellular kinase probes can contribute to the elucidation of signaling networks and in combination with polychromatic flow cytometry (11-color, 13 parameter) technology⁹ could aid an understanding of the complexity of intracellular signals in the immune system. Furthermore, the utility of these probes might be extended to clinical settings to determine intracellular kinase activities for cells derived from diseased patients as a diagnostic indicator of disease progression.

Experimental Procedures

Cell Culture and Primary cell Isolation

Jurkat T cells were maintained in RPMI1640, 10% FCS 1% PSQ (10% fetal calf serum, 1% penicillin-streptomycin (1000 units/ml and 2mM L-glutamine PSQ). NIH3T3 cells were maintained in DMEM (Dulbecco modified eagle medium), 10% donar calf serum, 1% PSQ. For preparation of primary cells, mononuclear cells were isolated from blood of healthy donors (Stanford Blood Bank, Stanford, California). Human peripheral blood monocytes were obtained by Ficoll-plaque density centrifugation (Amersham Pharmacia, Uppsala, Sweden) of whole blood and depletion of adherent cells by adherence to plastic culture dishes. Isolated cells were maintained in complete media. Magnetically activated cell sorting (MACS, Miltenyl Biotech, Baraisch-Gladbach, Germany) was used in some experiments as noted to enrich for CD3+ T cells by negative isolation using combinations of CD16, CD14, CD44, HLA-DR, or CD19 biotinylated antibodies (Dyna, Oslo, Norway) and streptavidin-magnetic beads (Dyna, Oslo, Norway). Chemical and biological agents used were either obtained from Sigma (St. Louis, Missouri), Cell Signaling Technologies (Beverly, Massachusetts), Roche Biochemicals (Indianapolis, Indiana), or Peprotech (Rocky Hill, New Jersey): ionomycin (1- μ M, Sigma), phorbol myristyl acetate (1 μ M, Sigma), anisomycin (10 μ M, Sigma), PDGF 10 pg/ml or as indicated (Roche Biochemicals), SB203580 (10 μ M, Cell Signaling Technology), LY294002 (10 μ M, Cell Signaling Technology), U0126 (10 μ M, Cell Signaling Technology), Bisindolymaleimide II (10 μ M, Sigma), Calyculin A

(10 μ M, Sigma), staurosporine (1 μ M, Sigma), genistein (10 μ M, Sigma), IL-2 (10 pg/ml, Peprotech), IL-6 (10 pg/ml, Peprotech), IL-1 α (Roche Biochemicals (10 pg/ml), epidermal growth factor (EGF, Roche Biochemicals), lambda phosphatase and calf intestinal phosphatase (CIP) (New England Biolabs, Beverly, Massachusetts), CD40L was a gift from Dr. Strobl (Department of Molecular Pharmacology, Stanford University). CD3 (OKT3) and CD28 endotoxin/azide free monoclonal antibodies were from PharMingen (La Jolla, California).

For serum starvation, Jurkat cells were deprived of serum for 24 hours and were considered uninduced with respect to kinase activity. Kinase activity inhibition was done with the appropriate kinase inhibitor added 30 minutes prior to being stimulated by the kinase activating agent. Stimulation by either kinase activating agent or exogenous stimuli was performed for 30 minutes before cells were prepared for flow cytometry. Stimulation of Jurkat cells with PMA/Ionomycin was for 2 hours at 1 μ M. Stimulation of PBMC was performed for 12 hours prior to 11-color analysis at 10 ng/ml of indicated cytokine. Chemical agents were dissolved in DMSO (Sigma), and uninduced cells received 0.1% DMSO carrier as control treatment. UV treatment for samples in Figure 1A was performed by incubation under a UV light in a typical tissue culture hood for the indicated time. Lambda phosphatase treatment (1000 units at 37°C for 15 min) and calf intestinal phosphatase treatment (CIP, 50 U at 37°C for 15 min) was performed after FACS permeabilization prior to intracellular staining.

Immunoblotting

Cell extracts were prepared by washing 2×10^6 cells in ice cold PBS and harvesting in lysis buffer (20mM Tris pH 7.5, 150 mM NaCl 1mM EDTA 1mM EGTA, 1% Triton X100, 2.5 mM Na_2PO_4 , 1mM β -glycerolphosphate, 1mM Na_3VO_4 , 1 μ g/ml Leupeptin, 1mM PMSF, protease inhibitor cocktail tablet (Boehringer Mannheim, Basel, Switzerland). Extracts were centrifuged 14,000 RPM for 5 min at 4°C and cell lysates (20 μ g as determined by BCA protein assay (Pierce, Rockford, Illinois) were fractionated on 12% or 15% SDS-polyacrylamide gel electrophoresis and transferred to PVDF membranes using standard procedures. Antibodies used are indicated adjacent to blots and were purchased from Cell Signaling Technologies (Beverly, Massachusetts). Cell treatments are indicated where appropriate.

Kinase assays

Kinase activity assay protocol was similar for AKT, p38, p44/42, JNK and substituted reagents used for respective kinase assay is noted. All reagents used for kinase activity assays were obtained from Cell Signaling Technologies (Beverly, Massachusetts). Kinase activity was

detected by immunoprecipitation of kinase AKT, p38, p44/42, or JNK using immobilized AKT1G1 monoclonal antibody, immobilized phospho-p38 MAPK monoclonal antibody, immobilized phospho-p44/42 monoclonal antibody, or c-Jun fusion protein beads respectively, from cells. Immobilized kinase was used in a kinase assay with GSK3 fusion protein, ATF-2 fusion protein, 5 Elk-1 fusion protein, or c-Jun fusion protein beads respectively. 2×10^6 NIH3T3 or 2×10^6 Jurkat cell were washed twice in PBS and cells were incubated with immobilized kinase antibody or beads (1:200) at 4°C with gentle rocking motion for 2 hrs. Immunocomplexes were washed 4X with cell lysis buffer and resuspended in 40 µl kinase buffer (25 mM Tris pH 7.5, 5 mM β-glycerolphosphate, 2mM DTT, 0.1 mM Na_3VO_4 , 10 mM MgCl_2) supplemented with 200 µM ATP and 1 µg of fusion protein (except for JNK activity assay, in which ATP was sufficient to induce 10 activation) for 30 min at 30°C. Kinase reaction was terminated with SDS sample buffer boiled for 5 min, and phosphorylation state of GSK3, ATF, Elk-1, or cJun was detected by immunoblotting with phospho specific antibodies phospho-GSK3αβ (ser21/9), phospho-Elk-1 (ser383), phospho-ATF2 (thr71) or phospho-cJun (ser63) respectively and visualized using ECL detection 15 (Amersham). Immunoblots are representative of 3 independent experiments.

Kinase Probe Generation

Antibody conjugation was done using standard procedures (<http://drmr.com.abcon>) or by using protein-protein/protein-dye crosslinking kits from Molecular Probes (Eugene, Oregon). 20 Phospho-specific and non-phospho specific antibodies were from Cell Signaling Technologies (Beverly, Massachusetts) or as indicated. Antibodies conjugated were the following: phospho-AKT Ser473 monoclonal anti-4E2, phospho p44/42 MAP Kinase (Thr202/Tyr204) monoclonal antibody, phospho-TYK2 (Tyr1054/1055) antibody, phospho-p38 MAP Kinase (Thr180/Tyr182) monoclonal antibody 28B10, phospho-PKC-PAN substrate antibody, 25 phospho-PKA-substrate, phospho-SAPK JNK (Thr183 Tyr 185) G9 monoclonal antibody, phospho-tyrosine monoclonal antibody (P-tyr-100), p44/42 MAPK, p38 MAPK, JNK/SAPK, and phosphoAKTthr308. Anti-TYK2 was obtained from Santa Cruz Biotechnologies (Santa Cruz, California). Phospho-PYK2 (pY402) antibody and phospho-FAK (pY397) were obtained from Biosource (Carillo, California). Alexa-Fluor dyes (Alexa Fluor 350, Alexa Fluor 430, Alexa Fluor 488, Alexa Fluor 546, Alexa Fluor 568, Alexa Fluor 594, Alexa Fluor 633, Alexa Fluor 660 Alexa 30 Fluor 680), Cascade Blue, Cascade Yellow and Rphycoerythrin (PE) were purchased from Molecular Probes (Eugene, Oregon). FITC, Rhodamine, and Texas Red were purchased from Pierce (Rockford, Illinois). Cy5, Cy5.5, Cy7 were obtained from Amersham Life Science (Pittsburgh, Pennsylvania). Tandem conjugate protocols for Cy5PE, Cy5.5PE, Cy7PE, 35 Cy5.5APC, Cy7APC can be found at www.drmr.com/abcon. Quantitation of fluorescent probe conjugation was assessed to determine degree of labeling and protocols including dye spectral

properties can be found at www.metazoa.com/UPL3419. In general, probes used had 29 moles of fluorescent dye per mole of protein.

Flow Cytometry and FACS Analysis

- 5 Intracellular and extracellular staining was initially performed as described (www.metazoa.com/UPL3287) but modified. Approximately $1-10 \times 10^7$ peripheral blood mononuclear cells were obtained by magnetically activated cell sorting, treated with a stimulatory condition for 12 hours, and prepared for flow cytometry. Surface stained for 20 minutes, and divided into aliquots for intracellular stain reagents (see below for details). Intracellular staining
- 10 was achieved by permeabilizing with CytoFix/CytoPerm buffer (PharMingen, La Jolla, California), intracellular stained for 30 minutes on ice, and final resuspension in 1% paraformaldehyde. Isotype control match antibodies were used for stain controls. Purified antibodies to cell surface markers listed in Table 1 in addition to the following were either obtained from PharMingen (La Jolla, California) or conjugated to the indicated fluorophore in the Herzenberg laboratory
- 15 (Department of Genetics, Stanford University): CD69-APC, CD69-PE, CD4-FITC, CD4-APC, CD4-PE, CD3-APC, CD3-PE, CD3-PERCP, CD19-FITC, CD19-APC.

- For each blood sample, PBMC were stained with all the reagents in stain combination 4, 5, 6 (Table 1) except for FITC, PE, Alexa 594, and APC. The stained cells were divided into aliquots
- 20 and then individually stained with different intracellular cocktail combinations as depicted in combinations 4, 5, 6 (Table 1). Intracellular FACS staining requires mild cell fixation and subsequent permeabilization. Though it has been possible to perform intracellular stains by using a two-step antibody staining protocol (primary plus conjugated secondary), optimal results were achieved with a primary conjugated antibody. Furthermore, conjugates comprised of
- 25 phycobiliproteins such as phycoerythrin (PE) and allophycocyanin (APC), required significant optimization of permeabilization conditions perhaps because phycobiliproteins are large and might contribute to background binding to intracellular surfaces. For these reasons, the phospho-specific probes generated using the Alexa-Fluor dye series were used in multi-label experiments. In addition, several phospho-specific probes developed for other kinase activities
- 30 (phospho-PYK2, phospho-FAK, among others) that recognize the phosphorylated form of these antibodies in standard western blots lost their ability to recognize the phosphoepitopes by FACS. This could be either due to the fixation process altering the epitope recognized by the antibody, or the epitope being occluded from antibody recognition in the protein's native form. After extensive testing we determined that in general, directly conjugated phospho-specific monoclonal
- 35 antibodies exhibited the greatest signal to noise ratio.

Single and four color flow cytometry data acquisition was collected on a FACSCalibur machine (Beckton Dickinson, San Jose California.) using CELLQuest software, analyzed and presented using FlowJo software (Tree Star, San Carlos, California). Data for four or less colors are representative of 3 independent experiments of 10^5 cells/sample and 50,000 events were collected and calibrated using Calibrite beads (Becton Dickinson, San Jose California). 11-color data acquisition was collected on a modified FACStarPlus (Becton Dickinson, San Jose, California) connected to MoFlo electronics (Cytomation, Fort Collins, Colorado). Data were collected by FACSDesk software and compensated, analyzed, and presented using FlowJo software. Data for 11-color flow cytometry collected a minimum of 200,000 events.

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Table 1

Top table shows single, 3, and 4 color combinations were used for Figure 1 and 2A-D. The antibody-dye sets for these can be found in rows 1, 2, and 3 above. The antibody-dye combinations are grouped according to their excitation wavelength as determined by the laser line applied during the experiment. 11-color staining combinations (rows 4, 5, 6) were used to generate the data in Figures 2E-F, 3, 4, and 5. Regions left blank indicate no reagent used. Conditions used ethidium monoazide bromide (EMA) to identify dead cells by intercalating into DNA and forming a covalent adduct prior to permeabilization of the cells. Serum underlays, as well as forward and side scatter gating were used to physically remove and identify dead cells, respectively (Fig. 4).

Bottom table illustrates Alexa fluorescent dye labeled probes used in these experiments. The various kinases were conjugated with one or more of 7 different Phospho-specific antibodies labeled with Alexa dyes are referred in the above tables as p-probe to indicate phospho specific probe was used in that color. They were applied in the various stains in the top section of Table 1 to allow for complementary dye combinations for multiparameter analysis.

Table 2 shows agents used to induce and inhibit specific kinase activity in serum starved Jurkat cells are denoted^{3,20-32}. 24 hour serum starvation reduced background phosphorylation significantly. Concentrations used other than those depicted is stated where appropriate. See Methods for treatment protocols.

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Example 2

LFA-1 functionally contributes to T-cell activation and polarized effector cell cytokine production to a TH1 profile

In this Example, using the methods and compositions of the present invention, the present inventors (also referred to herein as "we") demonstrate that LFA-1 stimulation by ICAM-2 functionally contributes in activating human naïve T-cells (CD3⁺CD4⁺CD8⁻CD62L⁺CD45RA⁺CD28⁺CD27⁺CD11a^{dim}) in the presence of CD3/CD28 co-stimulation as determined by simultaneous multidimensional (33 parameters) assessment of immunophenotype, phospho-protein signaling, cytokine production, and transcriptional activity.

Soluble ICAM-2 (sICAM-2), upon interacting with LFA-1, initiated phosphorylation and release of LFA-1 binding proteins cytohesin-1 and JAB1, which induced activation of the p44/42 MAPK pathway and cJun respectively. sICAM-2 enforced rapid kinetics of multiple active kinases (p44/42, p38, JNK MAPKs, Ick, AKT, PLCg) when combined with CD3/CD28 stimulation, lowering the CD3/CD28 stimulus threshold by 10,000 fold as determined by activation markers CD25 and CD69. CD3/CD28/LFA-1 signal integration resulted in enhanced nuclear localization of NF-kB, Creb, Atf2, c-Fos, and c-Rel. These conditions optimized T-cell activation as measured by IL-2 expression, NFAT, NF-kB, and AP-1 activity, and cell cycle progression. Furthermore, the CD3/CD28/LFA-1 trio-stimulation polarized naïve human T cells towards a TH1 phenotype, with CD3/CD28 stimulus favoring a TH2 phenotype. These results demonstrate that ICAM-2 interaction with LFA-1 can functionally contribute in T cell activation and differentiation.

In particular, this Example demonstrates that polychromatic flow cytometry based on the methods and compositions of the present invention allow for the rapid identification of specific

lymphocyte subsets with simultaneous measurement of multiple intracellular kinase activities. This approach provides functional signaling information on unique cellular processes, for example, T cell activation, at the single cell level.

5 Introduction

T-cell activation is a multi-step process involving integration of antigen receptor (TCR-CD3) signals with co-stimulatory signals such as B7/CD28 (Croft and Dubey, 1997). Engagement of the antigen-specific TCR-CD3 complex with a recognized peptide-MHC complex on antigen-presenting cells (APC) is necessary for in vivo T-cell activation. T-cell co-stimulatory surface
10 molecules CD28, CD2 (LFA-2), and their APC counter receptors B7 (CD80-86), CD58 (LFA-3), initiate T cell activation (Bjorndahl et al., 1989; Cefai et al., 1996; Cerdan et al., 1992; Chambers and Allison, 1999; Lenschow et al., 1996). Adhesion between T-cells and APCs is necessary for the formation of the immunological synapse and is mediated by T-cell surface molecules LFA-1 (CD11a/CD18) (de Fougerolles et al., 1991; Dustin et al., 1989; Koopman et al., 1992) and the
15 intercellular adhesion molecule ligands (ICAMs).

In contrast to the extensive data available concerning signals involved in TCR-CD3 and CD28 ligation, much less is known about specific signals generated by adhesion mediated events involved in T cell contact with APCs. However, there is evidence that optimal proliferation of T-cells and enhancement of IL-2 production requires stimulation other than TCR-CD3 and CD28
20 that results from adhesive interactions between costimulatory receptors on T-cells and their counter ligands on APCs (Fine and Kruisbeek, 1991; Salomon and Bluestone, 1998; Simmons, 1995; Starling et al., 1995; Zhang et al., 1997). Insect and fibroblast model systems utilized to study T cell activation have demonstrated that increasing antigen density by more than 10,000-fold fail to initiate naïve CD4+T cell proliferation or cytokine synthesis in the absence of an
25 ICAM/LFA-1 interaction (Abraham et al., 1999; Ragazzo et al., 2001). In addition, it has also been observed that the ICAM-1/LFA-1 interaction can promote the appearance of the hyper-phosphorylated p23 form of the TCR ζ chain (Ragazzo et al., 2001). Despite these and other observations that in co-ligation with TCR-CD3, LFA-1/ICAM interaction can lead to sustained intracellular calcium response, and increased inositol phospholipid hydrolysis (van Seventer
30 1992; Wulfig 1998; Rovere 1996), the specific functional consequences for an LFA-1 mediated "co-stimulatory" signal are not fully understood. Furthermore, these latter studies downplay a suggestion by some that LFA-1/ICAM interactions simply allow for longer TCR engagement and rather suggested to us that the LFA-1/ICAM interaction is capable by itself of initiating important intracellular signaling events.

35 Development of methodologies for intracellular phospho-protein staining has allowed us to simultaneously monitor multiple active kinases in up to 13 dimensions in single cells (Perez and

Nolan, 2002). Utilizing this in conjunction with surface immunophenotyping, transcription factor nuclear profiling and cytokine bead arrays, we observed that human naïve T cells (CD3+CD4+CD8-CD62L+CD45RA+CD11a^{dim}CD27+CD28+) are optimally activated upon CD3/CD28/LFA-1 stimulation. Co-stimulation with LFA resulted in an over 10,000 fold higher sensitivity to CD3/CD28 co-stimulation. In characterizing the mechanism of this observation, we demonstrate that cytohesin-1, an LFA-1 interacting protein, is released and phosphorylated upon LFA-1 stimulation, and mediates activation of the p44/42 MAPK pathway, correlated to activation of several downstream transcriptional regulators. In conjunction, JAB1, another interacting LFA-1 protein, is similarly released and phosphorylated upon LFA-1 stimulation, leading to c-Jun phosphorylation in the absence of JNK activity. The integration of signaling pathways from CD3, CD28, and LFA-1 converge to mediate activation of multiple pathways (MAPKs, PKC, Src kinases, and focal adhesion kinases) which enhanced NFAT, NF- κ B, and AP-1 activities, cell cycle entry, and expression of IL-2, CD25, and CD69. Furthermore, the CD3/CD28/LFA-1 stimulus polarized T cell differentiation towards a TH1 phenotype as compared to the TH2 phenotype observed for a CD3/CD28 stimulus. Thus, the additional LFA-1 signal contributes functionally to both T cell activation and effector cell differentiation.

Results

ICAM-2/LFA-1 interaction induces release and phosphorylation of JAB1 and cytohesin-1

In a series of kinase profiling experiments we had observed that a solubilized version of membrane- ICAM-2 (sICAM-2, see Materials and Methods) induced activation of the p44/42 MAPK through LFA-1 activation (*Perez and Nolan, manuscript submitted, Reviewers please see accompanying copy*) that kinetically differed from CD3 or CD28 stimulation (data not shown). It was also observed that LFA-1 stimulation induced c-Jun phosphorylation in the absence of JNK activity (data not shown). We therefore decided to investigate the functional differences of the ICAM-2-induced signaling in human naïve CD4+ T cells.

LFA-1 stimulation induced phosphorylation and nuclear translocation of c-Jun, an effect that was additive with CD3/CD28 stimulation (Figure 6A). Inhibition of JNK with the specific chemical inhibitor SP600125 (Bennett et al., 2001) did not abrogate the sICAM-2 induced cJun phosphorylation as determined by single cell analysis (Figure 6B, top panel). Recently, JAB1 has been identified as an LFA-1 binding protein that mediated cJun phosphorylation in the absence of JNK activity (Bianchi et al., 2000). Intracellular delivery of α -JAB1 antibodies (see Materials and Methods), abrogated LFA-1 induced c-Jun phosphorylation (Figure 6B, bottom panel). Visualizing both cytohesin-1 and JAB1 in T cells revealed membrane delocalization upon LFA-1 stimulation (Figure 6C). Cells artificially loaded with anti-JAB1 exhibited membrane

delocalized JAB1, in the absence of stimulus, and exhibited no change in cytohesin-1 localization (Figure 6C). Thus, antibody disruption of JAB1 from LFA-1 blocked the LFA-1 induced c-Jun phosphorylation, indicating that the receptor-ligand interaction was necessary for receptor signaling to c-Jun through JAB1.

5 We assessed if modifications to cytohesin-1 and/or JAB1 occurred upon LFA-1 stimulus. It was observed that LFA-1 stimulation induced threonine phosphorylation of cytohesin-1 and serine phosphorylation of JAB1, an effect that was not observed after CD3 stimulation (Figure 6D). We investigated if JAB1 or cytohesin-1 was responsible for LFA-1 induced p44/42 MAPK activity as previously observed (*Reviewers please see attached manuscript*) by using
10 intracellular delivery of antibodies and/or peptides to functionally modulate respective protein's signaling. Intracellular delivery of an α -cytohesin-1 antibody to the C-terminal blocked the LFA-1 induced p44/42 MAPK activity, a property shared by intracellular delivery of a peptide corresponding to the cytohesin-1 C-terminal (Figure 6E). Neither intracellular delivery of α -JAB1 or an IgG control affected the LFA-1 induced p44/42 MAPK activation (Figure 6E). In
15 contrast, intracellular delivery of antibodies directed at the cytohesin-1 N-terminal induced p44/42 MAPK activation (Figure 6E). These results are supported by the observation that the N-terminal of Cytohesin-1 contains the domain that directly interacts with LFA-1 (Geiger et al., 2000; Kolanus et al., 1996). Therefore, the dissociation of cytohesin-1 from LFA-1, here induced by antibodies to the N-terminal, led to dissociation of the LFA-1/cytohesin interaction in
20 vivo and consequently activated the p44/42 MAPK. The intracellular delivery of antibodies against JAB1 were capable of interfering with JAB1 binding and cJun phosphorylation whereas intracellular delivery of a corresponding C-terminal sequence of cytohesin-1 blocked LFA-1 induced p44/42 MAPK activity. Thus, intracellular delivery of antibodies can functionally modulate protein function and suggest that distinct signaling pathways are coupled to
25 cytohesin-1 and JAB1, two LFA-1 interacting proteins that were mobilized upon stimulus.

Integration of CD3, CD28, and LFA-1 signaling supercede signaling induced by CD3/CD28 co-stimulation

We had performed kinetic analyses of multiple active kinases by flow cytometry to CD3, CD28 and LFA-1 stimulation (*Reviewers please see Supplementary Figure 6*). Since activation rates
30 varied for individual stimuli tested (data not shown), we decided to study the combinations of CD3/CD28 versus CD3/CD28/LFA-1 in phospho-protein based analyses. We developed a phospho-protein array by spotting a combination of 96 phospho and non-phospho specific

antibodies for differential analysis of signaling proteins induced by CD3/CD28 versus CD3/CD28/LFA-1 stimulation in purified human naïve CD4 T cells. (Figure 7A). Enrichment of primary naïve T cells was performed by magnetically activated cell sorting using markers CD8, CD19, CD16, CD14, HLA-DR, and CD44 to remove cytotoxic T cells, B cells, NK cells, macrophages, and activated T cells from human PBMC respectively. After lysis of cells, equivalent protein lysates were labeled with Alexa532 or Alexa680, and the samples were co-hybridized to the array. The fluorescent intensities for both dyes were read by a fluorescent plate reader, and the log ratios were computed to obtain a differential signal between the stimulations tested. The fluorescent intensities for both channels is plotted on an XY coordinate grid (Figure 7A). The fold induction in phosphorylation, as determined by the log ratios, is color coded to represent fold change (from red being 0 fold induction to green being 5 fold induction) and is displayed on the plotted fluorescence intensity coordinates yielded for CD3/CD28 and CD3/CD28/LFA-1 stimulation (Figure 7A). Analysis identified a 2-5 fold phospho-induction of MAPK kinases (p44/p42, p38, JNK, raf, mek1/2, mek3/6, rsk, raf), src related kinases (src, pyk2), and integrin signaling kinases (FAK), among others (Figure 7A). Internal reference controls of non-phospho proteins (p44/42 MAPK, p38 MAPK, JNK, mek1/2, mek3/6, rsk, raf, src, pyk, fak) for these kinases showed induction in the range of 0.8-1.5 fold and were not expected to change significantly at the endpoint tested (data not shown). Antibodies against structural proteins (actinin, actin, tubulin) were used to verify equivalent protein quantities (data not shown). These results indicated that the triple CD3/CD28/LFA-1 stimulation induced overall phosphorylation of multiple proteins (kinases and transcription factors) to a greater extent than CD3/CD28 stimulation.

T cell activation requires cell cycle entry and subsequent cell proliferation, which is an important downstream outcome of cellular activity induced by signal transduction pathways. Cellular proliferation and concomitant cell cycle initiation was measured by flow cytometry staining for DNA content and Ki-67 proliferation antigen. Figure 7C demonstrates cell cycle entry after 12 hours in CD3/CD28/LFA-1 stimulus as noted by >2n DNA content and expression of nuclear antigen Ki-67. Proliferation associated antigen Ki-67 and total DNA content quantify the cells residing within different cell cycle phases (Endl et al., 2001). As shown in Figure 7C, CD3/CD28/LFA-1 stimulation invokes far greater percentages of cells in the S and G2/M phase than CD3/CD28 stimulation. Thus, a more profound cell cycle entry is observed in CD3/CD28/LFA-1 vs. CD3/CD28 stimulated cells.

Simultaneous assessment of immunophenotype, intracellular signaling, and cellular function in single cells

We were interested in determining the contribution LFA-1 signaling possessed for naïve T cell activation in conjunction with CD3/CD28 co-stimulation. As evidenced below, we multiplexed magnetically activated cell sorting, 13-dimensional multiparameter flow cytometry (surface immunophenotyping, intracellular production of active kinases and cytokines) with profiling of secreted cytokines (cytometric bead arrays) and nuclear localized transcription factor profiling to obtain a multidimensional assessment of T cell activation and effector cell function. Figure 8 represents a flow diagram of the various techniques multiplexed for functional analysis and these data sets were cross-referenced for correlatives of cell surface receptor activation, intracellular signaling, and functional outcomes.

Enrichment of primary naïve T cells was performed by magnetically activated cell sorting (MACS negative depletion) using markers CD8, CD19, CD16, CD14, HLA-DR, and CD44 to remove cytotoxic T cells, B cells, NK cells, macrophages, and activated T cells from PBMC respectively (Figure 8). We utilized 8 markers to identify naïve human T cells (CD3⁺CD4⁺CD8⁻CD45RA⁺CD62L⁺CD11a^{dim}CD27⁺CD28⁺). CD3, CD28, and LFA-1 single and combined stimulation was performed on enriched non-activated T cells. Mature naïve T cells, as determined by 8 immunophenotypical markers, were analyzed for CD69, CD25, and IL-2 expression, simultaneously by 13-dimensional flow cytometry as expression of these markers defines an activated T cell (Yu et al., 2000; Zola, 2000). Flow cytometric kinetic experiments identified signaling pathways as tested for individual stimulation to peak and subside within 0.5-4 hours post stimulation and were unique to each phospho-kinase tested (data not shown). We analyzed samples at 8, 12, and 24 hours post stimulation to observe the phospho-levels of stimulated cells. Post 8 hour stimulations, CD3 stimulation alone resulted in minor CD69 activation and low levels of phospho-p44/42 (Figure 8, panel I). CD3/CD28 co-stimulation induced more CD69 activation, but the levels of phospho-p44/42 MAPK returned back to a basal level (Figure 8, panel II). However, optimal activation of naïve T cells (CD3⁺CD4⁺CD8⁻CD45RA⁺CD62L⁺CD11a^{dim}CD27⁺CD28⁺) was observed in the presence of LFA-1 plus CD3/CD28 stimulation at 8 hours (Figure 8, panel III). sICAM-2 gave rise to some phospho-p44/42 induction and intermediate CD69 induction whereas CD28 stimulation alone was incapable of inducing either marker to express (data not shown, *Reviewers please see Supplementary Figure 7A*). At this timepoint, CD25 and IL-2 expression was observed for the CD3/CD28 stimulation, but the the levels were greater in the CD3/CD28/LFA-1 stimulated cells (Figure 8, panel III). Thus, these results suggested a signaling threshold was a consequence of 3 surface mediated signal integration for CD3, CD28, and LFA-1.

We performed titration and kinetic analyses for stimulus induced T cell activation as measured by IL-2 intracellular production and CD25 and CD69 surface expression by flow cytometry. The mean fluorescent intensities (MFI) of the intracellular IL-2 detection were computed, and the log ratio of the $MFI_{stimulated}$ to the $MFI_{unstimulated}$ was calculated and plotted as a function of the concentrations of the titrations for the stimulants used (both for single and the various ratios for the combined stimulants (Figure 9)). A color intensity gradient was applied to the range of values (red being high, blue being low) to visually plot the differences (Figure 9). At 6 hour stimulation, titration of LFA-1 and CD3/CD28 stimulation increased production of intracellular IL-2, depicting a threshold for IL-2 production at 0.1 μ g/ml of a 1:1:1 ratio of CD3/CD28/LFA-1 stimulation (Figure 9A, right panel). Similar levels of CD3 and CD28 titrations showed lower levels of detected intracellular IL-2 (Figure 9A, left panel). Higher levels than 0.1 μ g/ml of CD3/CD28/LFA-1 demonstrated a linear response in concentration of stimulus to that of IL-2 production (Figure 9A). Levels of inducer as high as 10 μ g/ml of CD3/CD28 did not produce the same intracellular levels of IL-2 after 6 hr CD3/CD28/LFA-1 stimulation (Figure 9A). Varying the ratio of either CD3 or CD28 up to 10,000 fold higher in the CD3/CD28/LFA-1 combination did not result in increased levels of intracellular IL-2 (data not shown). Therefore, ratiometric titration analyses indicates that the LFA-1 signal contribution cannot be compensated by increased CD3 or CD28 stimulation.

At 6 hr stimulation, 10,000 fold less CD3/CD28/LFA-1 stimulus was necessary than in CD3/CD28 costimulation for similar expression of CD69 and CD25 (Figure 9B). After 6 hrs of CD3/CD28/LFA-1 stimulation we observed detectable secreted IL-2, contrasting the 12 hrs necessary for first detection of CD3/CD28 mediated IL-2 secretion (data not shown). At 12 hrs post stimulation, extracellular IL-2 after CD3/CD28/LFA-1 stimulation was reflected by significant production of intracellular IL-2 and correlated with enhanced CD69 expression (Figure 9C). Furthermore, CD25 expression is maintained in the CD3/CD28/LFA-1 stimulated T cells at 48 hours, contrasting to a decrease observed in the CD3/CD28 stimulated cell population (data not shown, Reviewers please see *Supplementary Figure 7B*). Thus, the LFA-1 stimulus positively contributes to overcome the threshold of IL-2 production, allowing T cells to become activated more proficiently and with considerably enhanced longevity.

Stimulus through LFA-1 contributes to transcription factor nuclear translocation and activity

Transcription factor nuclear localization and activation profiling was performed in single CD3, CD28 and LFA-1 stimulations, as well as CD3/CD28 vs. CD3/CD28/LFA-1 stimulation.

Luciferase reporter constructs were used to verify NFAT, NF- κ B, and AP-1 activities. CD3/CD28/LFA-1 stimulus presented higher fold inductions of NFAT, NF- κ B, and AP-1 transcriptional activity than CD3 or CD3/CD28 stimulation (Figure 10A). Nuclear extracts of stimulated naïve CD4 T cells were bound to 96-well plates spotted with consensus sequences to the transcription factors NF- κ B p65, NF- κ B p50, cFos, CREB, *akt2*, and cRel. Detection of bound transcription factors was performed by standard ELISA techniques. Stimulation to CD3, CD28, and LFA-1 induced distinct profiles of nuclear localized transcription factors (data not shown, Reviewers see Supplementary Figure 8A). CD3/CD28 stimulus induced nuclear localization of all transcription factors tested here, with somewhat higher levels detected for the CD3/CD28/LFA-1 stimulus (Figure 5B). The presence of the LFA-1 stimulus was slightly additive with CD3/CD28 in enhancing the levels of nuclear localized transcription factors. Titration experiments of NFAT activity verified that the LFA-1 signaling cannot be compensated for by increased concentrations of CD3/CD28 (Figure 10C), results consistent with the expression of IL-2 (see Figure 9A). Thus, the addition of the LFA-1 signaling integrates with that of CD3 and CD28 co-stimulation to enhance gene expression of at least the T cell transcription activities tested here.

LFA-1 stimulation in the presence of CD3/CD28 polarizes T cell differentiation towards a TH1 phenotype

We performed titration experiments of both CD3 vs. CD28 and CD3/CD28 vs. LFA-1 stimulation and monitored intracellular production of IL-4 and IFN γ . Activated T cells, differentiate to either TH1 or TH2 effector T cells and can be preferentially directed towards one pathway by cytokines or additional stimuli (Asnagli and Murphy, 2001; Moser and Murphy, 2000). Effector cells can be identified by their cytokine secretion profile. TH1 cells are characterized by high IFN γ and low IL-4, whereas TH2 cells are characterized by high IL-4 and low IFN γ (Murphy et al., 2000). Computing the ratio of the median fluorescence of IL-4/IFN γ demonstrates that the LFA-1 stimulus, at all concentrations tested, favors a low IL-4/IFN γ ratio (Figure 11). This is consistent with the TH1 phenotype. In the absence of the LFA-1 stimulus, CD3 and CD28 titrations favored a TH2 phenotype.

We used cytometric bead arrays to assess the secretion of IFN γ , IL-10, IL-2, IL-4, IL-5, and TNF α in naïve T cells after 24 hours stimulation. The combination of CD3/CD28 presented high levels of IL-4, and low levels of IL-5, and IL-10. This contrasted CD3/CD28/LFA-1 stimulation which displayed similar levels of IFN γ , but significantly lower levels of IL-4 (Figure 12). The

triple stimulation reported elevated levels of IL-5 and IL-10, with similar levels of IL-2 at 24 hours (Figure 12). Neither stimulation reported elevated levels of TNF α (Figure 12) and these effects were not observed in single stimulated cells (data not shown, Reviewers see Supplementary Figure 8B). Thus, the stimulation by LFA-1 polarizes T cell differentiation toward a TH1 phenotype in the presence of CD3/CD28 stimulation.

Discussion

The work presented demonstrates that the ICAM-2/LFA-1 interaction contributes a distinct signal to CD3/CD28 co-stimulation and leads to rapid and optimal naïve T cell activation. Furthermore, it demonstrates that this accessory stimulation can, together with CD3 and CD28 and in the absence of additional signals, polarize T cell differentiation towards an apparent TH1 phenotype. Although it is expected that T cell signaling integrates numerous additional signals in the native environment, this finding suggests strongly that ICAM-2 induced LFA-1 signaling can be an important determinant of phenotypic outcomes in naïve T cell maturation.

Initiation of T cell activation requires cell-to-cell contact, an adhesion process primarily mediated by Leukocyte Function-Antigen 1 (LFA-1). Adhesive events involved in T cell contact have largely been unrecognized as transducing signals that may enhance T cell functions. However, there is increasing evidence for the importance of adhesion mediated events in optimal T cell activation (Neeson et al., 2000; Somersalo et al., 1995; Wulfig et al., 1998).

Model systems of T cell stimulation using insect or fibroblast transfected cells report ICAM/LFA-1 interactions to be necessary for T proliferation (Damle et al., 1992a; Damle et al., 1993; Damle et al., 1992b; Damle et al., 1992c; Deeths and Mescher, 1999). We utilized a solubilized form of a normally surface-bound ligand of LFA-1, ICAM-2, to study the contribution of LFA-1 signaling to naïve T cell activation as ICAM-2 was able to mediate the active form of LFA-1 and induce a calcium influx (characterized in Perez and Nolan manuscript submitted, *reviewers please see manuscript in Appendix*). We report here that two LFA-1 binding proteins, JAB1 and cytohesin-1, are released, and subsequently phosphorylated upon ICAM-2 binding. Utilizing intracellular delivery of specific antibodies/peptides towards JAB1 and cytohesin-1, these LFA-1 interacting proteins mapped to distinct signaling pathways upon stimulation.

Cytohesin-1 is a guanine exchange factor that has recently been identified to be phosphorylated by PKC δ in vitro (Dierks et al., 2001) and enhances LFA-1 mediated leukocyte adhesion in T cell lines (Weber et al., 2001). Recently, a study identified the human herpesvirus 8 protein kaposin A as capable of mediating signaling by recruitment of cytohesin-1 and that a cytohesin-1 mutant (defective in guanine nucleotide exchange) was defective in p44/42 MAPK activation (Kliche et al., 2001). Interestingly, antibodies and a peptide directed to cytohesin-1's C-terminal

(SEC7 domain), which contain the PKC consensus sites to threonine, blocked cytohesin-1 signaling to p44/42 MAPK. JAB1 another LFA-1 integrin binding protein, contains PKC serine consensus sites by sequence analysis, and has been identified as a mediator of cJun phosphorylation and AP-1 activity in cell culture models (Bianchi et al., 2000). Therefore these studies corroborate the direct observation that JAB1 and cytohesin-1 are involved in signaling upon LFA-1 stimulation in naïve CD4 T cells.

Multidimensional analysis of LFA-1 signaling events, in conjunction with CD3/CD28 stimulation, indicated that T cell activation was optimal upon triple stimulation. Differential phospho-protein analysis suggested that LFA-1 signaling induced phosphorylation of many signaling events. The integration of these signaling events mediated by LFA-1 in addition to CD3/CD28 costimulation is exemplified by the enhanced NFAT, NF- κ B, and AP-1 activities. The presence of LFA-1 signaling lowered the threshold for IL-2, CD25, and CD69 expression (see Figure 9) and initiated cell cycle progressions. Furthermore, the levels of nuclear translocated transcription factors as tested were greater in the CD3/CD28/LFA-1 stimulus vs. CD3/CD28, demonstrating the result of multiple kinase cascades. However the global analysis of transcriptional activity is expected to yield insights into the consequences of signal integration for factors not tested here as phosphorylation of other transcription factors, namely cJun, Creb, Rsk, JNK, p44/42, and elk were also observed (see Figure 7A).

Consequences of the LFA-1 integrin signaling included kinetically enhanced naïve T cell activation, lowering of the stimulus threshold for T cell activation, and apparent polarization of T cells to favor a TH1 phenotype, as determined by low IL-4 and high IFN γ levels. Of significance, T cell lines expressing a constitutively active LFA-1 respond to sub-threshold stimuli (Kaplan et al., 2000)(Damle et al., 1993) and blocking LFA-1/ICAM interactions has been demonstrated to favor TH2 cytokine production (Salomon and Bluestone, 1998). Therefore these results reconcile with the evidence provided here that LFA-1 plays an active role in T cell activation in addition to its adhesive properties. Based on these observations, it is predicted that LFA-1 knockout mice, will not only have impaired immune responses, but will be deficient in the ability to surmount an inflammatory response or recover from infections necessitating a TH1 response for clearance. Studies utilizing CD18 knockout mice, the β 2 integrin of LFA-1, do indeed show impaired responses in a delayed-type hypersensitivity (DTH) inflammation model (Grabbe et al., 2002), as well as a defect in the induction of peripheral immune responses (Scharffetter-Kochanek et al., 1998). Thus, LFA-1 is important for *in vivo* TH1 responses.

In conclusion, LFA-1 contributes significantly to naïve T cell functionality as demonstrated by multiple independent assays. Most importantly, it greatly potentiates CD3/CD28 costimulation allowing for T cell maturation (as measured by CD69/CD25 expression, IL-2 secretion and progression to the S+G2/M phases of the cell cycle. Under the defined circumstances it also appears to contribute to commitment decisions as measured by the enhanced expression of TH1 cytokines. Underlying these outcomes we correlated these results with stimulation of multiple transcription factors and kinases, such as p44/p42, known to play roles in expression of these genes. Inhibition of LFA-1 by small molecules has also shown efficacy in various inflammation murine models although the molecular mechanisms for these observations remain unknown. The small molecule LFA-1 antagonist BIX 642, inhibited the DTH response in a trans-vivo DTH model (Winqvist et al., 2001). Recently, an oral LFA-1 inhibitor LFA703 suppressed the inflammatory response in a murine model of peritonitis (Weitz-Schmidt et al., 2001). Furthermore, Leukocyte Adhesion Deficient (LAD) patients, a disease in which CD18 is either mutated or missing, display severe and recurrent bacterial infections (Hogg et al., 1999; Lipnick et al., 1996), in addition to LAD afflicted children having increased risk for infection of pseudomonas aeruginos (Pollard et al., 2001), an infection typically cleared by a TH1 response (Fruh et al., 1995; Moser et al., 2002). Therefore, the clinical manifestations reported in both LAD patients and LFA-1 knockout mice can validate the physiological significance of LFA-1's signaling role demonstrated here in T cell activation and effector cell differentiation. We expect, given our current studies, that LFA-1 signaling will be further implicated in important immune cell regulatory events.

Materials and Methods

Immunological and Chemical Reagents

The following phospho-antibodies (p-Ab) were obtained from Cell Signaling Technologies (CST): p-Raf1 (Ser259), p-MEK1/2 (Ser217/221), p-p44/42 MAP kinase (Thr202/Tyr204), p-p44/42 MAPK (Thr202/Tyr204) mAb, p-Elk-1 (Ser383), p-p38 MAP kinase (Thr180/Tyr182), p-cJun (S63), non-phospho specific antibodies to the above proteins were also obtained from CST. Anti-Ki-67 mAb (Transduction Laboratories). All surface antibodies, cytokine antibodies, and isotype controls were obtained from PharMingen: CD3, CD4, CD8, CD69, CD25, IL-2, IL-4, IFN γ on FITC/PE/PerCp/APC. Alexa-Fluor dyes (350, 430, 488, 532, 546, 568, 594, 633, 660, 680), Cascade Blue, Cascade Yellow and R-phycoerythrin (PE) were purchased from Molecular Probes (Eugene, Oregon). FITC and Texas Red were purchased from Pierce (Rockford, Illinois). Cy5, Cy5.5, Cy7 were obtained from Amersham Life Science (Pittsburgh, Pennsylvania). Tandem conjugate protocols for Cy5PE, Cy5.5PE, Cy7PE, Cy5.5APC, Cy7APC can be found at www.drmr.com/abcon. Antibodies to CD62L, CD28, CD27, CD45RA, CD11a,

- CD3, CD4, CD8, IL-2, IL-4, IFN γ (PharMingen) were conjugated to FITC, PE, Cy5PE, Cy5.5PE, Cy7PE, Cy5.5APC, Cy7APC, cascade blue, cascade yellow, or alexa fluor dyes as needed. Quantitation of fluorescent probe conjugation was assessed to determine degree of labeling and protocols including dye spectral properties can be found at www.metazoa.com/UPL3419.
- 5 In general, alexa conjugates used had 2-12 moles of fluorescent dye per mole of protein. The following antibodies were obtained from varying vendors: CD102-FITC (Research Diagnostics), anti-rabbit IgG HRP (CST), anti-mouse/rabbit Alexa 488/568/633 (Molecular Probes). Anti-JAB1, anti-cytohesin-1 C-terminal cytohesin-1 peptide, and anti-mouse IgG HRP (Santa Cruz Biotechnologies). Protein and chemical reagents used: DMSO, phorbol myristelated acetate
- 10 (PMA) ionomycin, phytohemmagglutinin A (PHA), propidium iodide (PI) RNASE (Sigma). U0126, PD98059, SB600125 (Calbiochem). Anti-CD3 (OKT3, NA/EN) and anti-CD28 mAb (NA/NE, PharMingen). ICAM-2-FC was purchased from Genzyme.

Cell Culture and Primary Cell Isolation

- 15 Jurkat T-cells were maintained in RPMI-1640, 10% FCS, 1% PSQ. Cells were maintained at 5% CO $_2$ /37°C humidified incubator and serum starved 12 hours for phospho analysis. For preparation of primary cells, mononuclear cells were isolated from blood of healthy donors (Stanford Blood Bank, Stanford, California). Human peripheral blood monocytes were obtained by Ficoll-plaque density centrifugation (Amersham Pharmacia, Uppsala, Sweden) of whole
- 20 blood and depletion of adherent cells by adherence to plastic culture dishes. Isolated cells were maintained in complete media. Magnetically activated cell sorting was to enrich for naïve CD4 T cells by negative isolation using combinations of CD16, CD14, CD44, HLA-DR, CD8 or CD19 biotinylated antibodies (DynaI, Oslo, Norway) and streptavidin-magnetic beads (DynaI, Oslo, Norway).

25 Immunoprecipitations and Immunoblotting

- Cell extracts were prepared by washing 2x10 6 cells (treated as indicated) in ice cold PBS and harvesting in lysis buffer (20 mM Tris pH 7.5, 150 mM NaCl 1 mM EDTA 1 mM EGTA, 1% Triton X-100, 2.5 mM Na $_2$ PO $_4$, 1 mM β -glycerolphosphate, 1 mM Na $_3$ VO $_4$, 1 μ g/ml Leupeptin, 1 mM PMSF, protease inhibitor cocktail tablet (Boehringer Mannheim). Extracts were centrifuged
- 30 14,000 RPM (5 min, 4°C) and 10 μ g (BCA protein assay (Pierce)) were immunoblotted using standard procedures. Immunoprecipitations (IP) were pre-cleared with protein A/G plus-agarose beads, incubated with primary ab (1h), protein A/G plus-agarose beads (1h) and

washed 4X with lysis buffer. Blots were incubated with the indicated antibodies and developed using ECL (Amersham).

Flow Cytometry and FACS Analysis

5 Intracellular and extracellular staining was performed as described for phospho-staining (Perez and Nolan, 2002). Approximately $1-10 \times 10^7$ peripheral blood mononuclear cells (unactivated CD4+ T cells) were obtained by magnetically activated cell sorting (negative isolation), treated with a stimulatory condition for indicated time, and prepared for flow cytometry. Brefeldin A ($10 \mu\text{M}$) was added for intracellular cytokine detection for 6 hours (or as indicated). Isotype control match antibodies were used for stain controls. Kinetic analyses was done on synchronized 96-
10 wells and directly fixed as described (Perez and Nolan, Submitted). Four or less color flow cytometry data acquisition was collected on a FACSCalibur machine (Beckton Dickinson, San Jose California.) using CELLQuest software, analyzed and presented using FlowJo software (Tree Star, San Carlos, California). Data for four or less colors are representative of 3 independent experiments of 10^6 cells/sample and 50,000 events were collected and manually
15 calibrated. 11-color data acquisition was collected on a modified FACStarPlus (Becton Dickinson, San Jose, California) connected to MoFlo electronics (Cytomation, Fort Collins, Colorado). Data were collected by FACS-Desk software and compensated, analyzed, and presented using FlowJo software. Data for 11-color flow cytometry collected a minimum of 200,000 events. Cytometric bead arrays (CBA) were from PharMingen. Ratios of intracellular
20 flow cytometric measurements were calculated from the following equation where MFI equals mean fluorescent intensity: $\text{Log} [(MFI_{\text{experimental}} - MFI_{\text{isotype mAb}}) / (MFI_{\text{control}} - MFI_{\text{isotype mAb}})]$. Values were then designated a color intensity of two chosen colors to depict high and low, and displayed where appropriate.

Phospho-protein arrays

25 Protein A coated 96 well plates were coated with phospho and non-phospho specific antibodies ($1 \mu\text{g/ml}$) for 1 hr. Plates were washed twice with PBS, and antibodies were covalently coupled with disuccinimidyl succinate as suggested by manufacturer (Pierce). Plates were washed three times (PBS) and blocked with 4% BSA (in PBS, 30 min). Protein samples from stimulated CD4+ naïve T cells were quantified and equivalent amounts conjugated to either succinimidyl
30 ester reactive Alexa 532 and 680 dyes in 0.1 M pH8.0 sodium bicarbonate. After 1 hr the reactions were quenched ($100 \mu\text{L}$ 1M Tris pH 8.0) and washed in centricon YM-10 spin columns. Labeled protein samples were resuspended in PBST (plus 1% BSA). Plates were hybridized with $30 \mu\text{g}$ of Alexa532 and Alexa680 conjugated protein samples for 1 hr, 25°C . Plates were washed three times in PBST (PBS, 0.1% Tween-20) and three times in PBS.

Fluorescent intensities were detected using a Molecular Devices Gemini Xs machine and fold induction was computed as the log ratio of the mean fluorescent intensities (MFI) of CD3/CD28/LFA-1 to CD3/CD28 stimulation, calculated from the following equation: $\text{Log} [(MFI_{CD3/CD28/LFA}) / (MFI_{CD3/CD28})]$.

5 Nuclear Transcription Factor profiling

Mercury TransFactor kits (Clontech) spotted for consensus sequences for NF- κ B p50, NF- κ B p65, c-Rel, c-Fos, CREB, and ATF2 were used to hybridize transcription factors from nuclear extracts of 1×10^7 stimulated CD4⁺ naïve T cells (prepared as suggested by manufacturer). Detection of bound protein to DNA complex was made by standard ELISA methods.

10 Laser scanning confocal microscopy

Cells were treated as indicated and adhered to poly-L-lysine (Sigma) coated sterilized coverslips (10 mg/ml, 30 minutes) by mild centrifugation (1000 RPM, 10 minutes), washed twice in phosphate buffered saline pH 7.4 (PBS) and fixed in 2.7% paraformaldehyde (in PBS). Cells were stained as described for flow cytometry: (primary at 0.1 mg/ml, secondary at 1:1000 dilution, in 4% FCS, with several washing steps in between). Stained coverslips were mounted onto glass slides with Prolong Antifade reagent (Molecular Probes) and visualized using a Zeiss laser scanning confocal microscope 510. Images were acquired using Zeiss LSM510 software and compiled using Adobe Photoshop 6.0.

Intracellular delivery of antibodies/peptides

20 Antibodies were buffer exchanged into PBS by spin dialysis (centricon YM-100 spin columns) and mixed in 1:20 ratio of Eugene (Roche) for 30 min. Liposome/antibody complex was mixed with 1×10^5 cells in 50 μ L, serum free media for 1 hr. Cells were washed twice (RPMI/4%FCS) and used for cellular assays within 2-4 hrs. Transfection efficiencies using FACS analysis of IgG-FITC was 30-40%.

25 Reporter Gene assays

Jurkat cells were transfected with 5 μ g of NF- κ B driven luciferase reporter gene (gift of S. Kinoshita, Stanford University) or AP-1 driven luciferase reporter gene (gift of J.F. Fortin, Stanford University) using Eugene 6 reagent (Boehringer Mannheim). NFAT-luciferase transfected cells were a gift of J.F. Fortin. Twenty-four hours after transfection, cells were

stimulated as indicated for 6 hr at 37°C. Luciferase activity was analyzed in cell lysates of 1×10^4 cells (Promega).

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Example 3

ICAM-2/LFA-1 activates p44/42 MAPK through PYK2 and SYK

In this Example, using the methods and compositions of the present invention, the present inventors (also referred to herein as "we") demonstrate that Leukocyte Function Antigen-1- (LFA-1) was found to activate the RAS/RAF/MEK/MAPK cascade upon engagement with ICAM-2. Dissection of the signaling pathway in revealed the ICAM-2/LFA-1 interaction depended on both PYK2 and SYK tyrosine kinases. Both PYK2 and SYK were found to associate with LFA-1 only after interaction with ICAM-2 and were rapidly re-distributed to the plasma membrane with concomitant phosphorylation of PYK2 and SYK as revealed by biochemical analysis and confocal microscopy. Chemical genetic approaches identify an LFA-1 mediated signal to phosphorylate PYK2, with subsequent signal relay to SYK to be contingent on PCK and PLC γ activities. Furthermore, cell-to-cell contact of ICAM-2+ cells with LFA-1+ initiated the transactivation of the p44/42 MAPK pathway in LFA-1 expressing cells and simultaneously activated the AKT cell survival pathway ICAM-2 presenting cells, as determined by single cell kinase profiling by multiparameter flow cytometry. These results highlight that specific trans-signaling events occur upon cellular adhesion events mediated by the interaction of ICAM-2 and LFA-1, implicating the importance of ICAM interactions in consequential processes such as cellular adhesion and T cell activation.

Introduction

Leukocyte mediated functions are largely governed by their ability to localize to appropriate tissue compartments via adhesion molecule recognition of ligands on target tissues. Targeting of leukocytes to distinct cellular environments is of pivotal importance in cellular processes such as blood clot formation (Bowes et al., 1995; Nagaoka et al., 2000; Schleef et al., 2001), immune surveillance (Patarroyo and Makgoba, 1989; Plate et al., 2000), inflammatory responses (Chihara et al., 2000; Cid et al., 2000; Krull et al., 1999; Zhang et al., 1999),

angiogenesis (Griffioen et al., 1998; Melder et al., 1996; Patey et al., 1996; Radisavljevic et al., 2000; Verkarre et al., 1999), and cellular migration (All et al., 2000; Sans et al., 2001).

Leukocyte targeting and adhesion is primarily mediated by the α and β integrin subunits and their intercellular adhesion molecule (ICAMs) -1, -2, -3, -4, -5 ligands (Douglas et al., 2000; Kruger et al., 2001; Sixt et al., 2001; Tian et al., 2000; van den Berg et al., 2001; Wang and Springer, 1998). The α and β integrin subunits form heterodimeric ICAM receptors such as leukocyte function antigen-1 (LFA-1, CD11a/CD18, or $\alpha_L\beta_2$), MAC-1 (macrophage receptor 1, CD11b/CD18, or $\alpha_M\beta_2$), CD11c/CD18 ($\alpha_X\beta_2$ or p150,95) and recently identified CD11d/CD18 ($\alpha_D\beta_2$) (Binnerts and van Kooyk, 1999; de Fougerolles et al., 1995; Gahmberg et al., 1998; Hayflick et al., 1998; Hermand et al., 2000). The ICAMs are members of the immunoglobulin superfamily, a subclass of the adhesion molecules that include cadherins, integrins, and selecting. ICAMs show differences in their cell type distribution- which is believed to reflect their varying functions. In addition, they can be differentially induced by various inflammatory cytokines (Hayflick et al., 1998; Simmons, 1995; Somersalo, 1996). ICAM molecules are expressed on lymphoid lineages that include B cells, T-cells, red blood cells, macrophages (Hayflick et al., 1998), and also expressed on a subset of vascular endothelial cells (Griffioen et al., 1996) and neurons (Tian et al., 2000).

Deficiencies or augmentation of cellular adhesion properties are both detrimental and life-threatening, as exemplified in a variety of immunopathologies such as leukocyte adhesion deficiency type I (LAD I, allergic encephalomyelitis, tumor vasculature, vascular permeability diseases, hypertension, severe sepsis syndrome/septic shock, tumor metastasis, and osteoporosis among others (Allende et al., 2000; Angelopoulou et al., 1999; Collins et al., 2000; Etzioni and Tonetti, 2000; Ghaisas et al., 2000; Kiarash et al., 2001; Lalancette et al., 2000; Schnitzler et al., 1999; Tanaka et al., 2000; Weigand et al., 1999). Although extensive work has been conducted on leukocyte cell surface molecules to correlate mutations in, or absence of, the integrin receptors to the aforementioned immunopathologies it is not clear to what extent the ICAMs contribute to the progression of these diseases (Craig et al., 2000; Daniel et al., 2000; Koga et al., 2000; Mori et al., 2000; Muanza et al., 1999; Oishi et al., 2000). There is increasing evidence that elucidation of mechanisms involved in ICAM mediated cell-to-cell contact (Gottlieb et al., 2000) and in mechanisms that underscore the irregular production of soluble ICAM molecules may be of therapeutic interest (De Vita et al., 1998; Guo et al., 2000; Salmaggi et al., 1999). Blockade of LFA-1/ICAM interaction by peptides and monoclonal antibodies is proving to be effective in inhibiting processes such as T cell activation (Tibbetts et al., 2000), attenuation of organ transplant rejection and various adhesion related diseases

(Gottlieb et al., 2000; Storck et al., 1994; Sun et al., 2000). Thus, ICAM molecules and their ligands are being recognized as important immunomodulators.

We have demonstrated that ICAM-2 can illicit a pro-survival signal upon surface clustering on human B cells rendering protection against FAS and TNF α induced apoptosis. Similarly, ICAM-1 has been shown to transmit an intracellular signal upon interaction with the extracellular matrix (Pluskota and D'Souza, 2000) and mice deficient in ICAM receptors or ICAM molecules have irregularities in a functional immune system (Ding et al., 1999; Gerwin et al., 1999; Igietsme et al., 1999; Schneeberger et al., 2000; van Den Engel et al., 2000). Crystallographic, molecular, and mutational analysis of ICAMs (1-4) and LFA-1 suggest that overlapping but not identical binding regions exist (Casasnovas et al., 1998; Casasnovas et al., 1999; Casasnovas et al., 1997; Edwards et al., 1998; Kotovuori et al., 1999[Hermand, 2000 #145]). In addition, dimerization requirements for ICAM-1 but not ICAM-2 or-3 binding to LFA-1 are indicative of differences in molecular interactions involving ICAM and LFA-1 and may allow differential cellular responses to distinct ICAM molecules (Casasnovas et al., 1998; Miller et al., 1995; Reilly et al., 1995). Finally, the repertoire of ICAM-binding integrin heterodimeric receptors and their subsequent signaling has not been investigated in depth.

Utilizing both retrovirally transduced NIH3T3 to express human ICAM-2 and a biochemically purified soluble ICAM-2, we demonstrate that in T cells, LFA-1 interaction with ICAM-2 induces activation of the RAS/RAF/MEK/ERK pathway (p44/42 Mitogen Activated Protein Kinase, MAPK). Monoclonal antibodies against LFA-1a (CD11a) or p2 integrin (CD18) blocked p44/42 MAPK activation. The signal was independent of ZAP-70, Src, FAK, and Lck as phosphorylation was not observed as a function of ICAM-2/LFA-1 interaction. ICAM-2/LFA-1 induced p44/42 MAPK activity was dependent on proline-rich tyrosine kinase 2 (PYK2) and spleen tyrosine kinase (SYK), members of the Src family non-receptor tyrosine kinases, and p44/42 activity was abrogated with specific pharmacological inhibitors piceatannol and tyrphostin A9 respectively. Confocal microscopy reveals that there is a re-distribution of both PYK2 and SYK to the cellular membrane upon LFA-1 engagement with ICAM-2 and that this interaction yields phosphorylation of PYK2. Furthermore, PYK2 and SYK co immunoprecipitated with LFA-1 only after engagement of ICAM-2, indicating a ligand induced conformational change was responsible for the interaction.

Development of a phospho-tyrosine protein elisa to screen for potential kinases involved in the LFA-1 to PYK2/SYK activation identify LFA-1 ligation to induce PYK2 phosphorylation, with subsequent SYK phosphorylation dependent on PCK and PLC γ . Biochemical purification of ICAM-2 binding proteins not only identified β 2 integrin, but also p1 and β 133 integrins as being

capable of interacting with ICAM-2, emphasizing that additional heterodimeric receptors involving p1 or p3 integrin subunits may exist for the ICAMs and that differential cellular responses can be regulated at the level of receptor subunit expression at the level of receptor avidity. Recently, other integrin heterodimeric receptors have been shown to influence the activity of p2-integrin receptors, illustrating that concerted cell adhesion might represent an additional level of intracellular signaling or regulation (May et al., 2000). Simultaneous measurement of intracellular kinase activities in mixed population experiments demonstrated that when ICAM-2 overexpressing fibroblasts engage LFA-1 expressing T-cells, activation of the AKT pathway is exclusively seen in the ICAM-2 expressing cells while activation of the p44/42 MAPK (ERK1/2) pathway is exclusively seen on the LFA-1 expressing T-cells. This trans-stimulatory effect of the ICAM-2/LFA-1 interaction is not dependent on additional co-stimulatory molecules, as ICAM2-/- fibroblasts did not result in either AKT or p44/42 MAPK activation. These results elucidate a previously unrecognized signaling mechanism for LFA-1 and have implications for its role in cellular adhesion and T cell activation.

Results

ICAM-2 clustering activates the RAF/MEK/ERK pathway

Previous work utilizing retrovirally transduced NIH3T3 to overexpress human ICAM-2 allowed for the dissection of an ICAM-2 induced signaling pathway that was dependent on PKB/AKT and mediated through the intracellular C-terminal. In this work, clustering of surface expressed ICAM-2 molecules by monoclonal antibodies revealed a distinct and separate activation of the RAF/MEK/ERK mitogen activated protein kinase pathway (p44/42 MAPK) that was not abrogated in the presence of PI3K inhibitors (data not shown). The extracellular portion of ICAM-2 was sufficient to induce MAPK activity as both an ICAM-2 C-terminal truncated molecule (ICAM2-AC) and full length ICAM-2 induced activation of p44/p42 MAPK when clustered on the cell surface. The ICAM2-DC molecule was not capable of transducing the PI3K/AKT intracellular signal characterized to ICAM-2 in a separate study. Expression of human ICAM-2 and ICAM2-AC was verified to target to the cellular membrane by flow cytometry (Figure 13A) and was confirmed by western blot analysis to present a similar electrophoretic mobility as the endogenous ICAM-2 protein in Jurkat cells (Figure 13B). NIH3T3 fibroblasts do not express levels of murine ICAM-3 but do express low levels of murine ICAM-1 and ICAM-2, however, retrovirally enforced ectopic expression of human ICAM-2 was found to be 5 magnitudes higher than endogenous levels as determined by quantitative flow cytometry.

ICAM-2 clustering initiated activation of the RAF/MEK/ERK cascade (p44/42 MAPK pathway) in NIH3T3 cells as a function of time as observed by immunoblotting for each respective kinase's active phosphorylated state. (Figure 13 C). ERK kinase activity led, as expected, to phosphorylation of Elk-1. Phosphorylation of Elk-1 has been demonstrated to be sufficient for its activation and nuclear translocation (Wang and Prywes, 2000). Incidentally, other p44/42 MAPK activated proteins capable of transcription activity, namely Rsk-1 (Figure 13 C) and CREB (data not shown), were also phosphorylated as a function ICAM-2 clustering. The ICAM-2 induced phosphorylation of p44/42 MAPK was verified by kinase activity assays and was present in both ICAM-2 and ICAM2-AC transduced fibroblasts, and by endogenous ICAM-2 in Jurkat T cells when crosslinked (Figure 13 D). Furthermore, activation of JNK, p38 MAPK or activities of MEK3/6, or MEK 4/7 were not observed following ICAM-2 crosslinking (data not shown). Verification of the truncated ICAM2-AC expression by intracellular flow cytometry (see Figure 13A) and its apparent inability to induce an intracellular signal led us to believe that the p44/42 MAPK activity observed was due by a surface-bound ICAM-2 interacting protein (discussed below).

ICAM-2 induced p44/42 MAPK activity attenuates growth kinetics and augments cellular contact inhibition

The p44/42 MAPK family is important in a variety of cellular process that govern growth, survival, and differentiation of cells (Chang and Karin, 2001). Several assays were used to assess p44/42 MAPK mediated events. Cell cycle analysis was performed to determine if the ICAM-2 induced p44/42 MAPK activity was able to initiate cell cycle re-entry. ICAM-2, ICAM2-ΔC, vector control, and varying cell types were serum starved for 72 hours to synchronize cells in the absence of chemical agents. Cells were then crosslinked for ICAM-2 using a monoclonal antibody and analyzed 18 hours later. The percentage of cells in S-phase increased following ICAM-2 crosslinking (Figure 14A). Cell cycle re-entry was dramatically apparent in ICAM2-AC (versus vector control and ICAM-2 cells). Continual stimulation of cell cycle re-entry is sufficient to override growth inhibition signals, often the case in cells that have lost the ability to arrest growth by contacting inhibition and exemplified in the metastatic spread of cancerous cells. Since ICAM-2 and ICAM2-AC presented p44/42 MAPK activity upon ICAM-2 clustering, we postulated that constitutive activation of the p44/42 MAPK pathway could potentially enhance growth kinetics, consequential to the cell cycle re-entry observed and may be sufficient to circumvent contact inhibitory mechanisms. We tested this by monitoring cellular respiration as a function of cell density to determine the extent of growth arrest induced, if any, by contact inhibition. We expected contact inhibition to induce a growth arrest in adherent cells, and therefore a decrease in cellular respiration as the cell density increased. Vector control cells (negative control) showed reduced cellular respiration at high cell densities

in contrast to both Jurkat cells (suspension cells not capable of contact inhibition) and 293T cells (transformed fibroblast that fail to contact inhibit) (Figure 14B). Both ICAM-2 and ICAM2-ΔC expressing cells exhibited enhanced cellular respiration at high cell densities over vector control (Figure 14B). Despite these observations, ICAM-2 and ICAM2-ΔC expressing cells were not found to be transformant, a property of cells that have lost contact inhibition, using a top agar transformation assay and did not form tumors when subcutaneously implanted in nude mice (data not shown). Irrespective of this, both ICAM-2 and ICAM2-ΔC expressing cells displayed enhanced growth kinetics over vector control cells (Figure 14C). Since the ICAM2-ΔC construct elicited p44/42 MAPK activity and was able to initiate a robust cell cycle re-entry in comparison to vector controls, this suggested that an ICAM-2 surface interacting protein was responsible for the activation of p44/42 MAPK. The observation that the p44/42 MAPK induction effect was intensified when ICAM-2 was relieved of C-terminal cytoskeleton interactions (Figure 14A) illustrates a potential role for cytoskeleton-mediated regulation of ICAM surface protein signal transduction (Heiska et al., 1998; Heiska et al., 1996; van Kooyk et al., 1999).

ICAM-2 interacts with β integrins 1, 2, and 3

A soluble form of ICAM-2 was used to verify the ability of ICAM-2 to bind to the cell surface. Full length ICAM-2 was immuno-depleted from ICAM-2 transduced cell lysates and conjugated to FITC (Figure 15A). The ICAM-2-FITC probe was used to stain the surface of NIH3T3 fibroblast cells, Jurkat T cells, and BaF3 proB cells (Figure 14B) on the presumption that ICAM-2 will bind, at least to its normal ligand partner, β 2 integrin on these cells. Pre-treatment of cells with the protease trypsin drastically decreased ICAM-2-FITC binding, suggesting that ICAM-2 was binding to a surface bound protein. The ICAM-2-FITC probe was found to be partially competed by antibodies to β 2-integrin on NIH3T3 fibroblasts, a cell line that lacks LFA-1 α expression (data not shown). ICAM-2-FITC binding was confirmed by epifluorescence microscopy and was titrateable and inhibited with an unlabeled control in a competition assay (data not shown). Since ICAM2-FITC binding was not completely inhibited by anti- β 2 integrin antibodies in NIH3T3, in addition to these cells lacking expression of LFA-1 α , it suggested that an additional molecule(s) capable of binding ICAM-2 existed on the surface of these cells.

A two-column affinity chromatography approach was utilized to determine surface bound target molecules capable of binding ICAM-2. ICAM-2, ICAM2-AC, and vector control cell surface proteins were biotinylated and subjected to affinity chromatography on an ICAM-2 coupled solid support, and subsequently passed through a streptavidin agarose support. A two-column

system was preferred in order to identify proteins that were both surface expressed (biotinylated) and ICAM-2 binding. Final protein elutions were tested for both biotinylation by immunoblotting techniques and -ICAM-2 interaction by the use of the ICAM-2FITC probe in a farwestern approach (Figure 15A). Protein bands of 112, 95, 67 kilodaltons (kDa) were gel eluted and sequenced by MALDI-MS. The biotinylation of the surface proteins interfered with a direct analysis of the proteins eluted, and fragments of peptide sequences generated suggested that the molecules contained RGD domains, with homologies to several surface bound receptors, although no apparent single protein could be identified from the peptide fragments generated. Therefore, we decided to test for integrin binding partners, given that ICAM-2 has been shown to bind β 2 integrin. Immunoblotting the final two-column elutions confirmed β 1, β 2, and β 3 integrins to be both surface expressed (biotinylated) and ICAM-2 interacting (Figure 15C). β 4, β 5, and β 6 integrins were not identified in immunoblotting final two-column elutions (data not shown). Performing reciprocal column systems (i.e. streptavidin-agarose support followed by ICAM-2 affinity column) in addition to, ICAM-2 immunoblots on β 1, β 2, and, β 3 integrin immunoprecipitations confirmed these results (data not shown). Additional integrins β 4, β 5, β 6, α 5, and α 1 were not found to coimmunoprecipitate with ICAM-2 (data not shown). Furthermore, expressed ICAM2 protein did not interact with the ICAM-2 coupled solid support, discrediting the possibility that ICAM-2 can dimerize, at least under the non-reducing conditions used here (data not shown).

Monoclonal antibody (mAb) blocking experiments were used to distinguish the contribution of the integrin binding partners to the ICAM-2 induced p44/42 MAPK activation. Pretreatment of ICAM-2, ICAM2-AC, and vector control cells with mAbs for β 1, β 2, and β 3 integrin prior to ICAM-2 clustering demonstrated occlusion of β 2 integrin to selectively inhibit ICAM-2 induced p44/42 MAPK activity (Figure 15C). Unexpected results were observed by using mAbs to β 1 and β 3, in that both partial inhibition and induction of p44/42 MAPK activity was observed (elaborated in the discussion). Therefore, differences observed in the antibody blocking experiments for β 1 and β 3 integrins cannot be differentiated between ICAM-2/integrin block or integrin-induced signaling given the nature of the experiment and were not explored any further. However, the experiments do illustrate that antibody blockade of β 2 integrin is sufficient to abrogate ICAM-2's induced p44/42 MAPK activity. Since ICAM-2 clustering via the extracellular domain induced p44/42 MAPK activity in the ICAM2-AC transduced cell population and was blocked by β 2 integrin monoclonal antibody, it suggested that the ICAM2-AC molecule was capable of interacting with β 2 integrin on the same cell. Thus, the surface interaction of these two molecules mediated p44/42 MAPK activity through the β 2 integrin. Interestingly, ICAM-2's natural receptor Leukocyte

Function-Antigen-1 (LFA-1) is an α , β heterodimeric receptor composed of both β 2 and α _L integrins (CD11 a) (Kaplan et al., 2000). Therefore, we decided to test the ICAM-2 induced p44/42 MAPK activation in Jurkat T-cells, which endogenously express the low levels of the LFA-1 receptor subunits, with a biochemically purified soluble ICAM-2 protein. Treatment of Jurkat cells with soluble ICAM-2 induced p44/42 MAPK phosphorylation and activity and was blocked by monoclonal antibodies to both β 2 and α _L integrins (Figure 15F) indicating that the ICAM-2/LFA-1 interaction can mediate activation of p44/42 MAPK in LFA-1 expressing cells.

ICAM-2/LFA-1 engagement induces p44/42 MAPK activity through PYK2 and SYK

To identify upstream kinases that would be responsible for signal transmission from LFA-1 to Raf (and subsequently to p44/42 MAPK), a series of kinase inhibitor screens were conducted. Tyrphostin A9 and piceatannol, specific inhibitors of proline-tyrosine kinase 2 (PYK2) and Spleen-tyrosine kinase (SYK) respectively (Avdi et al., 2001; Fuortes et al., 1999) were found to abrogate the ICAM-2 induced activation of Raf and subsequently p44/42 MAPK (Figure 16A). Other protein kinases that include p38 MAPK, and two members of the src related tyrosine kinases p56Lck and Src were not found to be involved by the inability of specific pharmacological inhibitors of these kinases to block ICAM-2 induced p44/42 MAPK activity. Furthermore, immunoblotting for phosphotyrosine residues of immunoprecipitated tyrosine kinases involved in T-cell activation, namely LAT and ZAP-70, and tyrosine kinases involved in integrin signaling, FAK and Src, did not implicate these kinases in the ICAM-2/LFA-1 mediated signal (data not shown). However the ICAM-2/LFA-1 activation of p44/42 MAPK was dependent on guanosine triphosphate, implying the involvement of Ras (data not shown).

It was apparent that p44/42 MAPK activation was a function of an ICAM2/LFA-1 interaction that was initiated when ICAM-2 engaged LFA-1. It has been reported that adhesion molecule receptors require an activation step in order for intracellular signaling to occur that usually results in changes in it's the receptor's avidity and/or affinity for its respective ligand or in its formation if composed of a heterodimeric association (van Kooyk et al., 1999). This prompted us to test whether contact of ICAM-2 with LFA-1 induced an "activation state" that allowed for the subsequent binding of PYK2 or SYK to transmit the signal through an SH2/GRB/Ras dependent pathway to p44/42 MAPK. Typical exogenous treatments with phorbol esters to activate LFA-1 could not be used, since we noted eventual activation of the p44/42 MAPK pathway following treatment (data not shown). Therefore, the interaction of PYK2 and SYK with both subunits of LFA-1, β 2 and α _L integrins was tested by co-immunoprecipitation as a function of soluble ICAM-2 and various T cell activating stimulants. Tyrosine phosphorylation of neither

$\beta 2$ nor α_L integrin was observed as a function of any stimulus tested (data not shown). An association between $\beta 2$ and α_L integrin was only observed after treatment with soluble ICAM-2 and was absent in unstimulated cells (Figure 16B). This suggested that the formation of the LFA-1 receptor can be initiated by a ligand induced event mediated by ligand induced recruitment of the α subunits. LFA-1 receptor integrity was found to be stabilized by treatment with various T cell stimulants in addition to soluble ICAM-2 resulting in an enhanced interaction between $\beta 2$ and α_L integrin.

SYKs interaction with α_L integrin was present under unstimulated conditions and was attenuated by stimulatory conditions with a concomitant association with $\beta 2$ integrin. This suggested that the interaction of $\beta 2$ and α_L integrins induced by ICAM-2 resulted in an activation of SYK - a process that can result from a conformational change proceeding LFA-1 formation as suggested by studies reporting avidity changes in ligand binding of LFA-1. PYK2 presented a weak association with $\beta 2$ integrin, an interaction that was enhanced upon ICAM-2 stimulus, and only interacted with α_L integrin after the ICAM-2 stimulus was provided (Figure 16B). Although the exact molecular details of the temporal kinetics of PYK2 and SYK association with LFA-1 as a function of stimulus were not investigated, the data would suggest that ICAM-2 could induce formation of its LFA-1 receptor by recruiting $\beta 2$ and α_L integrin. We speculate that "activated" LFA-1 induces a conformational change that is transmitted to PYK2 and SYK, either through direct physical contact, or exposure of binding sites that would in turn allow binding of PYK2 or SYK.

Ligand induced conformational changes of LFA-1 have been reported (Kotovuori et al., 1999; van Kooyk and Figdor, 2000), but lack linkage of LFA-1 to specific intracellular signaling mechanism. Furthermore, reciprocal coimmunoprecipitations of PYK2 and SYK association with $\beta 2$ integrin was observed to be maximal in the presence of one or more stimuli (Figure 16B). Both PYK2 and SYK2 were found to be phosphorylated in response to ICAM-2 as a function of time in LFA-1 expressing cells. Phosphorylation of PYK2 tyrosine residues tyr402 (autophosphorylation site) and tyr881 (SH2 binding site) was induced by ICAM-2 binding LFA-1 as determined by immunoblotting for respective phosphorylated residues. This confirmed that phosphorylation of PYK2 is mediated by an ICAM-2 induced LFA1 activation (Figure 16C).

Confocal laser scanning microscopy was used to assess the cellular distribution of PYK2 and SYK, and their respective localization and/or phosphorylation after treatment of Jurkat cells with soluble ICAM-2. A cellular redistribution of both PYK2 and SYK upon ICAM-2/LFA-1 interaction was observed as diffused cytoplasmic PYK2 and SYK localized to the plasma membrane (Figure 17, panel C and L). Phosphorylation of PYK2 was observed at the plasma

membrane only after the ICAM-2 stimulus was provided as detected by a phospho-specific antibody to PYK2 at tyrosine 402 (Figure 17, panel F and I). Recruitment of PYK2 to the plasma membrane by ICAM-2s interaction with LFA-1 resulted in PYK2 phosphorylation as PYK2 phosphorylation was not observed in the absence of the ICAM-2 stimulus. It is not clear which specific events mediate PYK2 or SYK recruitment to LFA-1 upon ligand interaction, at present, we hypothesize that a ligand induced conformational change to LFA-1 exposes protein binding sites for PYK2 and SYK interaction.

ICAM-2/LFA-1 interaction transmits distinct signals to both contacting cells

LFA-1 and ICAM- have been implicated in cellular adhesion of leukocytes, in essential processes that include lymphocyte trafficking (Etienne-Manneville et al., 2000; Sato et al., 2000; Sigal et al., 2000), stabilizing T-cell:antigen-presenting cell contact (Hwang et al., 2000; Neeson et al., 2000), metastatic spread (Dosquet et al., 1997; Griffioen et al., 1996; Griffioen et al., 1996; Regidor et al., 1998; Tang and Honn, 1994), and organ transplant rejection (Rentsch et al., 2000). Considering the severity of diseases associated with the absence of ICAM/LFA-1 interactions, we sought to understand the signaling mechanisms initiated upon cell-to-cell contact that utilize ICAM-2 and LFA-1. To do this, we developed specific kinase probes for AKT and p44/42 MAPK, among others, to simultaneously monitor kinase activity within individual cells. These probes detect phosphorylated AKTser473 and phosphorylated p44/42 MAPK (Thr202/Tyr204) by flow cytometry and phosphorylation at these residues was found to correlate with respective kinase activity by immunoblotting and kinase activity assays. These probes, in conjunction with specific cell type markers, allowed us to test for the activation of signaling pathways in heterogeneous cell mixing experiments by flow cytometry.

NIH3T3 cells were engineered by retroviral transduction to overexpress human ICAM-2 and were used as a presenting cell to Jurkat T cells. NIH3T3 lack expression of costimulatory molecules CD86 and CD80, and express low levels of murine ICAM1,-3 as determined by flow cytometry (data not shown). The engineered ICAM-2 overexpressing cells present 106 fold surface human ICAM-2 protein compared to vector control as determined by quantitative flow cytometry. ICAM-2 cells and vector control cells were mixed with Jurkat T cells and prepared for flow cytometry by surface staining for T cell markers CD3 and CD4, and intracellular detection of phosphorylated p44/42 and AKT. Figure 18 displays a FACS plot of both ICAM-2 (CD3 CD4-) and Jurkat cells (CD3+CD4+) as distinguished by CD4 and CD3 markers. Forward and scatter voltages were experimentally determined to allow visualization of both cellular populations denoting the autofluorescence of fibroblast cells. Intracellular kinase activities for

both AKT and p44/42 present two distinguishable peaks that can be attributed to distinct cell populations. Only single peaks were observed in homogenous control populations (data not shown). AKT^{high} activity is observed primarily in CD3/CD4 double negative cells while ERK^{high} activity is observed primarily in CD3/CD4 double positive cells. Therefore, ICAM-2/LFA-1 interaction between a presenting cell and a leukocyte activates AKT and p44/42 within each cell respectively.

Discussion

The results presented demonstrate (1) ICAM-2/LFA-1 interaction activates the p44/p42 MAPK cascade in LFA-1 expressing cells and can be blocked by antibodies to both α L and O2 integrins, (2) LFA-1 transmits a signal to PYK2 and SYK upon ligand binding, (3) ICAM-2/LFA-1 engagement activates AKT in ICAM2 expressing cells and p44/42 in LFA-1 expressing cells, a concerted transstimulation to contacting cells upon cell-to-cell contact respectively.

We have previously shown that ICAM-2 can intracellularly activate the PKB/AKT pathway protecting fibroblasts, T and B cells from apoptosis. Recently, it was shown that ICAM-1 interactions with fibrinogen can activate ERK1/2 and regulate endothelial cell survival (Pluskota and D'Souza, 2000). Therefore it is likely that ICAMs can transduce specific signals to their host cell upon engagement of LFA-1. Since the ICAMs are members of immunoglobulin superfamily, extensive work has been focused around their adhesive properties in cell-to-cell contact and lymphocyte recirculation (EtienneManneville et al., 2000; Geijtenbeek et al., 2000; Gerwin et al., 1999). This in part is due to the cell specific expression patterns exhibited by the ICAMs and their attenuation upon the presence of inflammatory cytokine (McLaughlin et al., 1999; Wolf et al., 2001). For example, ICAM-1 is expressed at low levels and is upregulated by inflammatory cytokines on leukocytes, endothelial cells, fibroblasts, and dendritic cells in contrast to ICAM-2 which is constitutively expressed on leukocytes, endothelial cells, and platelets (Staunton et al., 1989; Xu et al., 1996) and overexpressed in many lymphoproliferative diseases (Eichelmann et al., 1992; Molica et al., 1996).

Adhesion properties attributed to the ICAMs include cell migration (Ding et al., 1999; Nagaoka et al., 2000; Somersalo, 1996), lymphocyte trafficking (Devine et al., 1996; Geijtenbeek et al., 2000; Jutila, 1992), enhancing the avidity of the interaction between a T cell and antigen-presenting cell (APC) (Kotovuori et al., 1999; van Kooyk and Figdor, 2000; VanCompernelle et al., 2001; Wulfing et al., 1998) and contribute to the pathogenicity of viral infections (Belle and Rossmann, 1999; Hioe et al., 2001; Nagaoka et al., 2000; Shigeta, 1998). These adhesion roles partially overlap with the roles ascribed to the integrins, a distinct superfamily of $\alpha\beta$ heterodimeric transmembrane proteins that are essential in

adhesion/migration and proliferation/differentiation cellular processes (Geginat et al., 1999). It is now becoming clear that integrins themselves can function as signal transducers in integrin-mediated T cell co-stimulation (Gaglia et al., 2000; Goldstein et al., 2000), in interactions involving the extracellular matrix (ECM), in addition to migratory properties (Epler et al., 2000; Holly et al., 2000; Tarone et al., 2000). Signal transduction for the 19 superfamily has largely gone unrecognized, with the majority of attention focused on other cellular adhesion molecules that include the integrin, selectin, and cadherin family of proteins (Braga, 1999; Elangbam et al., 1997; Gonzalez-Amaro and Sanchez-Madrid, 1999). Furthermore, a subset of integrins more commonly known by their CD number as leukocyte antigens - $\beta 1$ (CD29), $\beta 2$ (CD18), $\beta 3$ (CD61) $\beta 4$ (CD 104) $\alpha 1-6$ (CD49a-f respectively), in addition to the ICAMs ICAM-1 (CD54), ICAM-2 (CD102), ICAM-3 (CD50) indirectly suggest that these molecules may possess specific signaling roles with functional consequences in lymphocyte populations.

Dissection of the downstream signaling pathways involving the LFA1/ICAM-2 interaction was found to be dependent on both PYK2 and SYK. PYK2 (also known as CAK-B or RAFTK) is a second member of the focal adhesion kinase family and activated by many types of cell surface receptors that include integrins, cytokines, immune receptors, stress stimuli, and PKC activation (Avraham et al., 2000; Hauck et al., 2000). Recently, phosphorylation of PYK2 has been associated with homotypic adhesion mediated by an LFA-1/ICAM-1 interaction in B cells (McDonald et al., 2000), and has also been implicated to be positively regulated by $\beta 1$ and $\beta 3$ integrin in myeloid cells (Litvak et al., 2000; Miura et al., 2000; van Seventer et al., 1998). Positive regulation of PYK2 by $\beta 2$ integrins has been reported in regards to cell spreading and migration in macrophages (Duong and Rodan, 2000) and to induce apoptosis in neutrophils (Avdi et al., 2001). In addition, activation of PYK2 has been stated to be downstream of SYK in Fc ϵ R1 signaling in basophils (Okazaki et al., 1997). Therefore PYK2 is recently being regarded as a key signaling molecule in transmitting adhesion mediated events intracellularly.

SYK is a spleen non-receptor tyrosine kinase that is essential in signal transmission of $\alpha III \beta 3$ inside out signaling (Saci et al., 2000), has roles in Fc ϵ R1 signaling (Jabril-Cuenod et al., 1996), and phosphorylated by integrin mediated signaling (Miller et al., 1999; Moores et al., 2000). Given the numerous reports depicting both PYK2 and SYK phosphorylation events, these events can be categorized by stimulatory conditions and present differential outcomes among different cell types: (1) signaling events involving β integrins with regards to B cell signaling (Babe et al., 2001; Li et al., 2001); (2) events involving $\beta 2$ integrins with regards to cellular migration and invasion (Holly et al., 2000); (3) activation induced by immune receptors CD2

and CD28 (Fukai et al., 2000; Tsuchida et al., 1999). Here we report that PYK2 and SYK2 associate with LFA-1 components α L and β 2 upon LFA-1s interaction with ICAM-2 in Jurkat T cells as determined by both co-immunoprecipitation experiments and confocal microscopy to demonstrate that the ICAM-2/LFA-1 interaction mediates phosphorylation of PYK2 at the plasma membrane. Furthermore, we are the first group to report that both PYK2 and SYK2 are necessary in transmitting the ICAM-2/LFA-1 induced signal to Raf, the upstream kinase in the RAF/MEK/ERK cascade, as determined by usage of specific pharmacological inhibitors piceatannol and tryphostin A9 in Jurkat T cells. PYK2 and SYK interactions have been reported in G protein coupled MAPK activity in PC12 cells (Dikic et al., 1996) and activation of HL40 cells (Miura et al., 2000), supporting the notion of PYK2 and SYK interactions in cellular processes.

Activation of the RAF/MEK/ERK cascade was monitored as a function of time per induction of ICAM-2 clustering. Blockade of p44/42 MAPK activity with monoclonal antibodies to β 2 and α L integrins indicated the ICAM-2 induced activity was mediated in part by interaction of these integrins on the cellular surface. Since monoclonal antibody treatment to other β -integrins, namely β 1 and β 3 induced p44/42 MAPK activity in vector control cells, it could suggest a general role for p44/42. MAPK activation by these specific integrins in the fibroblast cell line used, since this was not observed in Jurkat cells. The results of β 1 and β 3 mAb treatment can be accounted for by at least three possible interpretations on the ICAM-2 and ICAM2- Δ C expressing NIH3T3: (1) partial ICAM-2/partner inhibition; (2) partial integrin activation by antibody clustering; (3) combination of 1 and 2. Beta-integrins 1 and 3 have been reported to interact with Ras via G-proteins, and been implicated in the cellular adhesion/ECM signal transduction pathways resulting in p44/42 MAPK activity in adherent cell lines (Yokosaki et al., 1996; Zhu et al., 1999). Interestingly, β 1 integrin was recently implicated in p44/42 MAPK activity and subsequent cell cycle reentry (Qiang et al., 2000). Furthermore, integrin clustering initiated by extracellular matrix attachment can initiate specific signals as observed by antibody clustering (Chen et al., 1996; Hotchin and Hall, 1995).

Studies involving dominant negative Ras implicated β 2 integrin with the p44/42 MAPK in response to T cell receptor triggered ICAM-1 dependent adhesion (O'Rourke et al., 1998). Here we observed that p44/42 MAPK activity is due to ICAM-2 interacting with LFA-1, although both observations are not mutually exclusive and could suggest a mechanism in which low affinity ICAM/LFA-1 interactions are strengthened by a co-stimulatory condition such as that obtained through the T cell receptor in cell-to-cell contact processes. The ICAM-2 induced LFA-1 activation of p44/42 MAPK activity was confirmed by a soluble ICAM-2 molecule in Jurkat T cells and subsequently blocked by antibodies to the LFA-1 receptor subunits. Thus, LFA-1 is

capable of initiating a signal to p44/42 MAPK after binding to its ligand ICAM-2. Although ICAM-1 and ICAM-3 also bind LFA-1, further studies are warranted to generalize the LFA-1 induced signal to binding of all its cognate ligands, in addition to elucidating signaling pathways for other ICAM-2 receptors such as DC-SIGN.

5

Development of intracellular kinase probes to monitor kinase activity in distinct cell populations allowed us to determine the activation of p44/42 MAPK and AKT in cell-to-cell contact in LFA-1 and ICAM-2 expressing cells respectively. These adhesion signaling events initiated upon cellular contact were not only distinct to each participating cell type but illustrate that adhesion mediated events may transmit specific signaling events that have largely gone unrecognized. Detection of p44/42 MAPK and AKT activities in LFA-1/ICAM-2 engagement exemplify the significant impact that could result from deregulation of adhesion molecules, largely in part because p44/42 MAPK and AKT are two pivotal kinases involved in growth, proliferation, and survival pathways (Chang and Karin, 2001; Datta et al., 1999). The work presented herein brings to question the canonical segregation of integrins and leukocyte antigens as strictly playing adhesive/targeting roles and exemplifies that distinct signaling mechanisms are initiated upon ICAM-2 encountering LFA-1 to both interacting cells. Elucidating the roles of (I)CAM mediated adhesion in vivo could provide fundamental knowledge of their roles in functionally relevant cellular processes such as T cell activation, angiogenesis, and metastatic spread.

20

Materials and Methods

Immunological and Chemical Reagents

The following antibodies were obtained from Cell Signaling Technologies (CST): phospho-Raf1 (Ser259) polyclonal, phospho-MEK1/2 (Ser217/221) polyclonal, phospho-p44/42 MAP kinase (Thr202/Tyr204), phospho-p44/42 MAPK (Thr202/Tyr204) monoclonal, phospho-p90RSK (Ser381) polyclonal, phosphoElk-1 (Ser383) polyclonal, phospho-p38 MAP kinase (Thr180/Tyr182), polyclonal, immobilized phospho-p44/42 (Thr202/Tyr204) monoclonal, phospho-SAPK/JNK (Thr183/Tyr185) polyclonal, phospho-CREB (Ser133) polyclonal, phosphoSEK1/MKK4 (Thr261) polyclonal, phospho -Jun (Ser 63) polyclonal, phosphoMKK3/MKK6 (Ser189/207) polyclonal, phospho-AKT Ser473 monoclonal, nonphospho specific antibodies to the above proteins were also obtained from Cell Signal Technologies. The following antibodies were obtained from Transduction Laboratories: anti- β 1 monoclonal, anti- β 2 monoclonal, anti- β 3 monoclonal, anti- α_L monoclonal, anti- α_4 monoclonal, anti- α_5 monoclonal, anti- α_6 monoclonal, anti-LFA-1 monoclonal, anti- α_5 monoclonal, anti- α_1 monoclonal, anti- α_4 monoclonal, anti-PYK2, anti-SYK, anti-CD3.

The following antibodies were obtained from varying vendors: ICAM-2 monoclonal (IC2/2 Research Diagnostics), ICAM-2 Cterminal polyclonal (Santa Cruz Biotechnologies: SCBT), anti-PYK2p402 (Biosource), anti-phosphotyrosine-FITC (CST), CD3-APC (PharMingen), CD4 PerCP (PharMingen), CD80-PE (PharMingen), CD86-FITC (PharMingen), CD54PE (PharMingen), CD50 (PharMingen), CD102-FITC (Research Diagnostics), anti-rabbit IgG HRP conjugated (CST), anti-mouse IgG HRP conjugated (SCBT), anti-mouse IgG FITC conjugated (PharMingen), anti-rabbit-PE conjugated (PharMingen), anti-goat rhodamine conjugated (SCBT), anti-mouse Alexa Fluor 488 conjugated (Molecular Probes), anti-rabbit Alexa Fluor 488 conjugated (Molecular Probes) anti-rabbit Alexa Fluor 568 conjugated (Molecular Probes), anti-rabbit Alexa Fluor 568 conjugated (Molecular Probes), anti-rat IgG (PharMingen). Protein and chemical reagents used: streptavidin-FITC (SCBT), fluorescein isothiocyanate (FITC) (Pierce), R-phycoerythrin (Molecular Probes), Alexa488 (Molecular Probes), Alexa Fluor 546 (Molecular Probes), Elk-1 fusion protein (CST), p42 MAP kinase (Erk2) (CST), Tyrphostin A9 (Calbiochem), tyrphostin 18 (Calbiochem), SB203580 (CST), herbimycin A (Sigma), emodin (Sigma), piceatannol (Calbiochem), genistein (Sigma), DMSO (Sigma), phorbol myristelated acetate (PMA) (Sigma), ionomycin (Sigma), propidium iodide (PI) (Sigma), RNASE (Sigma), MTT (Sigma), streptavidin-agarose (SCBT).

Cell Culture and cDNA isolation and retroviral transduction

Full length ICAM2 cDNA was obtained from a retroviral cDNA library produced from Jurkat cDNA and cloned into retroviral vector PBM-Z-IN backbone (Kinoshita et al., 1998) at BamHI/Sal1 site. ICAM2-AC was generated by PCR and cloned into the BamHI/Sal1 site. Retroviral constructs containing ICAM-2 or ICAM2-AC were used to transduce NIH3T3 murine fibroblast. NIH3T3 cells were maintained in DMEM, 10% DCS, 1% PSQ (Duelbecco Modified Eagle Media, 10% Donor calf serum, 1% penicillinstreptomycin (1000 units/ml and 2mM L-glutamine PSQ). Jurkat T-cells were maintained in RPMI-1640, 10% FCS, 1% PSQ. BaF3 pro-B-cells were maintained in RPML1640, 10% FCS, 1% PSQ, 400 U/ml IL-3 (Peprotech). HL60 myeloid leukemia cells were maintained in Opti-Mem 1, 10% FCS, 1% PSQ. 293T human fibroblast cells were maintained in DMEM, 10% FCS, 1% PSQ. Cells were maintained at 5% CO₂ /37°C humidified incubator. Transduced cells were maintained in 500 1µg/ml G418 (Sigma) and proper expression of construct was verified by western blots and FACS analysis of full length ICAM-2 and ICAM2-AC as indicated. Vector controls consisted of pBMN-Z-IN (empty vector) transduction as indicated. The infection frequency of pBMN-Z-IN was 48% +/- 10 for three independent viral transductions. Continual neomycin selection yielded homogenous ICAM-2, ICAM2-ΔC, or control vector populations as routinely monitored by flow cytometry for protein expression.

Cross-linking

Antibody cross-linking experiments were performed using a mouse monoclonal anti-ICAM-2 that recognizes the N-terminal region (extracellular domain) (IC2/2 Research Diagnostics) at 10 µg/ml. Spin dialysis (Biorad) was used for buffer exchange of azide containing antibodies
5 (0.01%) to phosphate buffered saline buffer pH 7.4. Anti-mouse IgG (Sigma) was used as a control antibody in crosslinking experiments. 106 cells as indicated were serum starved for 3 hours or 4 hours respectively. Cells were incubated with either ICAM-2 (10 µg/ml) or mouse IgG (10 µg/ml) at 37°C for the indicated time. Time points began post serum starvation to attenuate most of the signaling pathways or as indicated. Cells extracts were taken and
10 subjected to either immunoprecipitation or immunoblotting.

Cell growth assays

Cell cycle analysis was performed as described (<http://www.metazoa.com/upl2072>) and analyzed by flow cytometry. Cellular respiration was determined by the metabolic indicator dye
15 3-(4,5-cimethylthiazot 2-yl)-2,5-diphenyl tetrazolium bromide) (MTT) (<http://www.metazoa.com/upl2102>) (Sigma).

Affinity Chromatography, ICAM-2-FITC probe generation, and surface protein biotinylation

20 Surface proteins of ICAM-2, ICAM2-ΔC, and vector control were biotinylated using sulfo-NHS-LC-Biotin (Pierce) as suggested by manufacturer and verified by flow cytometry and western blot techniques (data not shown). An ICAM-2 solid support was synthesized by coupling an anti-ICAM2 C-terminal antibody to an Affi-Prep 10 solid support (Biorad) as stated by manufacturer's instruction. The anti-ICAM-2 support was used to isolate ICAM-2 from
25 retrovirally transduced ICAM-2 expressing NIH3T3 cells, thoroughly washed, and antibody/protein complex stabilized by chemical crosslinking with bis(sulfosuccinimidyl) suberate (Pierce) as suggested by manufacturer. Alternating washes with low/high pH solutions removed contaminating proteins and or chemicals. Solid support was tested for human ICAM-2 coupling by immunoblot techniques (data not shown). The ICAM-2 solid support was designed
30 to orientate the anchored ICAM-2 molecule by the C-terminal (via an antibody), allowing the extracellular N-terminal to freely interact with passaged material (see Materials and Methods for elaboration).

Cell lysates were then passaged over the column and were initially subjected to a step gradient
35 elution using increments of 100, 250, 500 and 1 mM NaCl (in Tris pH 6.8) to elute binding

proteins. Dual column system utilized the ICAM-2 solid support, binding proteins eluted with 500 mM NaCl (in Tris pH 6.8), concentrated (Centricon 3 kDa cutoff), dialyzed using dialysis membrane (3 kDa cutoff) in PBS pH 7.4 overnight, and depleted for biotinylated proteins by streptavidin-agarose. Further procedure was similar to immunoprecipitation protocol described below. Interacting proteins that survived the procedure were reduced in SDS-PAGE buffer, boiled for 5 minutes, and subjected to SDS-PAGE analysis. Bands appearing were gel eluted and subjected to protein sequencing by MALDI-MS. Verification of the identity of the interacting proteins was done using monoclonal antibodies as indicated in immunoblots. ICAM-2-FITC generation was achieved by immunodepleting ICAM-2, antibody complex stripped by using 4.5 M NaCl (in Tris pH 6.8), concentrated, dialyzed, and chemically conjugated to NHS-Fluorescein (Pierce) as suggested by manufacturer. ICAM-2-FITC probe was stored in PBS containing 0.1 % azide.

Flow Cytometry

Intracellular and extracellular staining was performed as described (<http://www.metazoa.com/UPL3287>). Intracellular probes for AKT and p44/42 MAPK activity was made by conjugating monoclonal anti-AKTser473 antibody or monoclonal anti-p44/42 phospho (Thr202/Tyr204) (Cell Signaling Technology) to Alexa-568 dye or Alexa Fluor 488 (Molecular Probes), using Alexa Fluor protein conjugation kit (Molecular Probes) respectively. Phospho specificity was tested by western blotting and FACS analysis to a variety of PI3K activators (platelet derived growth factor) and inhibitors (LY294002) for AKT activity, and to epidermal growth factor and MEK1/2 inhibitors U0126 (CST) for p44/42 MAPK activity (data not shown). Intracellular staining by phospho-AKTser473Alexa568 and phospho-p44/42-Alexa488 reflected AKT kinase and p44/42 MAPK activity in Jurkat cells when stimulated and inhibited prior to stimulation. Quantitative FACS analysis was performed as described (Davis et al., 1998; Iyer et al., 1998; Lenkei et al., 1998).

In brief, R-phycoerythrin (PE) (Molecular Probes) was conjugated to indicated antibody as suggested by manufacturer of protein cross-linking kit (Molecular Probes), tested for proper stoichiometry (data not shown) and quantitated using QuantiBRITE-PE beads (BD systems). A quantitative calibration curve was generated using QuantiBRITE-PE beads that contain a known amount of PE molecule/bead. A linear regression analysis is performed using the following equation: $\log_{10}(\text{PE fluorescence}) = \text{slope} \times \log_{10}(\text{PE molecules/bead}) + \text{intercept}$. PE fluorescence is determined by taking the geometric mean of the PE channel. Quantitation is valid only for antibodies directly conjugated to PK. Using saturating amounts of antibody and thorough washing ensures all surface antigens are bound. Preparation of samples on ice reduces antibody internalization. Surface antigens of cells for an unknown cell population are

determined by computing the PE geometric mean using the calibration curve. Quantitative FACS analysis provides approximate estimates for surface antigen expression. Numbers plotted represent relative surface molecules and not absolute numbers since antibody valency was not investigated for the monoclonal antibodies used. Flow cytometry analysis was performed on a BD FACSCalibur machine and analyzed using FlowJo software (Tree Star). CellQuest was used for quantitative flow cytometry and linear regression analysis of QuantiBRITE-PE beads. Flow cytometry data are representative of 3 independent experiments of 106 cells/sample analyzed. 50,000 events were collected or otherwise noted and calibrated using Calibrite beads (BD systems). Data plotted in bar graph format is expressed as the mean (bar) +/- SD of triplicate experiments.

Confocal microscopy

Jurkat cells were treated as indicated and adhered to poly-L-lysine (Sigma) coated sterilized coverslips (10 mg/ml, 30 minutes) by mild centrifugation (1000 RPM, 10 minutes), washed twice in phosphate buffered saline pH 7.4 (PBS) and fixed in 2.7% paraformaldehyde (in PBS). Cells were permeabilized for 5 minutes with 0.1% saponin (in PBS), washed twice in PBS, blocked in 4% bovine serum albumin (BSA, in PBS), and subjected to antibody incubation: (primary at 0.1 mg/ml, secondary at 1:1000 dilution in 1% BSA, with several washing steps in between). Primary antibodies used: anti-PYK2 monoclonal, anti-SYK polyclonal, anti-PYK ptyr402 monoclonal, anti-SYK polyclonal, phospho-tyrosine-FITC. Secondaries used: anti-mouse-Alexa 488, anti-rabbit-Alexa 488, anti-rabbitAlexa-568, anti-rabbit-Alexa-568. Stained coverslips were mounted onto glass slides with Prolong Antifade reagent (Molecular Probes) and visualized using a Molecular Dynamics Multiprobe 2010 confocal laser scanning microscope. Images were compiled using Adobe Photoshop 6.0.

MAPK kinase assays

MAPK activity was detected by immunoprecipitation of active phosphorylated p44/42 from cells and used in a kinase assay with Elk-1 fusion protein (Cell Signaling Technologies; CST). 2×10^6 NIH3T3 or Jurkat cells were incubated with immobilized phospho-p44/42 monoclonal antibody (mAb) (1:200, CST) at 4°C with gentle rocking motion for 2 hrs. Immunocomplexes were washed 4X with cell lysis buffer and resuspended in 40 μ l kinase buffer (25 mM Tris pH 7.5, 5 mM β -glycerolphosphate, 2mM DTT, 0.1 mM Na_3VO_4 , 10 mM MgCl_2) supplemented with 200 μ M ATP and 1 μ g of Elk-1 fusion protein (CST) for 30 min at 30°C. Kinase reaction was terminated with SDS sample buffer boiled for 5 min, and phosphorylation state of Elk-1 was detected by immunoblotting and visualized using ECL detection (Amersham). Immunoblots are

representative of 3 independent virally transduced cell populations each repeated 3 times (n=9). MAPK activity was verified by MAPK phosphorylation by immunoblotting with phospho-specific antibodies

5 Immunoprecipitations and Immunoblotting

Cell extracts were prepared by washing 2×10^6 cells (treated as indicated) in ice cold PBS and harvesting in lysis buffer (20 mM Tris pH 7.5, 150 mM NaCl 1 mM EDTA 1 mM EGTA, 1 % Triton X-100, 2.5 mM Na_2PO_4 , 1 mM β -glycerolphosphate, 1 mM Na_3VO_4 , 1 $\mu\text{g/ml}$ Leupeptin, 1 mM PMSF, protease inhibitor cocktail tablet (Boehringer Mannheim)). Extracts were
10 centrifuged 14,000 RPM for 5 min at 4°C and cell lysates (20 μg as determine by BCA protein assay (Pierce)) were fractionated on 15% SDS-polyacrylamide gel electrophoresis and transferred to PVDF membranes using standard procedures. Immunoprecipitations (IP) were pre-cleared with protein A/G plus-agarose beads (Santa Cruz Biotechnologies; SCBT). IP was incubated for 1 hr with primary antibody, 1 hr with protein A/G plus-agarose beads and washed
15 4X with lysis buffer. Blots were incubated with the indicated antibodies and developed using ECL (Amersham). Fluorescent blots were visualized on a Kodak Digital Image station 440 C station. Immunoblots stripped and reprobed (as indicated) were done by incubating with stripping buffer (62.5 mM Tris, pH 6.8, 10% SDS, 1% β -mercaptoethanol) for 30 minutes at 55°C and subjected to a repeat of the entire immunoblot protocol.

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Example 4

30 Immune Cell Survival: Activation of the PKB/AKT Pathway Intracellular Adhesion Molecule-2 (ICAM-2)

In this Example, using the methods and compositions of the present invention, the present inventors (also referred to herein as "we") using a retroviral cDNA library in a functional genetic screen, identified intracellular adhesion molecule-2 (ICAM-2), a member of the immunoglobulin-
35 family of adhesion molecules, as a potent inhibitor of several activators of apoptosis in varying cell types and induction settings. The anti-apoptotic effect was mapped to the activation of the PI3K/AKT pathway. ICAM-2 induced tyrosine phosphorylation of ezrin recruited PI3 kinase to

the membrane and resulted in phosphatidylinositol 3,4,5 production, PDK-1 activity, AKT membrane translocation and activation and subsequent phosphorylation of AKT targets BAD, GSK3, FKHR and AFX. The ICAM-2 mediated survival function was abrogated by pharmacological inhibitors of Src (herbimycin A), Rho-dependent kineses (Y-27632),
5 phosphoinositide turnover (Psi-tectorigenin), and of PI3 kinase (wortmannin and LY294002), indicating activation of AKT and its downstream effectors were dependent on PI3K recruitment to the plasma membrane by an ICAM-2 induced ezrin activation. Primary CD19+ cells were protected from TNF α and Fas mediated apoptosis following ICAM-2 clustering and presented elevated AKT activity and phosphatidylinositol 3, 4, 5 (PIP3) as detected by direct single cell
10 flow cytometric measurements of intracellular AKT kinase activity and phosphatidylinositol levels. ICAM-2s engagement with its natural receptor, LFA-1, induced AKT activity similar to that of antibody crosslinking. This cell survival signal was shared only by ICAM-3 in primary B cells, and not by ICAM-1, CD43, and CD44, indicating that the binding of ezrin or interaction with LFA-1 is not sufficient for initiation of the cell survival signal as observed for ICAM-2.
15 These results attribute a novel survival signaling function to ICAM-2 that might provide an explanation for both the role of ICAM-2 overexpression in B-cell lymphomas and mechanisms by which ICAM-2 might signal intracellular communication in cell-to-cell contact at various immunological synapses.

20 Introduction

A cell's commitment to cellular proliferation or programmed cell death is a balance of the survival and death signals communicated from both the immediate environs, such as extracellular matrix (ECM) interactions, cell-cell contact, or more distant sites through endocrine and paracrine factors (Ruoslahti and Reed, 1994). Survival pathways are proposed
25 to function during early development where apoptosis plays a key role in shaping organogenesis (Raff, 1992) and are eminent in the maturation of the immune system. Clonal selection of antibody forming cells and maintenance of memory B cells are essential processes that require activation of cell survival pathways. Similarly, the lack of survival signals is exemplified in the phenomenon of anoikis, where epithelial cells undergo apoptosis when
30 ECM-attachment is blocked (Frisch and Ruoslahti, 1997). ECM survival signals are presumably mediated by activation of cell-surface receptors via integrin clustering, contributes to the specificity of cell cell/cell-matrix interactions, and deregulation of ECM-integrin signaling leads to anchorage-independent growth (Hulleman and Boonstra, 2001). Identification of analogous molecules in lymphoid cells as those that can antagonize cell death or override

growth inhibition programs would be pivotal in understanding intrinsic cell survival programs that can go awry in lymphatic disorders and possibly contribute to metastatic spread.

5 Functional characterization of molecules involved in cellular adhesion has been focused on signal transduction pathways attributed to integrins and cadherins, which can contribute to ECM-survival signals (de Fougerolles et al., 2000; Schneller, 2001; Shinohara et al., 2001). Adhesion molecules of the immunoglobulin (Ig) superfamily have been largely overlooked as potential signaling molecules. ICAM-2 (CD102) is a member of the Ig-superfamily of cell surface proteins and mediates leukocyte binding to LFA-1 (CD11a/CD18) and MAC-1 (CD11
10 b/CD18) (van Kooyk and Figdor, 2000). As a cell surface adhesion member involved in leukocyte recruitment in tissues, ICAM-2 is expressed in low levels on most leukocytes including T and B lymphocytes, monocytes, platelets and early CD34⁺ hematopoietic progenitor cells (de Fougerolles et al., 1991; Diacovo et al., 1994; Nortamo et al., 1991). In addition, ICAM-2 constitutive expression has been observed on all vascular endothelium (de Fougerolles
15 et al., 1991). Based on the expression pattern of ICAM-2, it has been postulated to be involved in lymphocyte recirculation, trafficking, and extravasation. ICAM-2 deficient mice support this by demonstrating that eosinophil trafficking is augmented in inflammatory responses (Gerwin et al., 1999). Surprisingly, this phenotype is relatively mild considering the range of cell types upon which ICAM-2 is expressed. It suggests that the roles ICAM-2 plays are subtle or are
20 redundant with other pathways. Thus, the ICAM-2 deficient mice do not sufficiently detail the roles of ICAM-2, warranting further characterizations of the mechanisms by which ICAM-2 acts are needed.

In contrast to ICAM-2's putative role as an adhesion molecule, it is surprising to find ICAM-2
25 highly expressed on both endothelial cells in lymphomas and CD5⁺ B cells from B-cell chronic lymphatic leukemia (B-CLL) patients (Molica et al., 1996; Renkonen et al., 1992). Lymphoproliferative diseases inevitably inhibit adhesion dependency-loss of adhesion is one of the primary means by which metastatic spread occurs. However, recent gene expression profiling of large B-cell lymphomas from patients indicated an elevated expression of ICAM-2
30 (Alizadeh et al., 2000). Interestingly, ICAM-2 is the only ICAM (1-5) that maps to chromosome 1 7q23-25 (Sansom et al., 1991), a segment associated with genomic instability and recently identified as a segment of high aberration in various cancers (Dion et al., 2000; Dobo et al., 1995; Russell et al., 2000; Sugai et al., 2000). Despite these observations, ICAM-2's role and molecular characterization in the progression of lymphatic disease states has not been
35 addressed.

We devised a genetic screen to identify genes that are involved in regulation of apoptosis and augmented in malignant phenotypes. Cells transduced with a human lymphoid cell line (Jurkat) retroviral vector cDNA library were screened for an anti-apoptotic phenotype. One cDNA demonstrated reproducible antiapoptotic activity in a phenotypic transfer assay. This cDNA encoded full-length intracellular adhesion molecule-2 (ICAM-2). Here we show functional characterization of the ICAM-2 signal as being capable of mediating a cell survival signal upon its ligation and interaction with its endogenous receptor leukocyte function-antigen-1 (LFA-1). ICAM-2 mediates a survival signal sufficient to block apoptosis by activation of the PI3K/AKT pathway, being contingent on ezrin activation by Src tyrosine phosphorylation and Rho-kinase dependent threonine phosphorylation. Ezrin activation recruited PI3K to the cell membrane in lymphoid cells, thereby initiating activation of the PI3K/AKT pathway. Furthermore, development of prob to measure intracellular AKT and PI3K activity by flow cytometry allowed for the simultaneous measurement of kinase activity as a function of ICAM-2 clustering on human peripheral blood monocytes (PBMC). Multi-parameter FACS analysis confirmed ICAM-2 clustering led to phosphatidylinositol 3,4,5 production and AKT activity in CD19⁺ primary human cells and subsequent protection from Fas and TNF α mediated apoptosis. Interestingly, activation of the PI3K/AKT pathway was shared by ICAM-3 and not by ICA-1, CD43, CD44 indicating that molecular differences exist amongst the 19 adhesion molecules that are capable of binding ezrin (Serrador et al., 1997; Yonemura and Tsukita, 1999). Ligation of ICAM-2 by its endogenous receptor LFA-1 was observed to induce AKT activity, illustrating that the ICAM-2/LFA-1 interaction is capable of activating the PI3K/AKT pathway. These observations identify a bimodal functional role for ICA-2 in mediating cell survival. Overexpression of ICAM-2 conferred an anti-apoptotic phenotype, suggesting that inappropriate expression of ICAM-2 could contribute significantly to the antiapoptotic phenotypes in the cumulative process that leads to tumor progression and metastatic spread. The findings also underscore a novel positive role for ICAM-2 in signaling pathways involving Src/Rho-Kinase/Ezrin/PI3K/PDK-1/AKT and downstream effectors upon LFA-1 engagement in cell-to-cell contact with implications for other ICAM-2 interactions such as ICAM-2/DC-SIGN and ICAM2/Mac-1.

RESULTS

Screening a retroviral cDNA library in an anti-apoptotic functional assay

We devised a retroviral library approach to screen for cDNAs that alter apoptotic signaling and encode for anti-apoptotic molecules (Figure 19A). cDNAs from a malignant cell type (Jurkat T cell leukemia) were screened for anti-apoptotic function in a recipient non-malignant (fibroblast)

cell. Apoptosis was induced with staurosporine (STP), a kinase-inhibiting plant-derived alkaloid, and a potent inducer of caspase-dependent cell death in most cell types. Induction conditions were determined that limited the level of spontaneous background while maximizing the extent of apoptotic death (minimal background of one surviving cell in a STP-treated mock library control in ~10⁴ starting cells). 10⁷ NIH3T3 fibroblasts were infected with a retroviral cDNA library derived from Jurkat cells at an infection frequency of 40% to limit the number of integrations to a single event per cell. Control retroviruses were prepared expressing LacZ- or Bcl2, and infected into a similar number of NIH3T3 cells. Apoptosis was induced in the transduced cell culture by staurosporine treatment and the cells were then allowed to recover (Figure 19A). Surviving cell clones were expanded, replated and retreated with staurosporine. The complexity of the expressed cDNAs in the surviving population was assessed with RT-PCR. A few major bands were observed following three rounds of selection (Figure 19B).

Replication-competent Moloney murine leukemia virus (MMLV) was applied to the enriched cell populations. Upon integration and expression of the MMLV proteins, resident retroviral constructs from the library were co-packaged into infectious virions and transferred to naive target NIH3T3 cells. The staurosporine selection process was then reapplied. In this manner true phenotype-inducing clones enrich at a rate of 1/background (see materials and methods for elaboration). Three of the pooled libraries showed enrichment for specific bands after PCR (see Figure 19B lanes 1 and 2). Several bands were rescued by PCR cloning and one of these bands was sequenced and demonstrated to encode full-length ICAM-2 cDNA (Staunton et al., 1989), an immunoglobulin superfamily member that regulates leukocyte adhesion through interaction with its integrin counter-receptor, LFA-1 (de Fougères et al., 1991; Simmons, 1995).

ICAM-2 mediates a survival signal

The ICAM-2 cDNA was cloned into a retroviral vector (pBMN-Z-IN) that carries an internal ribosome entry site (IRES) upstream of the neomycin resistance gene. NIH3T3 fibroblasts were infected with retroviruses capable of expressing the ICAM-2 gene, along with appropriate control vectors pBMN-Z-IN (vector control) and pBMN-Z-IN-GFP (GFP vector control). Expression of the 60 kD ICAM-2 glycoprotein in neomycin-selected NIH3T3 fibroblasts infected with the pBMN-Z-IN-ICAM-2 construct (referred to as ICAM-2 cells) was verified by immunoblotting (Figure 19C), flow cytometry (Figure 19 D3A), and immunoprecipitation (Figure 22A). The expressed ICAM-2 protein co-migrated with the native protein from Jurkat T cells (data not shown).

Staurosporine treatment is thought to mimic factor-withdrawal in cells (Raff, 1992), and induces apoptosis by inhibiting PKC, PKA, and PKG at nanomolar concentrations among other kinases (Meggio et al., 1995). ICAM-2 expression in NIH3T3 cells limited the extent of apoptosis induced by staurosporine treatment as determined by an annexin-V binding assay (Figure 20A), pyknotic nuclei count (Figure 20B) and the BrdU TUNEL assay (data not shown, Reviewers see Supplementary Figure 19A-B) confirming the clone's selection by phenotype in the screen. Interestingly, not all forms of apoptosis were inhibited by overexpression. ICAM-2 overexpressing Jurkat T cells did not inhibit apoptosis across a range of concentrations of a monoclonal antibody to the Fas receptor, contrasting inhibition of staurosporine induced apoptosis (Figure 19E, panel 2-3, elaborated below).

A report correlating ICAM-2 expression with a lymphoproliferative disorder, B-cell chronic lymphatic leukemia (B-CLL) (Molica et al., 1996), prompted us to test the anti-apoptotic activity of ICAM-2 in the mouse pre-B-cell line, 70Z/3. ICAM-2 expression inhibited staurosporine-induced apoptosis in 70Z/3 cells (Figure 20B). Given the indication that staurosporine mimics factor withdrawal, we assessed the ability of ICAM-2 expression to protect factor-dependent cells from factor withdrawal. ICAM-2 expression in the IL-3-dependent pro-B cell line BaF3 resulted in a delayed onset of apoptosis following IL-3 deprivation (Figure 20B). Furthermore, we tested the ability of ICAM-2 to rescue BaF3 cells from apoptosis induced by treatment with anti-Fas antibodies. ICAM-2 demonstrated a potent inhibition of apoptosis through the Fas- pathway in these cells (Figure 20B). These results are as good as or better than transduction controls with Bcl2 and therefore the apparent effects cannot be attributed to modest effects on apoptosis or retroviral transduction (Figure 20B, data not shown). There is, however, cell specificity to the anti-apoptotic effect by virtue of ICAM-2 ectopic expression as ICAM2 overexpressing Jurkat T cells were not protected from Fas-induced apoptosis but were protected from staurosporine induced apoptosis (Figure 20B). High throughput proteome analysis recently identified that in Jurkat T cells, Fas induced apoptosis results in a rapid destruction of 12 cytoskeletal proteins, alters phosphorylation states of various chaperone proteins and cytoskeletal structural proteins, and presents a more "aggressive" form of cell death than staurosporine induced apoptosis (Gerner et al., 2000). Therefore different cell death inducers have specific physiological characteristics in Jurkat T cells supporting the differences observed (Johnson et al., 2000). However, we later provide evidence for a bimodal function of ICAM-2, in which ligation of endogenous ICAM-2 on Jurkat

cells is sufficient to block Fas induced apoptosis suggesting that overexpression is insufficient to initiate cell survival in this cell type (described below).

The initial screening was performed in fibroblasts, and it is therefore unlikely that ICAM-2's anti-apoptotic effect was mediated through its lymphocyte-specific integrin ligand, LFA-1 (CD11a/CD18), since NIH3T3 do not express LFA-1 (data not shown). To address this possibility in lymphoid cells, we prepared an extracellular domain-deleted (AN) version of ICAM-2 and tested it in BaF3 cells. Expression of the cytoplasmic domain was sufficient to impart full anti-apoptotic activity (Figure 20B). This indicates that the cytoplasmic domain of ICAM-2 provides a survival signal when overexpressed ectopically, and likely exerts its effect by mimicking receptor crosslinking. Since signaling through ICAM-family cytoplasmic domains is poorly understood, we sought to identify the signaling pathway by which it conferred resistance to apoptosis. The ICAM-2 survival function maps to a membrane-cytoskeleton linker protein binding motif. ICAM-2 is regarded as an adhesion molecule and has been shown to be interconnected with the actin cytoskeleton by linker proteins α -actinin and ezrin, both of which bind the ICAM-2 cytoplasmic domain. (Heiska et al., 1996; Helander et al., 1996). To assess the potential role of α -actinin in mediating the ICAM-2 anti-apoptotic signal, the α -actinin recognition sequence was changed to a scrambled sequence unable to bind α -actinin in vitro (Heiska et al., 1996). Retroviral constructs encoding ICAM-2, a C-terminal deletion mutant (ICAM2-AC) and a full-length ICAM-2 protein with the mutated α -actinin binding motif (ICAM2C scrambled) were used to transduce NIH3T3 cells (Figure 20C). The infected cells were sorted by FACS using an ICAM-2 monoclonal antibody (IC2/2) recognizing only native N-terminal ICAM-2 (de Fougères et al., 1991). This verified that ICAM-2, ICAM2-AC, or ICAM2-C-scrambled proteins were properly folded and targeted to the outer face of the plasma membrane. The NIH3T3 cells expressing the ICAM-2 mutants were assayed for survival following both etoposide treatment (another potent inducer of apoptosis in NIH3T3 cells) and staurosporine treatment. ICAM-2 imparted potent long-term survival on NIH3T3 cells and the survival signal was lost when the terminal deletion of ICAM-2 (ICAM2-AC) was expressed in cells (Figure 20C). Mutation of the α -actinin binding site in the ICAM-2 cytoplasmic tail also did not provide any protection against etoposide or staurosporine treatments. Although the motif within ICAM-2s C-terminal is identified as an α -actinin binding site, recent studies have shown that the positively charged amino acid cluster the cytoplasmic domain can also interact with ezrin, a member of the ezrin/radixin/moesin (ERM) cytoplasmic linker protein family (Yonemura et al., 1998). Whereas both α -actinin and ezrin co-immunoprecipitated with only full length ICAM-2, a significant difference in the quantity of ezrin binding protein was observed relative to α -actinin (Figure 22A), supporting the postulate that these cytoskeleton linker proteins were necessary but may differentially relay an anti-apoptotic signal. Experiments designed to block

activation steps of each of these cytoplasmic linker proteins elucidated differences between fibroblastoid and lymphoid cell types and their requirements for a-actinin and/or ezrin to relay the anti-apoptotic signal (elaborated below).

5 ICAM-2 signals through PI3K/AKT kinase

As survival signaling through many adhesion molecules involves PI3 kinase (Frisch and Ruoslahti, 1997; Khwaja et al., 1997), we analyzed the role of this pathway in the ICAM-2 anti-apoptotic signal. Pretreatment of ICAM-2 expressing BaF3 cells with the PI3 kinase inhibitor wortmannin blocked the ICAM-2 mediated anti-apoptotic effect (Figure 21A, left panel),
10 implicating PI3 kinase in the ICAM-2 survival signaling cascade. Under these conditions vehicle-treated ICAM-2 expressing BaF3 cells were resistant to staurosporine induced apoptosis. Similarly, treating ICAM-2 transduced NIH3T3 with LY294002 abrogated ICAM-2's anti-apoptotic effect as measured by both annexin-V binding and BrdU TUNEL assay using flow cytometry (Figure 21A, data not shown, reviewers see Supplementary Figure 19).
15 Anti-apoptotic effect was not abrogated with inhibitors to p44/42 MAPK (U0126 and PD98059), p38 MAPK (SB 203580), PCK isozymes (bisindolymaleimide 1 and 11), or PKA (H-89), excluding the involvement of these kinsases in the signal transduction pathway (data not shown).

20 We utilized confocal microscopy to visualize the cellular distribution of PI3K, AKT, the p85 regulatory subunit of PI3K, and ezrin. We observe both PI3K and AKT to be concentrated at the outer periphery of ICAM-2 cells (Figure 21B). Ezrin and the p85 subunit was visualized to co-localize more extensively in ICAM-2 cells versus vector control (Figure 21C). ICAM-2 overexpression had augmented the distribution of PI3K and AKT to favor a membrane
25 localization which is required for their respective activation (Reviewers See Supplementary Figure 20A). Alpha-actinin and ezrin both co-immunoprecipitated with ICAM-2 (Figure 22A) and subsequently tested for PI3K interaction since direct PI3K interaction with ICAM-2 was not observed in the absence of either a-actinin or ezrin (data not shown).

30 Development of single cell based phosphatidylinositol 3,4,5 triphosphate (PIP3) and phosphatidylinositol 4,5 bisphosphate (PIP2) detection made it possible to assess PIP3/PIP2 levels within intact cells and reflect PI3K and PI4K/PLCy1 kinase activities (Perez and Nolan, manuscript in preparation). ICAM-2 cells presented uniformly high levels of PIP3 versus the asynchronous production of PIP3 in vector controls and were inhibited by serum starvation and
35 incubation with LY294002 (Figure 22B). Production of PIP3 serves as a secondary messenger

to recruit pleckstrin domain containing proteins, such as AKT, to the plasma membrane, and also serves to activate others, namely phosphatidylinositol-dependent kinases (PDK-1 and PDK-2) (Blume-Jensen and Hunter, 2001). The production of PIP3 in ICAM-2 cells paralleled the AKT localization pattern observed as expected for active PI3K.

5

PI3 kinase is known to regulate apoptosis through its downstream effector AKT (PKB) kinase (Downward, 1998). AKT kinase has a potent anti-apoptotic effect downstream of diverse apoptotic stimuli, including staurosporine (Marte and Downward, 1997)(data not shown). We assessed PDK-1 kinase activity in ICAM2 cells and found elevated PDK-1 activity in
10 comparison with vector control (Figure 22C). This effect reflected the increased levels of PIP3 detected in ICAM-2 cells as it was inhibited by incubation with LY294002, and to a lesser extent, Y27632, a specific pharmacological inhibitor of Rho-associated protein kinases. Cytosolic/membrane fractionation of ICAM-2 cells confirmed localization of AKT to the membrane versus cytoplasmic localization in vector controls (Figure 22D), congruent with
15 confocal microscopy of AKT localization. ICAM-2 expressing fibroblast demonstrated an elevated level of AKT activation that was absent in ICAM2-AC or mock transduced cells (Figure 22D). We further verified that this ICAM-2-dependent elevated AKT kinase activity is maintained following STP treatment using both kinase assays and by verifying phosphorylation of both ser473 and thr308 (Figure 22E). A target of AKT kinase is the pro-apoptotic Bcl2 family
20 member, BAD which is normally held in an inactive state by phosphorylation of serines 112 and 136, and is bound to 14-3-3. Upon induction of apoptosis, BAD is dephosphorylated at both ser112 and ser136, and dimerizes with Bcl-2. As a result, the equilibrium of Bcl2/Bcl-XL is shifted to favor release of Bcl-XL to mediate Cytochrome C release and apoptosis (Datta et al., 2000). BAD is dephosphorylated following staurosporine-treatment of ICAM2-AC and
25 mock-transduced NIH3T3 cells, concomitant with the induction of apoptosis (Figure 22F); however, ICAM-2 expression results in the maintenance of BAD phosphorylation consistent (Figure 22F) with sustained AKT kinase activity (Figure 22E).

BAD itself can integrate a number of signal transduction pathways with a similar outcome,
30 possessing 3 phosphorylation sites that can be phosphorylated independent of one another: ser112 mediated by active p90Rsk1, ser136 mediated by active AKT, and ser155 mediated by both active PKA and active p90Rsk1. ICAM-2 and its role in connection to BAD ser112 phosphorylation is to be described elsewhere (Perez and Nolan, manuscript in preparation). Phosphorylation of BAD ser155 was not observed (data not shown). The anti-apoptotic signal
35 had little apparent effects on the anti-apoptotic proteins Bcl2 and Bcl-XL (Figure 22E) and was found to be independent of caspase inhibitory proteins, cell cycle regulators, anti-apoptotic pathways using Bcl-2 or additional cell survival programs (data not shown). This indicated that

typical anti-apoptotic pathways did not mediate the survival signal (data not shown). Another target of AKT kinase involved in anti-apoptotic signaling, GSK3, was found to be phosphorylated in ICAM-2 expressing NIH3T3 and probably contributes to the anti-apoptotic effect (Figure 22F, bottom panel). Additional AKT substrates, namely p70S6K and FKHR were also found to be phosphorylated in addition to a decreased detection of cleaved caspase products in ICAM-2 cells induced to apoptose (data not shown).

ICAM-2 crosslinking confirms a role for endogenous ICAM-2 in providing an anti-apoptotic signal

The ICAM-2-dependent elevation of activated AKT levels in fibroblasts is independent of ligand interactions as determined by the fact that ICAM-2 Δ N can protect cells from apoptosis and ICAM2- Δ C cannot activate AKT. This is indicative of the signaling mechanism driven by oligomerization of surface ICAM-2 molecules, a common *modus operandi* for adhesion molecules. The surface oligomers may create interaction sites within their cytoplasmic domains, recruiting signaling molecules to participate in membrane proximal interactions. Quantitative FACS analysis revealed that the density of ICAM-2 molecules on the cell surface vary widely between cell types (Figure 23A, and, reviewers see APPENDIX for quantitative methodology see Experimental Procedures for elaboration) and correlated with varied expression patterns as previously described (de Fougerolles et al., 1991). Basal expression levels of ICAM-2 on lymphocytes was 2-4 orders of magnitude less than the enforced expression from the retroviral vector as determined by the number of ICAM-2 surface molecules. We postulated that antibody cross-linking could emulate ICAM-2s interaction with its receptor, and therefore decided to test whether endogenous ICAM-2, when cross-linked, could confer the phenotypes observed as in the overexpression systems.

We cross-linked (clustered) endogenous Jurkat T cell ICAM-2 surface molecules with monoclonal antibodies and assayed activation of the PI3K/AKT pathway. Ezrin was found to associate and become tyrosine phosphorylated as a function of ICAM-2 cross-linking as detected by co-immunoprecipitation with ICAM-2 (Figure 23B). This suggests that ezrin translocates to membrane in order to interact with the C-terminal of ICAM-2. Subsequently, ezrin becomes tyrosine phosphorylated upon interacting with ICAM-2 as a time delay is detected upon co-immunoprecipitation, indicating that phosphorylation is mediated by a membrane proximal protein tyrosine kinase, since ICAM-2 lacks tyrosine kinase activity (data not shown). Ezrin tyrosine phosphorylation and concomitant association with p85 was

observed as a function of ICAM-2 cross-linking, with a notable enhancement of ezrin threonine phosphorylation (Figure 23D) effects of which were not shared by α -actinin (data not shown).

5 In order to identify upstream kinases capable of tyrosine phosphorylating ezrin as a function of ICAM-2 ligation, we developed a phosphotyrosine-protein based elisa to screen for the inhibition of ezrin tyrosine phosphorylation in vivo by various known inhibitors of protein kinases (Perez and Nolan, manuscript in preparation). Of the inhibitors screened, herbimycin A, a potent inhibitor of src tyrosine kinase family members src and p62^{yes} (Haas et al., 2000; Schieven et al., 1992) was found to inhibit tyrosine phosphorylation of ezrin, comparable to
10 genistein, which served as a control for general tyrosine kinase inhibition (Figure 23C). Surprisingly, we also found inhibitors of ROCK, a Rho-dependent serine/threonine kinase (Y-27632), inhibitors of GTPases, and inhibitors of phosphoinositide turnover (Psi-Tectorigenin) to inhibit the tyrosine phosphorylation of ezrin, with minor effects by PKC isozyme inhibitors bisindolymaleimide I and II. Given the design of the screen to detect tyrosine residue
15 phosphorylation, we speculate that activation of Rho-dependent processes may be a prerequisite for ezrin to be tyrosine phosphorylated by src (elaborated below).

Ligation of ICAM-2 on the surface of Jurkat T cells induced PDK-1 activity (Figure 23E) and incidentally membrane localization of PI3K and AKT (Figure 23F). Pursuant AKT activation,
20 dual phosphorylation of AKTser473 and AKTthr308, and phosphorylation of AKT substrates BAD, GSK3, FKHR, and AFX followed ICAM-2 crosslinking in Jurkat T cells as a function of time (Figure 24A). Crosslinking of ICAM-2 on a different cell type, pro-B cells, which also express low endogenous levels of ICAM-2 (see Figure 23C) and displayed an antiapoptotic phenotype upon ICAM-2 transduction (Figure 20B) similarly activated AKT and phosphorylation
25 of AKT substrates BAD and FKHR in a time dependent manner (Figure 24B). ICAM-2 crosslinking was found to protect Jurkat T cells from staurosporine induced apoptosis verifying the resultant outcome of the PI3K/AKT pathway activation (Figure 24C). Incubation with LY294002 abrogated the antiapoptotic effects initiated by cross-linking ICAM-2 in Jurkat T cells and BaF3 cells (data not shown).

30

We had observed that overexpression of ICAM-2 was not sufficient to block Fas induced apoptosis in Jurkat T cells (Figure 19E, 20C). Contrary but not necessarily contradictory, we observed crosslinking of endogenous ICAM-2 on Jurkat T cells to efficiently protect against Fas induced apoptosis and be abrogated in the presence of GTP- γ S, Psi-tectorigenin, Y-27632,
35 and herbimycin A (Figure 24D), inhibitors identified to block ezrin tyrosine phosphorylation (see Figure 23C). These observations suggest that ICAM-2 clustering by virtue of antibody crosslinking is necessary for imparting the cell survival signal and that ICAM-2 overexpression

in this cell type is not sufficient to block Fas induced apoptosis. Furthermore, the abated ICAM-2 induced cell survival signal by compounds GTP γ S, Psi-tectorigenin, Y-27632, and herbimycin A in lymphoid cells contrasts the mild effects observed in ICAM-2 overexpressing NIH3T3 cell types (data not shown; Reviewers see Supplementary Figure 20B). This suggests
5 that ezrin is the predominant linker between ICAM-2 and PI3K in lymphoid cells, since these compounds affect the activation of ezrin and not a-actinin (Freeman et al., 2000; Izaguirre et al., 1999; Morishige et al., 2001; Nakamura et al., 2000). This was verified by ezrin, and to a lesser extent a-actinin, predominantly coimmunoprecipitating with PI3K proceeding Jurkat ICAM-2 crosslinking (Figure 24E), although PI3K co-immunoprecipitated with both ezrin and a-actinin in
10 reciprocal co-immunoprecipitations in fibroblastoid cells (see Figure 22C). In addition, we have observed ICAM-2 overexpressing fibroblastoid cells to dampen the dynamics of the cytoskeleton architecture and require this augmentation for maintenance of its survival signal, contrasting the requirements of a dynamic cytoskeleton architecture in lymphoid cells (Reviewers see Supplementary Figure 21A-C). Therefore, ICAM-2 ligation and ICAM-2
15 overexpression present molecular differences in their mode of activating the PI3K/AKT pathway in cells of different origins.

ICAM-2 protects primary CD19+ B cells from TNF α and Fas induced apoptosis via PI3K/AKT activation and is distinct from ICAM-1, ICAM-3, CD43, and CD44

20 Although the results for AKT activity were reproducible, we were unsatisfied with the western blot analysis/kinase activity assays for whole blood derived cells and so developed a new procedure for single cell analysis of AKT activity by flow cytometry described below (Perez and Nolan, submitted, reviewers please see affached manuscript). We also developed FACS based methodologies for phosphatidylinositol 3,4,5 triphosphate, and phosphatidylinositol 4,5
25 bisphosphate detection, reflective of phospholipid turnover, PI3K and PI4K/PLC γ activity respectively within single cells (Perez and Nolan, manuscript in preparation). We utilized FACS based AKT kinase assays and FACS based PIP3 and PIP2 measurements to attribute a physiological relevance for ICAM-2 in vivo. Human PBMC were crosslinked for ICAM-2 prior to treatment with TNF α and anti-Fas monoclonal antibody. Treated PBMC were subjected to flow
30 cytometry, gated by immunophenotype for CD4 (T cell marker) and CD19 (B cell marker) and subsequently assayed for AKT activity and apoptosis by multi-parameter FACS. Intracellular AKT kinase activity measurements were achieved by development of a phospho-specific fluorescent probe for AKT-phospho ser473 phosphorylation site on AKT detectable by FACS (Perez and Nolan, submitted). Both CD4+ T cells and CD19+ B cells exhibited AKT activity
35 when ICAM-2 was clustered and induced AKT activity preferentially protected B cells from

apoptosis to Fas and TNF- α induced apoptosis (Figure 25A). There was little apparent antiapoptotic effect on CD4⁺ T cells perhaps due to the fact that these T cells were not strongly induced by Fas or TNF- α to undergo apoptosis with our conditions and require prior stimulation with IL-2 or PMA/ionomycin (data not shown). Protection from apoptosis in primary B cells correlated perfectly with AKT phosphorylation at serine 473, confirming prior roles for this target serine in the protective effect of AKT against Fas and TNF- α induced apoptosis. FACS based kinase activity measurements for AKT allow unique activation profiles in distinct lymphocyte populations to be assessed and circumvent the functional augmentation often observed in pre-sorting for such cell populations (data not shown). Differences were observed when ICAM-2 was compared to other ezrin binding surface molecules, namely ICAM-1, -3, CD43, and CD44. PIP3 and PIP2 FACS based measurements identify ICAM-2 as increasing the ratio of PIP3/PIP2, congruent with PI3K activity (Figure 25B). ICAM-3 was found to increase both PIP3 and PIP2, with no effect on phosphoinositide levels by ICAM1 (Figure 25B). Although, ICAM-2, -3, CD44, and CD43 were found to activate AKT in primary CD4⁺ and CD19⁺ cells to varying extents (Figure 25B, lower panel), only ICAM-2 and ICAM-3 effectively blocked TNF α induced apoptosis (Figure 25C) and Fas induced apoptosis (data not shown) in CD19⁺ B cells, indicative of functionally distinct properties possessed by ICAM-3, CD44, and CD43. The ICAM-2 anti-apoptotic effect was abrogated by incubation with Psitectorigenin and Y-27632 in CD19⁺ human B cells (Figure 25E), verifying the Rhokinase and phosphoinositide dependency in primary human cells. Incubation with GTP γ S loading conditions and herbimycin A induced apoptosis in primary cells for unknown reasons (data not shown). Thus, ICAM-2 oligomerization leads to the apparent membrane recruitment of PI3-kinase, results in PIP3 production, and subsequent activation of AKT kinase, initiating a potent inhibition of apoptosis in Jurkat T cells, BaF3 pro-B cells, and human primary lymphoid cells.

25

ICAM-2 interactions with its natural receptor LFA-1 activates AKT

We decided to test whether ICAM-2 oligomerization by virtue of interacting with its natural receptor, LFA-1 was sufficient to activate the PI3K/AKT pathway as elucidated by antibody crosslinking experiments. We developed an overlay assay, using the ICAM-2 retrovirally transduced NIH3T3 cells as ICAM-2 presenting cells to LFA-1 + Jurkat cells, to see if mixing of these two cell populations would induce AKT activation in the ICAM-2 expressing cells. Pre-incubation of the ICAM-2 cells with LY294002 abolished the intrinsic AKT activity due to the consequences of ICAM-2 overexpression in these cells. Provided that PI3K inhibition for prolonged periods of time might present an irreversible inhibition, the overlay assay was designed to identify AKT activity in the ICAM-2 cells after one hour blockade of PI3K at a concentration (2 μ M) close to the IC₅₀ of the compound. This time period and concentration was found to block AKT activity in control experiments and be inducible upon ICAM-2

35

crosslinking (data not shown). The overlay assay takes advantage of our recently developed FACS based AKT assay, in conjunction with immunophenotypic markers to identify which specific cells in a heterogeneous mixture present AKT kinase activity. Jurkat cells express low levels of LFA-1 and no detectable levels of Mac-1, two endogenous receptors for ICAM-2 (data not shown). ICAM-2 cells express full length human ICAM-2, and these NIH3T3 derived cells express low levels of mouse ICAM-1, -2 and no detectable levels of mouse ICAM-3 (data not shown), interactions of which are thought to be insignificant with human LFA-1, given the species and stoichiometric ratios of human ICAM-2 to murine ICAM-1, -2 (see Figure 23C, data not shown). We mixed Jurkat cells with vector control and ICAM2 cells with conditions that included prior blocking of Jurkat surface LFA-1 and Mac-1 proteins by incubation with monoclonal antibodies (mAb). We identified the ICAM-2 cells by size differences and absence of the CD4 T cell marker. We observed that incubation of LFA-1 positive Jurkat cells activates AKT in ICAM-2 expressing cells and is inhibited by prior treatment of Jurkat cells with LFA-1 monoclonal antibodies when compared to Jurkat mixing with vector control cells (Figure 25E, Reviewers see attached manuscript similar experiment in heterogenous lymphoid populations). Prior treatment of Jurkat cells with Mac-1 mAb also inhibited AKT activation of ICAM-2 expressing cells, indicating the common $\beta 2$ -integrin subunit between LFA-1 and Mac-1 is necessary in ICAM-2 oligomerization by these heterodimeric receptors. Because the ICAM-2 induced signal had to overcome the transient PI3K inhibition in our conditions, we did not expect to see a robust activation of AKT, but rather relied on the cell-by-cell analysis to see if activation of AKT was inducible in the presence of LFA+ Jurkat cells and inhibited by LFA-1 monoclonal antibodies. Although additional studies are necessary to verify the activation state of LFA-1 and Mac-1, in addition to the extent of ICAM-2 oligomerization, the data suggests that a receptor induced ICAM-2 oligomerization is sufficient to activate ICAM-2s intracellular PI3K/AKT signal and attributes a physiological role for ICAM-2 as a signaling molecule in cell-to-cell contact. Wulfig et al have visualized ICAM-1 surface clustering upon cell-to-cell contact, supporting that LFA can induce ICAM surface oligomerization required for intracellular signaling by ICAM-2 (Wulfig and Davis, 1998; Wulfig et al., 1998). In a separate study, we have adopted the T cell/B cell mixing conditions of Wulfig et al, and find dual activation of the ICAM2 AKT signal and a novel LFA-1 mediated p44/42 MAPK activation measured simultaneously in leukocyte mixing reactions by FACS based kinesis assays (Perez and Nolan, submitted), further supporting an in vivo signaling role upon ICAM-2/LFA-1 contact.

Discussion

A model for ICAM-2 mediating a cell survival signal

Our results demonstrate that ICAM-2 ectopic expression in a wide variety of cell types, and ligation of endogenous ICAM-2 in lymphoid cells including primary CD19+ B cells can initiate a survival signal. ICAM-2 induced AKT activation is a survival signal that can render a cell resistant to chemical (staurosporine and etoposide), and physiological (TNF α and Fas ligand) apoptotic inducers and is contingent on ezrin activation, a tyrosine phosphorylation event requiring both src tyrosine kinase and Rho-dependent serine/threonine kinase. This results in PI3K recruitment to the plasma membrane, PIP3 production and subsequent activation of AKT, which is capable of initiating a variety of cell survival signals through phosphorylation of downstream targets.

10

Although interactions with critical membrane-cytoskeletal linker proteins α -actinin and ezrin was demonstrated through co-immunoprecipitation studies, we note apparent differences between the dependency of these linker proteins to relay the ICAM-2 survival signal in fibroblastoid and lymphoid cell types. ICAM-2's antiapoptotic effect was dependent on the 21 amino acid cytoplasmic domain known to bind α -actinin. Notably, α -actinin has been shown to interact directly with PI3 kinase through the p85 SH3-domain (Shibasaki et al., 1994) and was found to co-immunoprecipitate with PI3K in ICAM-2 overexpressing transduced NIH3T3 and associate with PI3K in ICAM-2 crosslinked Jurkat cells (Figure 24E). We did not observe phosphorylation of α -actinin (data not shown), but did observe ezrin phosphorylation as a function of CAM-2 crosslinking (Figure 23B), suggestive of an ICAM-2 induced ezrin activation mechanism.

20

The ICAM-2 cytoplasmic domain has been found to bind ezrin, a member of the ezrin/radixin/moesin (ERM) family of membrane-cytoskeleton linker proteins (Helander et al., 1996) and notably bound ICAM-2 in contrast to α -actinin in our experiments (Figure 22A). ERM-proteins regulate cell morphology, adhesion and growth by promoting membrane-cytoplasmic linkages (Tsukita and Yonemura, 1997) and include the tumor suppressor, merlin/schwannomin, implicated in neurofibromatosis type II. In addition, ezrin and other ERM proteins have been implicated in different diseases for their modulation of adhesion molecule distribution (Ichikawa et al., 1998; Stokowski and Cox, 2000; Turunen et al., 1998; Vinore et al., 1995). Interestingly, ezrin overexpression has been shown to induce ICAM-2 clustering (Helander et al., 1996) suggesting that inappropriate surface protein clustering and subsequent signaling can be strongly influenced by ERM proteins. Additional studies are needed to address if the deregulation of the ICAM-2/ezrin protein stoichiometry by mutually exclusive pathways can impart similar cell survival capabilities.

35

Ezrin has recently been reported to mediate adhesion dependent survival in epithelial cells through a direct interaction with PI3-kinase and pursuant AKT activation (Gautreau et al.,

1999). Here we report that ICAM-2 clustering induced ezrin tyrosine phosphorylation and enhanced threonine phosphorylation, concomitant with p85 and PI3K association in Jurkat T cells (Figure 23D and 24E). This was sufficient to translocate PI3K to the plasma membrane (Figure 23F) and initiate production of PIP3 as measured directly by single cell analysis within
5 primary cells (Figure 25B) and indirectly through a PDK-1 activity assay (Figure 23E). Although PI3K/AKT membrane localization, PIP3 production, ezrin phosphorylation, and PDK-1 activity was also observed in ICAM-2 overexpressing transduced NIH3T3 cells, the profound effects of ezrin tyrosine phosphorylation as induced by ICAM-2 clustering and inhibition of the ICAM-2 anti-apoptotic effect with Y-27632 and herbimycin A (Figure 23D, 24D) preferentially in
10 Jurkat cells lead us to propose that ICAM-2 mediates a cell survival signal through a Rho-dependent ezrin activation (Reviewers see Supplementary Figure 20B). This then would initiate PI3K recruitment to the membrane and resultant PIP3 induced PDK-1 and AKT activity. We postulate that the increased binding of ezrin to the positively charged amino acid cluster in ICAM-2s C-terminal is probably enhanced by the oligomerization of ICAM-2 as induced by
15 antibody clustering and LFA-1 interaction, since we did not detect ICAM-2 to be tyrosine phosphorylated. This could provide a mechanistic explanation as to why ezrin is strongly associated with ICAM-2 in overexpression models, but is only observed to associate with ICAM-2 upon ICAM-2 clustering of endogenous levels.

20 Current research focusing on the activating mechanisms of ezrin, report a variety of differences among different cell types. A two-step activation process has been proposed for ezrin (Bretscher et al., 2000). ERM proteins are believed to reside in a dormant closed conformation mediated by a Gto-N terminal interactions (Bretscher et al., 1997; Gary and Bretscher, 1995). Upon phosphorylation at threonine and possibly serine residues (Bretscher, 1989; Simons et
25 al., 1998), a conformational change unmask the F-actin binding sites in the C-terminal and the surface protein interacting sites in the N-terminal (Bretscher et al., 2000). RhoGDI (Rho-GDP-dissociation inhibitor) has been observed to bind only activated ERM proteins (Takahashi et al., 1998), suggesting that dissociation of Rho/RhoGDI is a prerequisite for Rho kinase activation (Straw et al., 1998; Yasui et al., 1998). Dissociation of RhoGDI is a
30 prerequisite for activation of Rhodependent pathways, including ROCK, and is dependent for GTP to GDP conversion.

Evidence exists for ezrin functioning as both an upstream regulator and as a downstream regulator of small GTP binding proteins, making elucidation of molecular requirements and
35 ordering of activation systems across cell types difficult (Kosako et al., 2000; Kotani et al.,

1997; Mackay et al., 1997; Matsui et al., 1998; Shaw et al., 1998). In the systems utilized here, we observed the ICAM-2 anti-apoptotic effect to be abrogated by Y-27632 and GTP γ S, suggesting the Rho-dependent kinases and GTP are necessary in relaying the ICAM-2 signal to PI3K. In addition, inhibitors of src/p62 ^{yes} similarly antagonized the ICAM-2 induced tyrosine phosphorylation. We did not find a dependency on PCK isozymes, as PKC isozyme inhibitors bisindolymaleimide I, bisindolymaleimide II, and staurosporine did not abrogate the ICAM-2 induced survival signal and suggest that the PCK α and PCK θ dependent threonine phosphorylation of ezrin for its activation as reported by others contributes minimally in this system (Ng et al., 2001; Pietromonaco et al., 1998). We do note an increase of threonine phosphorylation of ezrin as a function of ICAM-2 crosslinking (Figure 23D) but its presence at the zero time point and difference observed in the fold induction of tyrosine phosphorylation of ezrin lead us to believe that although the threonine phosphorylation state may be contributory zrin tyrosine phosphorylation is responsible for relaying the ICAM-2 survival signal. Furthermore, site-directed mutagenesis of ezrin tyrosine residue Y353 to phenylalanine (Y353F) has been shown to abrogate PI3K binding in epithelial cells (Gautreau et al., 1999), suggesting that kinases involved in phosphorylating Y353 may be key regulators in ezrins capability to activate the PI3K/AKT pathway. Additional tyrosine residues (Y154 and Y353) have been observed to be phosphorylated by several membrane proteins possessing tyrosine kinase activity, namely platelet derived growth factor receptor (PDGF-R), epidermal growth factor receptor (EGF-R3 and hepatocyte growth factor (HGF); however, mutants lacking both tyrosine residues are still observed to be tyrosine phosphorylated, with various augmentation in signaling properties (Bretscher et al., 2000; Crepaldi et al., 1997; Jiang et al., 1995). This indirectly suggests that ezrin's multiple tyrosine phosphorylation sites may possess distinct and unique signaling properties within different cells, suggestive of a potential explanation on how ezrin can integrate and respond differently to a variety of signals, either through direct interactions with key signaling molecules or indirectly via adapter proteins. The identification of protein kinases responsible for specific tyrosine residue phosphorylation remains controversial, although preliminary studies for tyrosine phosphorylation at Y353 have pointed to src and p62^{yes} in vitro (Dumenil et al., 2000; Moller et al., 1994). We see ablation of ezrin tyrosine phosphorylation in the presence of herbimycin A in vivo, an inhibitor of both src and p62 ^{yes}, and further studies are needed to differentiate between these two src family protein tyrosine kinases and specific phosphorylation at Y353 or potentially other sites.

Our data suggest that ROCK is involved in the activation of ezrin that leads to its phosphorylation by src/p62 ^{yes} since incubation with Y-27362, a specific ROCK inhibitor, abolished tyrosine phosphorylation of ezrin. We had noted that ezrin threonine phosphorylation, though initially present, was intensified upon ICAM-2 ligation contrasting the induction of ezrin

tyrosine phosphorylation (see Figure 23D), and its ablation in the presence of inhibitors of Rho-dependent processes (see Figure 23C). Although this suggests that src/p62^{yes} tyrosine phosphorylation of ezrin is upstream of a Rho-dependent threonine phosphorylation, further studies are needed to conclusively delineate the temporal kinetics of ezrin activation as induced by ICAM-2 oligomerization. The ICAM-2 PI3K/AKT survival signal requirements for Rho-dependent kineses and phosphoinositides is apparent in Jurkat cells and in primary B cells (Figure 24D, 26C), however these requirements are less constraining in ICAM-2 overexpressing cells (data not shown, Reviewers see Supplementary Figure 20B) illustrating differences between the ICAM-2 overexpression and ICAM-2 clustering induced activation of the PI3K/AKT pathway.

The PI3K/AKT signaling system is a general mediator of extracellular stimuli, including growth factors, cytokines, as well as adhesion to extracellular matrices (Downward, 1998). The activation of AKT is dependent on phosphorylation at two sites, threonine-308 by PDK-1 and serine-473 by an unidentified PDK-2 (Downward, 1998). The amino terminus of AKT contains a pleckstrin homology (PH) domain that is thought to bind directly to phospholipid products of PI3K. This binding recruits AKT to the membrane, which has been proposed to induce a conformational change allowing phosphorylation at threonine-308 and serine473 by PDKI and PDKII respectively (Alessi et al., 1996). Dually phosphorylated AKT is then active and can phosphorylate a number of downstream effectors contribute to cell survival that include, apart from those investigated here, human caspase 9, and eNOS (endothelial cells) which promotes angiogenesis of vascular endothelium (Cardone et al., 1998; Kureishi et al., 2000) AKT hyperactivity has been observed in a number of disorders (Haas-Kogan et al., 1998) (Kulik et al., 1997; Shan et al., 2000; Wick et al., 1999) and therefore, elucidating mechanism that initiate the activation of AKT may be of therapeutic interest. Here, we demonstrate that ICAM-2 induced activation of AKT kinase results in the activation of several downstream effectors as detected by phosphorylation of BAD, GSK3, FKHR and AFX, all of which can contribute to favor cell survival (Brunet et al., 1999; Graff et al., 2000; Pap and Cooper, 1998; Zha et al., 1996). These observations support that ICAM-2 activation would be expected to lead to an extremely powerful anti-apoptotic signal in a variety of cells. It is apparent that a major regulation of the PI3K/AKT survival pathway is conditional on the upstream activating events, since general PI3K activation is a commonly observed in a variety of cancers and tumor models (Blume-Jensen and Hunter, 2001; Roymans and Slegers, 2001). A graphical illustration of ICAM-2 activating the PI3K/AKT pathway is depicted in Figure 25F, although differences in

molecular mechanisms may exist at the ICAM-2/ezrin and ICAM2/ α -actinin interface as alluded to above.

The data presented here illustrates that molecular differences exist between the induction of a cell survival signal in an ICAM-2 overexpression and ICAM-2 clustering model and further suggest that these processes may be cell type dependent. The anti-apoptotic effects mediated by ICAM-2 overexpression and by ICAM-2 clustering illustrate that inappropriate ICAM-2 expression may confer resistance to apoptotic stimuli but more importantly, ICAM-2 oligomerization can initiate a cell survival signal upon contact with LFA-1 bearing cells. This activation of the PI3K/AKT pathway by ICAM-2 can be viewed as both anti-apoptotic, if a cell where in an environment that predisposed it to apoptose, or as a cell survival signal that is mediated by cell-to-cell contact. As an adhesion molecule, ICAM-2 has been attributed a role in lymphocyte recirculation and a role in enhancing and stabilizing T-cell:B-cell contact via interactions with its receptor LFA-1 (Damle et al., 1992; Douglas et al., 2000). Here, we suggest that in addition to possessing adhesion molecule properties, ICAM-2 is capable of transmitting an intracellular signal when LFA-1 engagement clusters it on the cell surface. The significance of ICAM-2 clustering is twofold (1) by virtue of its overexpression it could lead to apoptotic resistance of cells otherwise destined to die by apoptosis; (2) receptor-mediated engagement of ICAM-2 can mediate cell survival selectively to cells that engage LFA-1 bearing cells, a process that occurs at the immunological synapse (Perez and Nolan, manuscript in preparation). We speculate that ICAM-2 clustering, as induced by antibody crosslinking or by engagement of its natural receptor LFA-1 presents a regulateable cell survival signal that is only initiated when an ICAM-2 cell comes in contact with an LFA-1 expressing cells. Such cell-to-cell contact is of functional necessity in T cell activation (Perez and Nolan, manuscript submitted) and may be important in other ICAM-2 / receptor interactions such as those recently reported between ICAM-2/DC-SIGN interactions in lymphocyte extravasation and ICAM/LFA-1 enhanced HIV infection/transmission (Barbeau et al., 1998; Butini et al., 1994; Geijtenbeek et al., 2000; Hioe et al., 2001). Thus, differential physiological roles for CAM-2 may exist. ICAM-2 overexpression could lead to tumor drug resistance and cytoproliferative disorders such as B-cell chronic lymphatic leukemia (B-CLL) since it has been reported that ICAM-2 is highly expressed on CD5⁺ B cells from B-CLL patients (Molica et al., 1996) and on large B-cell lymphomas derived from patients (Alizadeh et al., 2000). These observations, in conjunction with the work described, here proposes that ICAM-2 or other adhesion molecules may have a role in certain classes of leukemias. In normal fibroblastoid cells, ICAM-2 mediated PI3K activation may likely regulate the endothelial transmigration of eosinophils (Gerwin et al., 1999) and potentially dendritic cells (via DC-SIGN) (Geijtenbeek et al., 2000), in addition to lymphocyte extravasation, two pivotal cell-contact dependent processes.

Our results illustrate that ICAM-2 overexpression and clustering can provide an anti-apoptotic signal in cell lines and primary human B cells rendering them resistant to physiological death programs induced by TNF α and Fas ligand. Although ICAM-3, CD43, and CD44 induced AKT activity in primary cells as determined by single cell analysis, only ICAM-2 and ICAM-3 were able to block apoptosis to FAS and TNF α , with CD44 and CD43 actually perpetuating cell death. ICAM-3 crosslinking intensified PIP2 production in addition to a temporal delay in AKT activation (data not shown), which differed from ICAM-2 crosslinking and suggests that ICAM-3 is affecting other signaling pathways. Surprisingly, ICAM-1 was completely dissimilar to ICAM-2 in the experiments conducted. These observations illustrate that molecular differences exist in intracellular signaling pathways for adhesion molecules capable of binding the same ERM proteins and interacting with at least one shared receptor. Interestingly, it has been reported that inflammatory cytokines can modulate ICAM-2 expression on antigen presenting cells (APC) and that dendritic cell maturation upregulates expression of ICAM-3 (Maki et al., 1998; McLaughlin et al., 1998; McLaughlin et al., 1999; Ohh and Takei, 1996), suggesting that the similarities of ICAM-2 and ICAM-3 observed may have functionally similar roles in vivo. The work presented herein attributes a previously unrecognized function for ICAM-2. The finding that ICAM-2 can exert cell survival signaling, ectopically and endogenously, as well as elicit an anti-apoptotic phenotype in a variety of cell lines and human primary cells suggests that deregulation or induced clustering is likely to contribute significantly to the anti-apoptotic phenotype that leads to tumor progression and/or cell survival.

Development of anti-metastatic approaches targeted to block cancer cell adhesion by general blockade of adhesion molecules warrants caution as the development of such agents may have detrimental effects in normal physiological process that are dependent on cell-to-cell contact interaction such as those mediated by ICAM/LFA-1. Thus, pathways involved in the regulation of ICAM-2 or other adhesion molecule expression, distribution, and activation could be alternative targets to the development of anti-tumor therapies and potentially as immunosuppressants.

EXPERIMENTAL PROCEDURES

Antibodies and Chemicals

The following antibodies were purchased from Cell Signaling Technologies (Beverly, MA): anti-phospho-AKT-ser473 mAb, anti-phospho-AKT-thr308, anti-AKT, anti-phospho FKHR, anti-FKHR, anti-phospho-Gsk3 alp21, anti-cleaved PARP, anti-cleaved caspase 9, anti-cleaved caspase 3, anti-cleaved caspase 7, anti-caspase 9, anti-caspase 7, anti-pAFX, anti-PARP,

anti-BAD, anti-BADser112, anti-BADser136, anti-phospho-BADser155, anti-rabbit-HRP. Anti-ICAM-2 IC2/2 mAb, anti-ICAM-2-FITC mAb (IC2/2) were purchased from Research Diagnostics (Flanders, NJ). Anti-ICAM-2 N-terminal, anti-ICAM-2 Cterminal, anti-a-actinin, anti-Bcl2, anti-Bclx/s, anti-p27, anti-p21, anti-cFLIP, antiMYC, anti-p53, anti-NFKB, anti-Ikk α , anti-Ikk β , anti-phospho-tyrosine, antiphospho-threonine, anti-mouse IgG-HRP, anti-goat-HRP, annexin-V-biotin, were purchased from Santa Cruz Biotechnologies (Santa Cruz, CA). Anti-ezrin mAb, anti-PI3K, were purchased from Transduction Laboratories (San Diego, CA). Anti-PDK-phospho-serine, anti-PDK-phospho-threonine, Y-27362, U0126, bisindolymaleimide I and II, psi-tectorigenin, etoposide, PD98059, LY294002, TNF α , IL-2 were purchased from Calbiochem (San Diego, CA). Staurosporine, wortmannin, acridine orange, ethidium bromide, propidium iodide, GTP γ S were purchased from Sigma (St. Louis, MO). Anti-mouse-alexa fluor 568, anti-rabbit alexa fluor 488, anti-mouse alexa fluor 488, anti-rabbit alexa fluor 568, Prolong antifade, annexin-633, annexin-488, streptavidin alexa fluor 488, streptavidin alexa fluor 633, anti-phosphoinositide 3,4,5 triphosphate, anti-phosphoinositide 4,5 bisphosphate, were purchased from Molecular Probes (Eugene, OR). Annexin-V-FITC, anti-rabbit PE, anti-mouse-FITC, anti-goat FITC, anti-goat texas red, Streptavidin-PE, were from BD Biosciences (San Jose, CA). CD4-APC, CD19-PerCP, anti-ICAM-1, anti-CAM-3, anti-CD43, anti-CD44, ANTI-LFA-1, anti-Mac1, were purchased from PharMingen (San Diego, CA). Additional reagents are described where appropriate.

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Retroviral cDNA screening

Production and infection of retroviruses was carried out as described (Kitamura et al., 1995; Onishi et al., 1996; Rayner and Gonda, 1994). A cDNA library was prepared from Jurkat T cells in the pBabe-S vector (Hitoshi et al., 1998). The library (2×10^6 primary transformants) was used to transfect $\sim 10^7$ PHOENIX-E packaging cells. The pBMN-LacZ control vector was spiked into the library at 1:10 to monitor the transfection/infection efficiency. Viral supernatants were harvested and used to infect $\sim 10^7$ exponentially growing NIH3T3 cells; 48 hours post infection the cells were split and analyzed for β -galactosidase activity. The library-transduced NIH3T3 cells were plated at $\sim 25\%$ confluency; 24 hours later the medium was replaced with DMEM containing 1 μ M staurosporine (STP). After 24 hours treatment the STP-containing medium was replaced with complete DMEM/10% donor bovine serum. Surviving cells were allowed to reach $\sim 80\%$ confluency (7 days). Survivor cells were retreated as described two additional rounds.

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Phenotypic transfer assay

An aliquot of the surviving library-infected cells were super-infected with Moloney Murine Leukemia Virus (gift of T.Kinsella, Stanford). Super-infected cells were passaged for

two-weeks to allow efficient spread of the helper virus. Naive NIH3T3 cells were infected with viral supernatants and allowed to express for 48 hours. Re-infected cells were then treated with staurosporine as described above. Surviving cells were cultured for 7 days and analyzed for transfer by RT-PCR.

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RT-PCR cloning of proviral-cDNA inserts and construct generation

RNA was prepared by the acid-guanidium-phenol-chloroform method as described (Chomczynski and Sacchi, 1987). cDNA was prepared from 4 µg heat-denatured total RNA in a 50 µl reaction volume (Promega, Madison, WI) and incubated at 42°C and 55°C for 45 minutes.

- 10 The cDNA reaction products (5 µl) were amplified (primers: 5'-GATCCTCCCTTTATCCAG; 5'-GAATGAAAGACCCACCTGT) in a 50 µl PCR reaction mix (Perkin-Elmer, Foster City, CA) at 95°C/30sec; 55°C/60sec; 72°C/2min for 25 cycles. Full length ICAM2 cDNA was cloned into retroviral vector PBM-Z-IN backbone (Kinoshita et al., 1998) at BamHI/Sal1 site. ICAM2- ΔC, ICAM2-ΔN and ICAM2-C-terminal scrambled were generated by PCR and cloned into the
- 15 BamHI/Sal1 site.

Cell culture and preparation of primary cells

NIH3T3 murine fibroblast were maintained in DMEM, 10% DCS, 1% PSQ (Duelbecco Modified Eagle Media, 10% Donar calf serum, 1 % penicillin-streptomycin (1000 units/ml and 2mM L-glutamine PSQ). Jurkat T-cells and CH27 transformed B-cells were maintained in

20 RPMI-1640, 10% FCS, 1% PSQ. BaF3 pro-B-cells were maintained in RPMI-1640, 10% FCS, 1% PSQ, 400 U/ml IL-3 (PeproTech). 70Z/3 pre-B-cells were maintained in RPMI-1640, 10% FCS, 1 % PSQ, 50 µm β-mercaptoethanol. HL60 myeloid leukemia cells were maintained in Opti-Mem I, 10% FCS, 1% PSQ. 293T human fibroblast cells were maintained in DMEM, 10% FCS, 1% PSQ. Cells were maintained at 5% CO₂ /37°C humidified incubator. Transduced cells were maintained in 500 µg/ml G418 (Sigma) and proper expression of construct was verified by western blots and FACS analysis of full length ICAM-2, ICAM-2 ΔC, ICAM-ΔN as indicated. Staurosporine treatment was for 24 hrs at 1 µM (unless otherwise indicated). LY294002 treatment was at 10 µM 30 minutes before any subsequent treatment. Wortmannin treatment

30 was for 24 hrs at 100 nM. Chemicals were dissolved in DMSO (final being 1:1000 for solvent dilution) and vector controls were incubated with 1% DMSO as negative control. Vector controls consisted of pBMN-Z-IN (empty vector) transduction or pBMN-Z-IN-GFP as indicated. The infection frequency of pBMN-Z-IN-GFP was 48% ± 10 for three independent viral transductions. Continual neomycin selection yielded homogenous ICAM-2, ICAM2- ΔC,

35 ICAM2-ΔN, or control vector populations as routinely monitored by flow cytometry for protein

expression. For preparation of primary cells, mononuclear cells were isolated from human peripheral blood by Ficoll-plaque density centrifugation and depletion of adherent cells on adherent plastic culture dishes. Isolated cells were maintained in complete media and analyzed by FACS for CD4, CD3, CD19, and ICAM-2 expression.

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Cross-linking

Antibody cross-linking experiments were performed using mAbs to anti-ICAM-1, anti-ICAM-2, anti- ICAM-3, anti-CD43, or anti -CD44 that recognizes the N-terminal region (extracellular domain) at 10 µg/ml. Spin dialysis (Biorad) was used for buffer exchange of azide containing antibodies (0.01%) to phosphate buffered saline buffer pH 7.4. Anti-mouse IgG (Sigma) was used as a control antibody in cross-linking experiments. 1×10^6 Jurkat, 1×10^6 BaF3, ar 1×10^7 PBMC cells were serum starved for 4 hours respectively (except PBMC). Cells were incubated with either mAb (10 µg/ml) or mouse IgG (10 µg/ml) at 37°C for the indicated time. Time points began post serum starvation to attenuate most of the signaling pathways or as indicated. Serum starvation of 4 hrs lacked biochemical initiations of apoptosis as tested by caspase 3 activity, cell cycle analysis, annexin V/PI staining (data not shown). Cells extracts were taken and subjected to either immunoprecipitation or immunoblotting were appropriate. Primary cells were subjected to apoptotic treatment followed by preparation for flow cytometry. GTPγS was made cell permeant by pre-incubating compound with Eugene (Roche Biochemicals) in a 1:4 molar ratio for 30 minutes at 25°C prior to being applied to cells. Liposome mediated delivery of radiolabeled/fluorescent triphosphates yields high cell incorporation of highly charged molecular species (Perez, O,D unpublished results).

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Apoptosis assays

Apoptosis was determined either by counting pyknotic nuclei, annexin-V binding by flow cytometry, or TUNEL BrdU staining by flow cytometry as indicated. Apoptosis was induced either by staurosporine (1 µM), treatment with anti-mouse Fas (5 µg/ml Santa Cruz Biotechnology (SCB), anti-human Fas (5 µg/ml Jo2 Biomed) or anti-human Fas (10 ng/ml CH11 Kamiya Biomedical) as indicated in appropriate experiment, IL-3 withdrawal (24 hrs) or as indicated. For pyknotic nuclei count, NIH3T3 cells were grown on glass coverslips and assayed for apoptosis. Suspension cells were analyzed by nuclear staining with 25 µM acridine orange/25 µM ethidium bromide or 5 µg/ml Hoechst 33342 (Molecular Probes, Eugene, OR). Apoptosis was identified by the presence of characteristic pyknotic nuclei as described (Jacobson and Raff, 1995). At least 5 individual fields were photographed and counted using a Zeiss axioscope fluorescence microscope. For the survival assay 10^6 NIH3T3 cells were plated in 10 cm dishes and incubated overnight at 37°C. The cells were treated with 1 µM staurosporine for 24 hours. The cells were washed and cultured for 24 hours in complete

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medium. The cells were trypsinized and viable cells counted following trypan blue staining. Untreated cells were counted at the time of treatment. % survival represents (number of viable cells post-treatment)/(number of viable cells pretreatment). FACS based annexin-V-FITC/PI staining was done as described (PharMingen). TUNEL assay was performed as described by
5 manufacturer of Apoptosis BrdU kit (Promega). Flow cytometry data acquisition was performed 24 post staurosporine treatment (1 μ M) for both BrdU (10,000 events collected) and pyknotic nuclei count and 12 hr post-staurosporine treatment for annexin V binding (100,000 events collected). Annexin-V positive cells will have compromised membranes after 24 hr treatment. Flow cytometry analysis was performed on a BD FACSCalibur machine with CELLQuest and
10 analyzed using FlowJo software (Tree Star).

Kinase assays

PDK-1 activity was determined by a PDK-1 immunoprecipitation kinase assay kit (Upstate Biotechnology, Lake Placid, NY) as recommended by manufacturer. In brief, PDK-1 was
15 immunoprecipitated from 1×10^6 treated cells, incubated with inactivated SGK enzyme, and detection of (γ - 32 P) ATP incorporation to a SGK peptide substrate was measured by scintillation counting. PI3K activity was detected by developed PI3K FACS based assay (see below). AKT activity was detected by immunoprecipitation of AKT1 from cells and used in a kinase assay with GSK3 fusion protein (CST). 2×10^6 NIH3T3, 2×10^6 Jurkat, or 5×10^5 BaF3 cells were
20 incubated with immobilized AKT 1G1 monoclonal antibody (mAb) (1:200, CST) at 4°C with gentle rocking motion for 2 hrs. Immunocomplexes were washed 4X with cell lysis buffer and resuspended in 40 μ l kinase buffer (25 mM Tris pH 7.5, 5 mM p-glycerolphosphate, 2mM DTT, 0.1 mM Na_3VO_4 , 10 mM MgCl_2) supplemented with 200 μ M ATP and 1 μ g of GSK-3 fusion protein (CST) for 30 min at 30°C. Kinase reaction was terminated with SDS sample buffer
25 boiled for 5 min, and phosphorylation state of GS3K was detected by immunoblotting and visualized using ECL detection (Amersham). Immunoblots are representative of 3 independent virally transduced cell populations each repeated 3 times (n=9). AKT activity was verified by AKT phosphorylation by immunoblotting with phospho-specific antibodies to ser473 and thr308 (NEB). Kinase assays are representative of at least three independent measurements and are
30 represented as standard deviations were appropriate.

Cell Fractionation, Immunoprecipitations and Immunoblotting

Triton-X-100 fractionation was carried out as described (Ref xxx). Cell extracts were prepared by washing 2×10^6 cells in ice cold PBS and harvesting in lysis buffer (20 mM Tris pH 7.5, 150
35 mM NaCl 1 mM EDTA 1 mM EGTA, 1% Triton X100, 2.5 mM Na_2PO_4 , 1 mM β

-glycerolphosphate, 1 mM Na_3VO_4 , 1 $\mu\text{g/ml}$ Leupeptin, 1 mM PMSF, protease inhibitor cocktail tablet (Boehringer Mannheim)). Extracts were centrifuged 14,000 RPM for 5 min at 4 °C and cell lysates (20 μg as determined by BCA protein assay (Pierce)) were fractionated on 12% or 15% SDS-polyacrylamide gel electrophoresis and transferred to PVDF membranes using standard procedures. Immunoprecipitations were pre-cleared with protein A/G plus-agarose beads (SBC). IP was incubated for 1 hr or overnight with primary antibody, 1 hr with protein A/G plus-agarose beads and washed 4X with lysis buffer. Blots were incubated with the indicated antibodies. Secondary antibodies used: anti-rabbit HRP (1:5000 NEB), anti-mouse HRP or anti-goat HRP 1:5000, SBC) and visualized using ECL detection (Amersham) or with a Kodak Digital Science 440C station. Blots are representative of at least 3 independent experiments.

Flow Cytometry and FACS analysis

Intracellular and extracellular staining was performed as described (www.metazoa.com/UPL3287) using indicated antibodies. Intracellular probe for AKT activity was made by conjugating monoclonal anti-AKTser473 antibody (Cell Signaling Technology) to Alexa-568 dye (Molecular Probes), using Alexa-568 protein conjugation kit (Molecular Probes). Phospho specificity was tested by western blotting and FACS analysis to a variety of PI3K activators and inhibitors (data not shown, Perez and Nolan, submitted). Intracellular staining by phosphoAKTser473-Alexa568 reflected AKT kinase activity when NIH3T3 cells were stimulated with platelet derived growth factor (Sigma) and inhibited by LY294002 prior to stimulation. Intracellular probe for phosphatidylinositol 3,4,5 triphosphate, and phosphatidylinositol 4,5 bisphosphate was made by conjugating mAb to PIP3 or PIP2 to alexa fluor 488 or alexa fluor 633 using alexa fluor protein conjugation kits (Molecular Probes). PIP3 and PIP2 FACS based measurements reflected PI3K and PI4K kinase activity (Perez and Nolan, manuscript in preparation). Quantitative FACS analysis was performed as described (Davis et al., 1998; Iyer et al., 1998; Lenkei et al., 1998). In brief, phycoerythrin (PE) (Molecular Probes) was conjugated to a anti-ICAM2 N-terminal monoclonal antibody (Research Diagnostics IC2/2) as suggested by manufacturer of protein cross-linking kit (Molecular Probes), tested for proper stoichiometry (data not shown) and quantitated using Quantibrite-PE beads (BD systems). A quantitative calibration curve was generated using Quantibrite-PE beads that contain a known amount of PE molecule/bead. A linear regression analysis is performed using the following equation: $\log_{10}(\text{PE fluorescence}) = \text{slope} \cdot \log_{10}(\text{PE molecules/bead}) + \text{intercept}$. PE fluorescence is determined by taking the geometric mean of the PE channel. Quantitation is valid only for antibodies directly conjugated to PK. Using saturating amounts of antibody and thorough washing ensures all surface antigens are bound. Preparation of samples on ice reduces antibody internalization. Surface antigens of cells for an unknown cell population are determined by computing the PE geometric mean using the calibration curve. Quantitative

FACS analysis provides approximate estimates for surface antigen expression. Numbers plotted represent relative surface molecules and not absolute numbers since antibody valency was not investigated for the monoclonal antibodies used. Flow cytometry analysis was performed on a BD FACSCalibur machine and analyzed using FlowJo software (Tree Star).
5 CellQuest was used for quantitative flow cytometry and linear regression analysis of Quantibrite-PE beads. Flow cytometry data are representative of 3 independent experiments of 10^6 cells/sample analyzed. 100,000 events were collected or otherwise noted and calibrated using Calibrite beads (BD systems). Data plotted in bar graph format is expressed as the mean (bar) \pm SD of triplicate experiments.

Laser Scanning Confocal microscopy

Cells were grown in coverslips, washed twice in phosphate buffered saline pH 7.4 (PBS) and fixed in 2.7% paraformaldehyde (in PBS). Cells were permeabilized for 5 minutes with 0.1% Triton-X-100 washed twice in PBS, blocked in 4% bovine serum albumin (BSA, in PBS), and
15 subjected to antibody incubation: (primary at 0.1 mg/ml, secondary at 1:1000 dilution in 1% BSA, with several washing steps in between). Stained coverslips were mounted onto glass slides with Prolong Antifade reagent (Molecular Probes) and visualized using a Molecular Dynamics Multiprobe 2010 confocal laser scanning microscope. Images were compiled using Adobe Photoshop 6.0.

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5 Example 5

Activation of PKB/AKT-Dependent Cell Survival by Intracellular Adhesion Molecule-2 (ICAM-2)

In this Example, using the methods and compositions of the present invention, the present
10 inventors (also referred to herein as "we") using a retroviral cDNA library in a functional genetic
screen identified intracellular adhesion molecule-2 (ICAM-2), a member of the immunoglobulin-
family of adhesion molecules, as a potent inhibitor of several activators of apoptosis. ICAM-2
expression blocked the onset of apoptosis in several cell types and induction settings.
ICAM-2's anti-apoptotic effect was mapped to the activation of the PI3K/AKT pathway and
15 resulted in subsequent phosphorylation of BAD, GSK3, and FKHR. ICAM-2's cell survival
signal was found to be independent of caspase inhibitory proteins, cell cycle regulators, or
anti-apoptotic pathways using Bcl-2. The survival function was dependent on the cytoplasmic
tail of ICAM-2, a region that bound membrane-cytoskeleton linker proteins α -actinin and ezrin.
The ICAM-2 mediated survival function was abrogated by two pharmacological inhibitors of P13
20 kinase, wortmannin and LY294002, indicating activation of AKT and its downstream effectors
were dependent on P13K recruitment to the plasma membrane. Antibody clustering of
endogenous ICAM-2 on BaF3 pro-B cells and Jurkat T-cells stimulated AKT kinase activity and
phosphorylation of downstream effectors BAD, GSK3, FKHR, and AFX, congruent with the
ectopic expression results. Primary CD4+ and CD19+ cells were protected from TNF α and Fas
25 mediated apoptosis following ICAM-2 clustering and presented elevated AKT activity as
detected by direct single cell flow cytometric measurement of intracellular AKT kinase activity.
These results attribute a novel survival signaling function to ICAM-2 that might provide an
explanation for both the role of ICAM-2 over-expression in B-cell lymphomas and mechanisms
by which ICAM-2 might signal intracellular communication in a variety of important cell types.

30

Introduction

Cellular homeostasis is achieved through a tightly regulated balance of proliferation and
apoptosis. Apoptosis is a distinct form of cell death that is initiated by a number of events
including DNA damage, growth factor deprivation, receptor-mediated death signaling, or by
35 xenobiotic induction, among others. Regulation of apoptotic signaling pathways is largely

governed by a cell's ability to sense its environment. Cellular apoptotic death can occur by many paths with distinct apoptotic stimuli initiating varied cell death mechanisms. For instance, chemically induced apoptosis occurs with the release of cytochrome C and activation of caspase 9 (Budihardjo et al., 1999). Receptor induced apoptosis (Fas/TNF α) is mediated through activation of caspase 8 (Holmstrom and Eriksson, 2000; Kruidering and Evan, 2000; Rath and Aggarwal, 1999; Schneider and Tschopp, 2000) and can operate independent of the mitochondria (Gottlieb, 2000; Kroemer, 1999; Kuida, 2000). Both caspase 8 and 9 converge on effector caspases 3 and 7 to mediate the characteristic hallmarks of apoptosis (Earnshaw et al., 1999; Grutter, 2000).

A cell's commitment to cellular proliferation or programmed cell death is a balance of the survival and death signals communicated from both the immediate environs, such as extracellular matrix (ECM) interactions, cell-cell contact, or more distant sites through endocrine and paracrine factors (Ruoslahti and Reed, 1994). Survival pathways are proposed to function during early development where apoptosis plays a key role in shaping organogenesis (Raff, 1992). Similarly, the lack of survival signals is exemplified in the phenomenon of anoikis, where epithelial cells undergo apoptosis when ECM-attachment is blocked (Frisch and Ruoslahti, 1997). ECM survival signals are presumably mediated by activation of cell-surface receptors via integrin clustering, contributes to the specificity of cell-cell/cell-matrix interactions, and prevents unwarranted cell migration. ECM-interactions are required for growth factor signaling in adherent cells (Hulleman and Boonstra, 2001). Deregulation of ECM-integrin signaling leads to anchorage-independent growth, which correlates with malignancy. Hence it is clear that deregulation of ECM signaling can desensitize a cell to apoptotic stimuli and contribute to a malignant phenotype that allows cells to survive in foreign milieus and under adverse conditions.

Functional characterization of molecules involved in cellular adhesion has been focused on signal transduction pathways attributed to integrins and cadherins (de Fougères et al., 2000; Schneller, 2001; Shinohara et al., 2001). Adhesion molecules of the immunoglobulin (Ig) superfamily have been largely overlooked as potential signaling molecules. ICAM-2 (CD102) is a member of the Ig-superfamily of cell surface proteins and mediates leukocyte binding to LFA-1 (CD11a/CD18) and MAC-1 (CD11b/CD18) whose expression is restricted to leukocytes (van Kooyk and Figdor, 2000). As a cell surface adhesion member involved in leukocyte recruitment in tissues, ICAM-2 is expressed in low levels on most leukocytes including T and B lymphocytes, monocytes, platelets and early CD34+ hematopoietic progenitor cells (de Fougères et al., 1991; Diacovo et al., 1994; Nortamo et al., 1991). In addition, ICAM-2 constitutive expression has been observed on all vascular endothelium (de Fougères et al.,

1991). Based on the expression pattern of ICAM-2, it has been hypothesized to be involved in leukocyte recirculation. ICAM-2 deficient mice support this by demonstrating that eosinophil trafficking is augmented in inflammatory responses (Gerwin et al., 1999). Surprisingly, this phenotype is relatively mild considering the range of cell types upon which ICAM-2 is expressed. It suggests that the roles ICAM-2 plays are subtle or are redundant with other pathways. Thus, the ICAM-2 deficient mice do not sufficiently detail the roles of ICAM-2, warranting further characterizations of the mechanisms by which ICAM-2 acts are needed.

In contrast to ICAM-2's putative role as an adhesion molecule, it is surprising to find ICAM-2 highly expressed on both endothelial cells in lymphomas and CD5⁺B cells from B-cell chronic lymphatic leukemia (B-CLL) patients (Molica et al., 1996; Renkonen et al., 1992). Lymphoproliferative diseases inevitably inhibit adhesion dependency-loss of adhesion is one of the primary means by which metastatic spread occurs. However, recent gene expression profiling of large B-cell lymphomas from patients indicated an elevated expression of ICAM-2 (Alizadeh et al., 2000). Interestingly, ICAM-2 is the only ICAM (1-5) that maps to chromosome 17q23-25 (Sansom et al., 1991), a segment associated with genomic instability and recently identified as a segment of high aberration in various cancers (Dion et al., 2000; Dobo et al., 1995; Russell et al., 2000; Sugai et al., 2000). Despite these observations, ICAM-2's role and molecular characterization in the progression of lymphatic disease states has not been addressed.

We devised a genetic screen to identify genes that are involved in regulation of apoptosis and augmented in malignant phenotypes. Cells transduced with a human transformed T cell (Jurkat) retroviral vector cDNA library were screened for an anti-apoptotic phenotype. One cDNA demonstrated reproducible anti-apoptotic activity in a phenotypic transfer assay. This cDNA encoded full-length intracellular adhesion molecule-2 (ICAM-2). Here we show that ICAM-2 was identified as being capable of transducing an anti-apoptotic effect. Functional characterization of ICAM-2 reveals that it mediates a survival signal sufficient to block apoptosis by activation of the PI3K/AKT pathway. Furthermore, development of probes to measure intracellular AKT activity by flow cytometry allowed for the simultaneous measurement of AKT activity as a function of ICAM-2 clustering on human peripheral blood monocytes (PBMC). Multi-parameter FACS analysis confirmed ICAM-2 clustering led to AKT activity in CD4⁺ and CD19⁺ primary human cells and subsequent protection from Fas and TNF α mediated apoptosis. These observations identify ICAM-2 as having a role in mediating cell survival by activation of the PI3K/AKT cell survival pathway. These finding suggest that

inappropriate expression of ICAM-2 could contribute significantly to the anti-apoptotic phenotypes in the cumulative process that leads to tumor progression and metastatic spread. The findings also underscore a positive role for ICAM-2 in signaling pathways involving AKT and downstream effectors.

5

RESULTS

Screening a retroviral cDNA library in an anti-apoptotic functional assay

To identify genes that alter signaling in apoptosis we devised a retroviral library approach to screen for cDNAs that encode anti-apoptotic molecules (Figure 26A). cDNAs from a malignant cell type (Jurkat T cell leukemia) were screened for anti-apoptotic function in a recipient non-malignant (fibroblast) cell. Apoptosis was induced with staurosporine (STP), a kinase-inhibiting plant-derived alkaloid, activity, and a potent inducer of caspase-dependent cell death in most cell types. Induction conditions were determined that limited the level of spontaneous background while maximizing the extent of apoptotic death (minimal background of one surviving cell in a STP-treated mock library control in $\sim 10^4$ starting cells). 10^7 NIH3T3 fibroblasts were infected with a retroviral cDNA library derived from Jurkat cells at an infection frequency of 40% to limit the number of integrations to a single event per cell. Control retroviruses were prepared, expressing LacZ- or Bcl-2, and infected into a similar number of NIH3T3 cells. Apoptosis was induced in the transduced cell culture by staurosporine treatment and the cells were then allowed to recover (Figure 26A). Surviving cell clones were expanded, replated and retreated with staurosporine. The complexity of the expressed cDNAs in the surviving population was assessed with RT-PCR. A few major bands were observed following three rounds of selection (Figure 26 B).

Replication-competent Moloney murine leukemia virus (MMLV) was applied to the enriched cell populations. Upon integration and expression of the MMLV proteins resident retroviral constructs from the library were co-packaged into infectious virions that can be transferred to naïve target NIH3T3 cells. The staurosporine selection process was then reapplied. In this manner true phenotype-inducing clones enrich at a rate of 1/background (see materials and methods for elaboration). Three of the pooled libraries showed enrichment for specific bands after PCR (see Figure 26B lanes 1 and 2). Several bands were rescued by PCR cloning and one of these bands was sequenced and demonstrated to encode full-length ICAM-2 cDNA (Staunton et al., 1989), an immunoglobulin superfamily member that regulates leukocyte adhesion through interaction with its integrin counter-receptor, LFA-1 (de Fougerolles et al., 1991; Simmons, 1995).

ICAM-2 mediates a survival signal

The ICAM-2 cDNA was cloned into a retroviral vector (pBMN-Z-IN) that carries an internal ribosome entry site (IRES) upstream of the neomycin resistance gene. NIH3T3 fibroblasts were infected with retroviruses capable of expressing the ICAM-2 gene, along with appropriate control vectors pBMN-Z-IN (vector control) and pBMN-Z-IN-GFP (GFP vector control).
5 Expression of the 60 kD ICAM-2 glycoprotein in neomycin-selected NIH3T3 fibroblasts infected with the pBMN-Z-IN-ICAM-2 construct (referred to as ICAM-2) was verified by immunoblotting (Figure 27A), flow cytometry (Figure 27B, 28B), and immunoprecipitation (Figure 28C). The expressed ICAM-2 protein co-migrated with the native protein from Jurkat T cells (data not shown).

10 Staurosporine treatment is thought to mimic factor-withdrawal in cells (Raff, 1992), and induces apoptosis. ICAM-2 expression in NIH3T3 cells limited the extent of apoptosis induced by staurosporine treatment as determined by pyknotic nuclei count (Figure 27C, 27D) as well as an annexin-V binding assay (Figure 27E) and the BrdU TUNEL assay (Figure 29B) confirming
15 the clone's selection by phenotype in the screen. Interestingly, not all forms of apoptosis were inhibited. Fas induced apoptosis in ICAM-2 expressing Jurkat T cells was not inhibited across a range of concentrations of a monoclonal antibody to the Fas receptor (Figure 27C, panel 3).

A report correlating ICAM-2 expression with a lymphoproliferative disorder, B-cell chronic
20 lymphatic leukemia (B-CLL) (Molica et al., 1996), prompted us to test the anti-apoptotic activity of ICAM-2 in the mouse pre-B-cell line, 70Z/3. ICAM-2 expression inhibited staurosporine-induced apoptosis in 70Z/3 cells (Figure 28A). Given the indication that staurosporine mimics factor withdrawal, we assessed the ability of ICAM-2 expression to protect factor-dependent cells from factor withdrawal. ICAM-2 expression in the
25 IL-3-dependent pro-B cell line BaF3 resulted in a delayed onset of apoptosis following IL-3 deprivation (Figure 28A). Furthermore, we tested the ability of ICAM-2 to rescue BaF3 cells from apoptosis induced by treatment with anti-Fas antibodies. ICAM-2 demonstrated a potent inhibition of apoptosis through the Fas pathway in these cells (Figure 28A). These results are as good as or better than transduction controls with Bck2 and therefore the apparent effects
30 cannot be attributed to modest effects on apoptosis (data not shown). There is, however, cell specificity to the anti-apoptotic effect as ICAM-2 over-expressing Jurkat T cells were not protected from Fas-induced apoptosis (Figure 27C) but were protected from staurosporine induced apoptosis (Figure 28A). High throughput proteome analysis recently identified that in Jurkat T cells, Fas induced apoptosis results in a rapid destruction of 12 cytoskeletal proteins,
35 alters phosphorylation states of various chaperone proteins and cytoskeletal structural proteins,

and presents a more "aggressive" form of cell death than staurosporine induced apoptosis (Gerner et al., 2000). Therefore different cell death inducers have specific physiological characteristics in Jurkat T cells supporting the differences observed. In addition, effector caspases have been demonstrated to be dispensable in chemical induced apoptosis (Johnson et al., 2000).

The initial screening was performed in fibroblasts, and it is therefore unlikely that ICAM-2's anti-apoptotic effect was mediated through its lymphocyte-specific integrin ligand, LFA-1 (CDT 1a/CD18). To address this possibility we prepared an extracellular domain-deleted (ΔN) version of ICAM-2 and tested it in the BaF3 cells. Expression of the cytoplasmic domain was sufficient to impart full anti-apoptotic activity (Figure 28A). This indicates that the cytoplasmic domain of ICAM-2 provides a survival signal when over-expressed ectopically, and likely exerts its effect by mimicking receptor crosslinking. Since signaling through ICAM-family cytoplasmic domains is poorly understood, we sought to identify the signaling pathway by which it conferred resistance to apoptosis.

The ICAM-2 survival function maps to a membrane-cytoskeleton linker protein binding motif.

ICAM-2 had been regarded as an adhesion molecule and has been shown to be interconnected with the actin cytoskeleton by linker proteins α -actinin and ezrin, which bind the ICAM-2 cytoplasmic domain. (Heiska et al., 1996; Helander et al., 1996). To assess the potential role of α -actinin in mediating the ICAM-2 antiapoptotic signal, the α -actinin recognition sequence was changed to a scrambled sequence unable to bind α -actinin in vitro (Heiska et al., 1996). Retroviral constructs encoding ICAM-2, a C-terminal deletion mutant (ICAM2- ΔC) and a full-length ICAM-2 protein with the mutated α -actinin binding motif (ICAM2-C scrambled) were used to transduce NIH3T3 cells (Figure 28B). The infected cells were sorted by FACS using an ICAM-2 monoclonal antibody (IC2/2) recognizing only native N-terminal ICAM-2 (de Fougères et al., 1991). This verified that ICAM-2, ICAM2- ΔC , or ICAM2-C-scrambled proteins were properly folded and targeted to the outer face of the plasma membrane. The NIH3T3 cells expressing the ICAM-2 mutants were assayed for survival following both etoposide treatment (another potent inducer of apoptosis in NIH3T3 cells) and staurosporine treatment. ICAM-2 imparted potent long-term survival on NIH3T3 cells and the survival signal was lost when the C-terminal deletion of ICAM-2 (ICAM2- ΔC) was expressed in cells (Figure 28B). Mutation of the α -actinin binding site in the ICAM2 cytoplasmic tail also did not provide any protection against etoposide or staurosporine treatments. Furthermore, both α -actinin and ezrin co-immunoprecipitated only with full length ICAM-2 (Figure 28C), supporting the postulate that these cytoskeleton linker proteins were necessary to relay an anti-apoptotic signal.

ICAM-2 signals through AKT kinase

As survival signaling through many adhesion molecules involves PI3 kinase (Frisch and Ruoslahti, 1997; Khwaja et al., 1997), we analyzed the role of this pathway in the ICAM-2 anti-apoptotic signal. Pretreatment of ICAM-2 expressing BaF3 cells with the PI3 kinase inhibitor wortmannin blocked the ICAM-2 mediated anti-apoptotic effect (Figure 29A), implicating PI3 kinase in the ICAM-2 survival signaling cascade. Under these conditions vehicle-treated ICAM-2 expressing BaF3 cells were resistant to staurosporine induced apoptosis. Similarly, treating ICAM-2 transduced NIH3T3 with LY294002 abrogated ICAM-2's anti-apoptotic effect as measured by both annexin-V binding and BrdU TUNEL assay using flow cytometry (Figure 29A, 29B, and, reviewers see APPENDIX for flow cytometry plots).

PI3 kinase is known to regulate apoptosis through its downstream effector AKT (PKB) kinase (Downward, 1998). AKT kinase has a potent anti-apoptotic effect downstream of diverse apoptotic stimuli, including staurosporine (Marte and Downward, 1997) (data not shown). We assayed ICAM-2 expressing fibroblasts for AKT kinase activity. NIH3T3 fibroblasts expressing ICAM-2 showed an elevated level of activated AKT (Figure 30A). This increase in AKT kinase activity was not seen in ICAM2-ΔC or mock transduced cells. We further verified that this ICAM-2-dependent increase in activated AKT is maintained following STP-treatment using both kinase assays and by verifying phosphorylation of both ser473 and thr308 (Figure 30A). A target of AKT kinase is the pro-apoptotic Bcl2 family member, BAD. BAD is normally held in an inactive state by phosphorylation of serines 112 and 136, and is bound to 14-3-3. Upon induction of apoptosis, BAD is dephosphorylated at both ser112 and ser136, and dimerizes with Bcl-2. As a result, the equilibrium of Bcl-2/Bcl-X_L is shifted to favor release of BCL-X_L to mediate Cytochrome C release and apoptosis (Datta et al., 2000). BAD is dephosphorylated following staurosporine-treatment of ICAM2-ΔC- and mock-transduced NIH3T3 cells, concomitant with the induction of apoptosis (Figure 30B). However, ICAM-2 expression results in the maintenance of BAD-phosphorylation consistent (Figure 30B) with sustained AKT kinase activity (Figure 30A).

BAD has 3 phosphorylation sites, ser112 mediated by active p90Rsk1, ser136 mediated by active AKT, and ser155 mediated by both active PKA and active p90Rsk1. ICAM-2 and its role in connection to BAD ser112 phosphorylation is to be described elsewhere (Perez and Nolan, manuscript in preparation). Phosphorylation of BAD ser155 was not observed (data not shown). Another target of AKT kinase involved in anti-apoptotic signaling, GSK3, was found to be phosphorylated in ICAM-2 expressing NIH3T3 and probably contribute to the anti-apoptotic

effect (Figure 30B, bottom panel). The anti-apoptotic signal had no apparent effects on the anti-apoptotic proteins Bcl-2 and Bcl-X_L indicating that typical anti-apoptotic pathways did not mediate the survival signal (Figure 30C).

5 **ICAM-2 crosslinking confirms a role for endogenous ICAM-2 in providing an anti-apoptotic signal**

This ICAM-2-dependent elevation of activated AKT levels in fibroblasts is independent of ligand interactions as determined by the fact that ICAM-2ΔN can protect cells from apoptosis and ICAM2-ΔC cannot activate AKT. This is indicative of the signaling mechanism driven by
10 oligomerization of surface ICAM-2 molecules, a common *modus operandi* for adhesion molecules. The surface oligomers create interaction sites within their cytoplasmic domains, recruiting signaling molecules to participate in membrane proximal interactions. Quantitative FACS analysis revealed that the density of ICAM-2 molecules on the cell surface vary widely between cell types (Figure 30D, and, reviewers see APPENDIX for quantitative methodology;
15 see Experimental Procedures for elaboration) and correlated with varied expression patterns as previously described (de Fougerolles et al., 1991). Basal expression levels of ICAM-2 on lymphocytes was 24 orders of magnitude less than the enforced expression from the retroviral vector as determined by the number of ICAM-2 surface molecules. We therefore decided to test whether endogenous ICAM-2, when cross-linked, could confer the phenotypes observed
20 above.

To test this hypothesis in a physiological setting, we cross-linked (clustered) endogenous pro-B cell ICAM-2 surface molecules with monoclonal antibodies and assayed for activation of AKT kinase activity using GSK3 as a target. We first chose BaF3 cells as these displayed low
25 endogenous ICAM-2 levels (Figure 30D) and an anti-apoptotic phenotype upon ICAM-2 transduction (Figure 28A). As shown in Figure 31A, basal AKT activity is unaffected by incubation with isotype control sera. In contrast, AKT kinase activity increased 3.5-fold with fifteen minutes following addition of anti-ICAM-2 and resulted in the subsequent phosphorylation of BAD ser136, AKT ser473 thr308, and FKHR. This level of AKT kinase
30 activity is congruent with that measured in following IGF-1-treatment of BaF3 cells (A. Bartels and J.B.L., unpublished results). Though the results for AKT-activity were reproducible we were unsatisfied with the western blot analysis and so developed a new procedure for single cell analysis of AKT activity by flow cytometry, described below (Perez and Nolan, submitted, reviewers please see attached manuscript). Cross-linking ICAM-2 on a different cell type,
35 Jurkat T cells, also protected against staurosporine-induced apoptosis as assessed by flow cytometry analysis of annexin-V and propidium iodide staining (Figure 31B-C). AKT phosphorylation and activation was observed within 15 minutes. Subsequently the AKT targets

GSK3 and FKHR were phosphorylated by cross-linking of ICAM-2 in a time dependent manner (Figure 31D). Incubation with LY294002 abrogated the anti-apoptotic effects initiated by cross-linking ICAM-2 in Jurkat T cells and BaF3 cells (data not shown).

5 It was of interest to understand if ICAM-2 antibody clustering can initiate a cell survival signal able to overcome the apoptotic machinery after induction of an apoptotic program. To test this, Jurkat T cells were treated with staurosporine for one hour to initiate an apoptotic program and then challenged to survive by ICAM-2 antibody clustering. After 1 hour of staurosporine treatment, hallmarks of apoptosis initiation were evident as detected by cleavage of PARP,
10 cleavage of caspase 9, and cleavage of effector caspases 3 and 7 (Figure 31E). Interestingly, detection of these cleavage apoptotic indexes decreased as ICAM-2 clustering stimulated AKT activity as a function of time (Figure 31E-F). This was not observed in cells that were treated with staurosporine but ICAM-2 crosslinked (data not shown). ICAM-2 clustering initiated these hallmark cell survival signals in the presence of the staurosporine induced death signal as
15 detected by increases in AKT phosphorylation and AKT activity. In addition, BAD phosphorylation also followed ICAM-2 clustering in these challenged T cells, demonstrating that activation of the PI3K/AKT pathway is sufficient to override apoptotic paradigms.

To attribute a physiological relevance to ICAM-2 induced AKT activity in vivo, human PBMC
20 were crosslinked for ICAM-2 prior to treatment with TNF α and anti-Fas monoclonal antibody. Treated PBMC were gated by immunophenotype for CD4 and CD19 and subsequently assayed for AKT activity and apoptosis by muHi-parameter FACS. Intracellular AKT kinase activity measurements were achieved by development of a phospho-specific fluorescent probe for AKT-phospho ser473 phosphorylation site on AKT detectable by FACS (Perez and Nolan,
25 submitted). Both ICAM-2+/CD4+ T cells and ICAM-2+/CD19+ B cells exhibited AKT activity when ICAM-2 was clustered (Figure 32A). ICAM-2 induced AKT activity protected B cells from apoptosis to Fas and TNF- α induced apoptosis (Figure 32B). There was little apparent anti-apoptotic effect on CD4+ T cells (Figure 32B) perhaps due to the fact that these T cells were not strongly induced by Fas or TNF- α to undergo apoptosis with our conditions.
30 Protection from apoptosis in primary B cells correlated perfectly with AKT phosphorylation at serine 473, confirming prior roles for this target serine in the protective effect of AKT against Fas and TNF- α induced apoptosis. Thus, ICAM-2 oligomerization leads to the apparent membrane recruitment of PI3-kinase and subsequent activation of AKT kinase, resulting in a potent inhibition of apoptosis in Jurkat T cells, BaF3 pro-B cells, and human primary T and B
35 cells.

Discussion

A model for ICAM-2 mediating a cell survival signal

Our results demonstrate that ectopic expression, or crosslinking of endogenous, ICAM-2 in a wide variety of cell types as well as in primary CD19⁺ B cells can initiate a survival signal to cells. ICAM-2 induced AKT activation is a survival signal that can render a cell resistant to chemical (staurosporine and etoposide), and physiological (TNF α and Fas ligand) apoptotic inducers.

ICAM-2 co-immunoprecipitated with both α -actinin and ezrin, demonstrating interactions with critical membrane-cytoskeletal linker proteins. ICAM-2's anti-apoptotic effect was dependent on the 21 amino acid cytoplasmic domain known to bind α -actinin. Notably, α -actinin was shown to interact directly with PI3 kinase through the p85 SH3-domain (Shibasaki et al., 1994). Furthermore, the ICAM-2 cytoplasmic domain binds to members of the ezrin/radixin/moesin (ERM) family of membrane-cytoskeleton linker proteins (Helander et al., 1996). ERM-proteins regulate cell morphology, adhesion and growth by promoting membrane cytoplasmic linkages (Tsukita and Yonemura, 1997) and include the tumor suppressor, merlin/schwannomin, implicated in neurofibromatosis type II. In addition, ezrin and other ERM proteins have been implicated in different diseases for their modulation of adhesion molecule distribution (Ichikawa et al., 1998; Stokowski and Cox, 2000; Turunen et al., 1998; Viores et al., 1995). Adhesion dependent survival was recently reported to be mediated by ezrin in epithelial cells through a direct interaction with PI3-kinase and pursuant AKT activation (Gautreau et al., 1999). Interestingly, ICAM-2 clustering was induced by errin overexpression (Helander et al., 1996) suggesting that inappropriate surface protein clustering and subsequent signaling can be strongly influenced by ERM proteins. Hence, we propose that the ICAM-2 cytoplasmic domain interacts with membrane-cytoskeletal linker proteins such as α -actinin and/or ezrin, leading to the recruitment and activation of PI3 kinase to the membrane and the subsequent activation of AKT kinase-mediated survival activity.

The PI3K/AKT signaling system is a general mediator of extracellular stimuli, including growth factors, cytokines, as well as adhesion to extracellular matrices (Downward, 1998). The activation of AKT is dependent on phosphorylation at two sites, threonine-308 and serine-473 (Downward, 1998). The amino terminus of AKT contains a pleckstrin homology domain that is thought to bind directly to phospholipid products of PI3K. This binding recruits AKT to the membrane that has been proposed to induce a conformational change allowing phosphorylation at threonine-308 and serine-473 by PDKI and PDKII respectively (Alessi et al., 1996). Dually phosphorylated AKT is then active and can phosphorylate a number of

downstream effectors. Another mode by which AKT is thought to inhibit cell death is by preventing release of cytochrome C from mitochondria (Kennedy et al., 1999). Activation of AKT kinase induced by ICAM-2 clustering resulted in activation of several downstream effectors as detected by phosphorylation of BAD, GSK3, FKHR and AFX. Phosphorylation of BAD at ser-136 results in binding to 14-3-3 instead of BCL-X_L (Zha et al., 1996), shifting the equilibrium of Bcl-2 family members to favor cell survival. GSK3 activity has been shown to be necessary for apoptosis resulting from PI3K inhibition (Pap and Cooper, 1998). Phosphorylation of GSK3 inhibits its activity and therefore would present an anti-apoptotic signal. Phosphorylation of FKHR inhibits its ability to translocate to the nucleus and initiate FKHR-dependent transcription. Within the nucleus, dephosphorylated FKHR can induce target genes such as Fas ligand in certain cells and trigger apoptosis (Brunet et al., 1999). Phosphorylation of AFX inhibits transcriptional activation of p27, a cell cycle inhibitor (Graff et al., 2000). In addition, AKT has the ability to phosphorylate and inactivate caspase 9 (human caspase 9), phosphorylate eNOS (endothelial cells) and promote angiogenesis of vascular endothelium, and potentially other substrates (Cardone et al., 1998; Kureishi et al., 2000). AKT hyperactivity has been observed in a number of disorders (Haas-Kogan et al., 1998) (Kulik et al., 1997; Shan et al., 2000; Wick et al., 1999). All these data support the notion that ICAM-2 activation would be expected to lead to an extremely powerful anti-apoptotic signal in cells. A model of ICAM2's interaction with the PI3K/AKT pathway is depicted in Figure 32C.

The anti-apoptotic effects mediated by ICAM-2 overexpression and by ICAM-2 clustering suggest that inappropriate ICAM-2 expression contributes an anti-apoptotic phenotype. As an adhesion molecule, ICAM-2 has been attributed a role in lymphocyte recirculation and through interactions with its receptor LFA-1, a role in enhancing and stabilizing T-cell:B-cell contact (Damle et al., 1992; Douglas et al., 2000). Here, we suggest that in addition to possessing adhesion molecule properties, ICAM-2 is capable of transmitting an intracellular signal when it is clustered on the cell surface. The significance of ICAM-2 clustering, by virtue of its overexpression in a cancer or by receptor-mediated engagement, could lead to survival in cells otherwise destined to die by apoptosis. This could lead to tumor drug resistance and cytoproliferative disorders such as B-cell chronic lymphatic leukemia (B-CLL). For instance, it was reported that ICAM-2 is highly expressed on CD5⁺ B cells from B-CLL patients (Molica et al., 1996) and on large B-cell lymphomas derived from patients (Alizadeh et al., 2000) indicating that ICAM-2 or other adhesion molecules may have a role in certain classes of leukemias. In normal cells, ICAM-2 mediated PI3K activation may likely regulate the

endothelial transmigration of eosinophils (Gerwin et al., 1999), and potentially dendritic cells (via DC-SIGN) (Geijtenbeek et al., 2000).

Our results illustrate that ICAM-2 overexpression and clustering can provide an, anti-apoptotic signal in B cells rendering them resistant to physiological death 2 programs induced by TNF α and Fas ligand. Notably ICAM-2 expression is upregulated in many tissue culture cell lines in comparison to their normal counterparts, suggesting that cells in an ex vivo context need to initiate cell survival programs (de Fougères et al., 1991). The work presented herein attributes a previously unrecognized function for ICAM-2. The finding that ICAM-2 can exert cell survival signaling, ectopically and endogenously, as well as elicit an anti-apoptotic phenotype in a variety of cell lines and human primary cells suggests that deregulation or induced clustering is likely to contribute significantly to the anti-apoptotic phenotype that leads to tumor progression. Thus, pathways involved in the regulation of ICAM-2 expression and distribution or other adhesion molecules could be important in the development of anti-tumor therapies.

EXPERIMENTAL PROCEDURES

Retroviral cDNA screening

Production and infection of retroviruses was carried out as described (Kitamura et al., 1995; Onishi et al., 1996; Rayner and Gonda, 1994). A cDNA library was prepared from Jurkat T cells in the pBabe-S vector (Hitoshi et al., 1998). The library (2×10^6 primary transformants) was used to transfect $\sim 10^7$ PHOENIX-E packaging cells. The pBMN-LacZ control vector was spiked into the library at 1:10 to monitor the transfection/infection efficiency. Viral supernatants were harvested and used to infect $\sim 10^7$ exponentially growing NIH3T3 cells; 48 hours post infection the cells were split and analyzed for β -galactosidase activity. The library transduced NIH3T3 cells were plated at $\sim 25\%$ confluency; 24 hours later the medium was replaced with DMEM containing $1 \mu\text{M}$ staurosporine (STP; Sigma, St. Louis MO). After 24 hours treatment the STP-containing medium was replaced with complete DMEM/10% donor bovine serum. Surviving cells were allowed to reach $\sim 80\%$ confluency (7 days). Survivor cells were retreated as described two additional rounds.

Phenotypic transfer assay

An aliquot of the surviving library-infected cells were super-infected with Moloney Murine Leukemia Virus (gift of T. Kinsella, Stanford). Super-infected cells were passaged for two-weeks to allow efficient spread of the helper virus. Naive NIH3T3 cells were infected with viral supernatants and allowed to express for 48 hours. Re-infected cells were then treated with

staurosporine as described above. Surviving cells were cultured for 7 days and analyzed for transfer by RT PCR.

RT-PCR cloning of provirap-cDNA inserts and construct generation

- 5 RNA was prepared by the acid-guanidium-phenol-chloroform method as described (Chomczynski and Sacchi, 1987). cDNA was prepared from 4 μ g heat-denatured total RNA in a 50 μ l reaction volume (Promega, Madison, WI) and incubated at 42°C and 55°C for 45 minutes. The cDNA reaction products (5 μ l) were amplified (primers: 5'-GATCCTCCCITTATCCAG; 5'-GAATGAAAGACCCACCTGT) in a 50 μ l PCR reaction mix
10 (Perkin-Elmer, Foster City, CA) at 95°C/30sec; 55°C/60sec; 72°C/2min for 25 cycles. Full length ICAM2 cDNA was cloned into retroviral vector PBM-Z-IN backbone (Kinoshita et al., 1998) at BamHI/Sal1 site. ICAM2-AC, ICAM2-AN and ICAM2-C-terminal scrambled were generated by PCR and cloned into the BamHI/Sal1 site.

15 Cell culture and preparation of primary cells

- NIH3T3 murine fibroblast were maintained in DMEM, 10% DCS, 1% PSQ (Duelbecco Modified Eagle Media, 10%Donar calf serum, 1 % penicillin streptomycin (1000 units/ml and 2mM L-glutamine PSQ). Jurkat T-cells and CH27 transformed B-cells were maintained in RPMI-1640, 10% FCS, 1% PSQ. BaF3 pro-B-cells were maintained in RPMk1640, 10% FCS,
20 1% PSQ, 400 U/ml IL-3 (Peprotech). 70Z/3 pre-B-cells were maintained in RPMI-1640, 10% FCS, 1 % PSQ, 50 μ m β -mercaptoethanol. HL60 myeloid leukemia cells were maintained in Opti-Mem I, 10% FCS, 1% PSQ. 293T human fibroblast cells were maintained in DMEM, 10% FCS, 1%PSQ. Cells were maintained at 5% CO₂ /37°C humidified incubator. Transduced cells were maintained in 500 μ g/ml G418 (Sigma) and proper expression of construct was verified by
25 western blots and FACS analysis of full length ICAM-2, ICAM-2 Δ C, ICAM- Δ N as indicated. Staurosporine (Sigma) treatment was for 24 hrs at 1 μ M (unless otherwise indicated). LY294002 (New England Biolabs) treatment was at 10 μ m 30 minutes before any subsequent treatment. Wortmannin (Sigma) treatment was for 24 hrs at 100 nM. Chemicals were dissolved in DMSO (final being 1:1000 for solvent dilution) and vector controls were incubated with 1 %
30 DMSO as negative control. Vector controls consisted of pBMN-Z-IN (empty vector) transduction or pBMN-Z-IN-GFP (http://www.stanford.edu/group/nolan/plasmid_maps/pmmaps.html) as indicated. The infection frequency of pBMN-Z-IN-GFP was 48% \pm 10 for three independent viral transductions. Continual neomycin selection yielded homogenous ICAM-2, ICAM2- Δ C, ICAM2- Δ N, or control
35 vector populations as routinely monitored by flow cytometry for protein expression. For

preparation of primary cells, mononuclear cells were isolated from human peripheral blood by Ficoll-plaque density centrifugation and depletion of adherent cells on adherent plastic culture dishes. Isolated cells were maintained in complete media and analyzed by FACS for CD4, CD3, CD19, and ICAM-2 expression.

5

Cross-linking

Antibody cross-linking experiments were performed using a mouse monoclonal anti-ICAM-2 that recognizes the N-terminal region (extracellular domain) at 10 µg/ml. Spin dialysis (Biorad) was used for buffer exchange of azide containing antibodies (0.01%) to phosphate buffered saline buffer pH 7.4. Anti-mouse IgG (Sigma) was used as a control antibody in cross-linking experiments. 10⁶ Jurkat or BaF3 cells were serum starved for 3 hours or 4 hours respectively. Cells were incubated with either ICAM-2 (10 µg/ml) or mouse IgG (10 µg/ml) at 37°C for the indicated time. Time points began post serum starvation to attenuate most of the signaling pathways or as indicated. Serum starvation of 3 or 4 hrs lacked biochemical initiations of apoptosis as tested by caspase 3 activity, cell cycle analysis, annexin V/PI staining (data not shown). Cells extracts were taken and subjected to either immunoprecipitation or immunoblotting. ICAM-2 clustering of primary human cells was performed using anti-human ICAM-2 (IC2/2 Research Diagnostics) conjugated to FITC. The pursuant staining for ICAM-2 resulted in ICAM-2 clustering and signal transduction.

20

Apoptosis assays

Apoptosis was determined either by counting pyknotic nuclei, annexin-V binding by flow cytometry, or TUNEL BrdU staining by flow cytometry as indicated. Apoptosis was induced either by staurosporine (1 µM), treatment with anti-mouse Fas (5 µg/ml Santa Cruz Biotechnology (SCB), anti-human Fas (5 µg/ml Jo2 Biomed) or anti-human Fas (10 ng/ml CH11 Kamiya Biomedical) as indicated in appropriate experiment, IL-3 withdrawal (24 hrs) or as indicated. For pyknotic nuclei count, NIH3T3 cells were grown on glass coverslips and assayed for apoptosis. Suspension cells were analyzed by nuclear staining with 25 µM acridine orange/25 µM ethidium bromide or 5 µg/ml Hoechst 33342 (Molecular Probes, Eugene, OR). Apoptosis was identified by the presence of characteristic pyknotic nuclei as described (Jacobson and Raff, 1995). At least 5 individual fields were photographed and counted using a Zeiss axioscope fluorescence microscope. For the survival assay 10⁶ NIH3T3 cells were plated in 10 cm dishes and incubated overnight at 37°C. The cells were treated with 1 µM staurosporine for 24 hours. The cells were washed and cultured for 24 hours in complete medium. The cells were trypsinized and viable cells counted following trypan blue staining. Untreated cells were counted at the time of treatment. % survival represents (number of viable cells post-treatment)/(number of viable cells pretreatment). FACS based annexin-V-FITC

(SCB)/PI (Clontech) staining was done as described (PharMingen). TUNEL assay was performed as described by manufacturer of Apoptosis BrdU kit (Promega). Flow cytometry data acquisition was performed 24 post staurosporine treatment (1 μ M) for both BrdU (10,000 events collected) and pyknotic nuclei count and 12 hr post-staurosporine treatment for annexin V binding (100,000 events collected). Annexin-V positive cells will have compromised membranes after 24 hr treatment. Flow cytometry analysis was performed on a BD FACSCalibur machine with CELLQuest and analyzed using FlowJo software (Tree Star).

AKT kinase assays

AKT activity was detected by immunoprecipitation of AKT1 from cells and used in a kinase assay with GSK3 fusion protein (NEB). 2×10^6 NIH3T3, 2×10^6 Jurkat, or 5×10^5 BaF3 cells were incubated with immobilized AKT 1G1 monoclonal antibody (mAb) (1:200, NEB) at 4°C with gentle rocking motion for 2 hrs. Immunocomplexes were washed 4X with cell lysis buffer and resuspended in 40 μ l kinase buffer (25 mM Tris pH 7.5, 5 mM β -glycerolphosphate, 2mM DTT, 0.1 mM Na_3VO_4 , 10 mM MgCl_2) supplemented with 200 μ M ATP and 1 μ g of GSK-3 fusion protein (NEB) for 30 min at 30°C. Kinase reaction was terminated with SDS sample buffer boiled for 5 min, and phosphorylation state of GS3K was detected by immunoblotting and visualized using ECL detection (Amersham). Immunoblots are representative of 3 independent virally transduced cell populations each repeated 3 times (n=9). AKT activity was verified by AKT phosphorylation by immunoblotting with phospho-specific antibodies to ser473 and thr308 (NEB).

Immunoprecipitations and Immunoblotting

Cell extracts were prepared by washing 2×10^6 cells in ice cold PBS and harvesting in lysis buffer (20 mM Tris pH 7.5, 150 mM NaCl 1 mM EDTA 1 mM EGTA, 1% Triton X-100, 2.5 mM Na_2PO_4 , 1 mM β -glycerolphosphate, 1 mM Na_3VO_4 , 1 μ g/ml Leupeptin, 1 mM PMSF, protease inhibitor cocktail tablet (Boehringer Mannheim)). Extracts were centrifuged 14,000 RPM for 5 min at 4°C and cell lysates (20 μ g as determine by BCA protein assay (Pierce)) were fractionated on 12% or 15% SDS-polyacrylamide gel electrophoresis and transferred to PVDF membranes using standard procedures. Immunoprecipitations were pre-cleared with protein A/G plus-agarose beads (SBC). IP was incubated for 1 hr or overnight with primary antibody, 1 hr with protein A/G plus-agarose beads and washed 4X with lysis buffer. Blots were incubated with the following antibodies: ICAM2 N-terminal (1:500 SBC), anti-ICAM2 C-terminal (1:500 SBC), anti-ICAM2 IC2/2 mAb (1:500 BD Biosciences), anti- α -actinin (1:500 SBC), anti-ezrin mAb (1:500 Transduction Laboratories), anti-Bcl2 (1:500 SBC), anti-p27 (1:1000 SBC),

anti-p21 (1:500 SBC), anti-cFLIP (1:500 SBC), anti-phospho-AKT-ser473 (1:1000 NEB), antiphospho-AKT-thr308 (1:1000 NEB), anti-AKT (1:1000 NEB), anti-phosphoBADser155 (1:1000 NEB), anti-FKHR (1:1000 wb, 1:200 IP NEB), anti-phospho FKHR (1:1000 NEB), anti-phospho-Gsk3 α /021 (1:500, NEB), anti-cleaved PARP (1:1000 NEB), anti-cleaved caspase 9 (1:1000 NEB), anti-cleaved caspase 3 (1:1000 NEB), anti-cleaved caspase 7 (1:1000 NEB), anti-pAFX (1:1000 NEB), anti-caspase 9 (1:1000 NEB), anti-caspase 7 (1:1000 NEB), anti-PARP (1:1000 NEB). BAD phosphorylation was determined by immunoprecipitating total BAD (1:200 NEB) and immunoblotting with anti-BADser112 (1:1000 NEB) and antiBADser136 (1:1000 NEB) as recommended by manufacturer of BAD phosphorylation detection kit (NEB). Secondary antibodies used: anti-rabbit HRP (1:5000 NEB), anti-mouse HRP or anti-goat HRP 1:5000, SBC) and visualized using ECL detection (Amersham) or with a Kodak Digital Science 440C station. Blots are representative of at least 3 independent experiments.

15 Flow Cytometry and FACS analysis

Intracellular and extracellular staining was performed as described (www.metazoa.com/UPL3287). Antibodies used: anti-ICAM2 N-terminal (SBC), anti-ICAM2 N-terminal (SBC), anti-ICAM2-FITC mAb (BD Biosciences/Fisher), annexin-V-FITC (BD Biosciences), annexin-V-biotin (SBC), PI (Cloneteck). Secondaries used: anti-rabbit PE (BD Biosciences), anti-mouse-FITC (BD Biosciences), anti-rabbit FITC (PharMingen), anti-goat FITC (BD Biosciences), anti-goat texas red (BD Biosciences), Streptavidin-PE (BD Biosciences), streptavidin-FITC (BD Biosciences), anti-human CD4-APC (PharMingen), anti-human CD19-PerCP (PharMingen), anti-human ICAM-2-FITC (Research Diagnostics IC2/2) Intracellular probe for AKT activity was made by conjugating monoclonal anti-AKTser473 antibody (Cell Signaling Technology) to Alexa-568 dye (Molecular Probes), using Alexa-568 protein conjugation kit (Molecular Probes). Phospho specificity was tested by western blotting and FACS analysis to a variety of PI3K activators and inhibitors (data not shown). Intracellular staining by phospho-AKTser473-Alexa568 reflected AKT kinase activity when NIH3T3 cells were stimulated with platelet derived growth factor (Sigma) and inhibited by LY294002 prior to stimulation. Quantitative FACS analysis was performed as described (Davis et al., 1998; Iyer et al., 1998; Lenkei et al., 1998). In brief, R-phycoerythrin (PE) (Molecular Probes) was conjugated to a anti-ICAM2 N-terminal monoclonal antibody (Research Diagnostics IC2/2) as suggested by manufacturer of protein cross-linking kit (Molecular Probes), tested for proper stoichiometry (data not shown) and quantitated using Quantibrite-PE beads (BD systems). A quantitative calibration curve was generated using Quantibrite-PE beads that contain a known amount of PE molecule/bead.

A linear regression analysis is performed using the following equation: $\log_{10}(\text{PE fluorescence}) = \text{slope} \cdot \log_{10}(\text{PE molecules/bead}) + \text{intercept}$. PE fluorescence is determined by taking the geometric mean of the PE channel. Quantitation is valid only for antibodies directly conjugated to PK. Using saturating amounts of antibody and thorough washing ensures all surface antigens are bound. Preparation of samples on ice reduces antibody internalization. Surface antigens of cells for an unknown cell population are determined by computing the PE geometric mean using the calibration curve. Quantitative FACS analysis provides approximate estimates for surface antigen expression. Numbers plotted represent relative surface molecules and not absolute numbers since antibody valency was not investigated for the monoclonal antibodies used. Flow cytometry analysis was performed on a BD FACSCalibur machine and analyzed using FlowJo software (Tree Star). CellQuest was used for quantitative flow cytometry and linear regression analysis of Quantibrite-PE beads. Flow cytometry data are representative of 3 independent experiments of 10^6 cells/sample analyzed. 100,000 events were collected or otherwise noted and calibrated using Calibrite beads (BD systems). Data plotted in bar graph format is expressed as the mean (bar) \pm SD of triplicate experiments.

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Example 6**ICAM-2 (CD102) signaling events necessary for human CD56⁺CD8⁺ T cell cytotoxicity**

In this Example, using the methods and compositions of the present invention, the present
5 inventors (also referred to herein as "we") show that Leukocyte Function Antigen-1 (LFA-1) is
essential in the formation of immune cell synapses and has a role in the pathophysiology of
various autoimmune diseases. In this Example, using the methods and compositions of the
present invention, the present inventors demonstrate that ICAM-2 induced an LFA-1 signal
transduction pathway that is linked to receptor clustering and activation by both the microtubule
10 and actin cytoskeleton. ICAM-2 exhibited a 21.7 pM/cell binding affinity as determined by single
cell analysis. ICAM-2/LFA-1 engagement induced activation of PKC and a reorganization of
both the actin and microtubule cytoskeleton. These events resulted in a Syk dependent
activation of the p44/42 MAPK pathway upon cytotoxic T cell effector-target cell binding via
active LFA-1. ICAM-2 mediated human CD56⁺CD8⁺ perforin release and resultant cytotoxicity
15 to target leukemia cells. In comparison to the other ICAMs, ICAM-3 was found to be most
similar to ICAM-2's effect and dissimilar to ICAM-1. In IL-2 pre-activated human PBMC, ICAM-2
> ICAM-3 >> ICAM-1 in mediating perforin release of a CD56⁺CD8^{med} population. All ICAMs
contributed to perforin and granzyme-A loss in CD56⁺CD8^{high} populations. These results identify
a specific functional consequence for ICAM-2/LFA-1 in subset-specific cytotoxic T cell
20 immunity.

Introduction

Leukocyte Function Antigen -1 (LFA-1) is an α,β heterodimer integrin involved in leukocyte
adhesion (van Kooyk and Figdor, 2000). At present, it is well understood that LFA-1
25 participates in lymphocyte adhesion, with prominent roles in the formation of the immunological
synapse (Dustin and Shaw, 1999), and lymphocyte extravasation and recirculation (Volkov et
al., 2001). LFA-1 adhesion is governed by the intercellular adhesion molecule (ICAMs) -1, -2,
and -3 ligands (van Kooyk and Figdor, 2000). Patients afflicted with Leukocyte Adhesion
Deficiency disorder (LAD), a syndrome in which the LFA-1 integrin is mutated or missing, suffer
30 sever recurrent bacterial infections and impaired overall immunity (Bunting et al., 2002).
Among these clinical manifestations, the LFA-1 knockout mouse has suggested that LFA-1
may have a potential role in mediating tumor regression in adoptive immunotherapy (Mukai et
al., 1999; Nishimura et al., 1999). Although these studies genetically link a lymphocyte
adhesion molecule with impaired immune function, the molecular details that mediate these
35 immunopathologies are less well understood.

Investigations of LFA-1 have primarily focused on the integrin's adhesive role. It is unclear as to how the physical processes of LFA-1 integrin activation and receptor clustering are interconnected and translated into cellular signals upon ligand binding. It is less understood how the absence of these events leads to the devastating effects of LAD and the impaired immune responses in LFA-1 knockout mice. We therefore sought to decipher the molecular details of a model interaction of ICAM-LFA-1 to understand LFA-1 signaling mechanisms initiated upon cell-to-cell contact. Utilizing multiparameter single cell analysis to monitor LFA-1 receptor dynamics upon treatment with a soluble ICAM-2, we found that both the actin and the microtubule cytoskeleton couple ICAM-2 adhesion to LFA-1 activation and clustering. The microtubule cytoskeleton constrained the LFA-1 conformational change (activation), an event that preceded LFA-1 clustering as measured by multiparameter flow cytometry. The induced LFA-1 activation led to the activation of the p44/42 Mitogen Activated Protein Kinase pathway (MAPK; RAF/MEK/ERK), an event that was dependent on both Pyk2 and Syk kinase activities.

The present inventors investigated these molecular details of the ICAM-2 mediated LFA-1 activation in the adhesion between cytotoxic T cells and a target leukemia cell, an event that requires cell-to-cell contact. ICAM-2 stimulation of human CD56⁺CD8⁺ T cells could induce perforin/granzyme-A mediated cytotoxicity of leukemia cells. This directed killing was shared by ICAM-3 and to a lesser extent by ICAM-1, two other LFA-1 ligands. These results distinguish a signaling mechanism for ICAM-2/LFA-1 directed cytotoxic T lymphocyte immunity and suggest possible mechanisms by which tumor secretion of ICAM-2 and possibly ICAM-3 might allow for evasion of a directed cytotoxic T cell immune response.

Results

Recombinant ICAM-2 promotes LFA-1 mediated adhesion

The present inventors chose a model Jurkat T cell line as a system to initially dissect the LFA-1 signaling mechanism and then verified the findings in human T cells. A biochemically purified ICAM-2 protein was produced to study ICAM-2/LFA-1 interactions in the absence of other ligands (*Reviewers please see Supplementary Figure 33A-B*). We purified human ICAM-2 from retrovirally transduced NIH3T3 cells using immunoaffinity chromatography and subsequent gel filtration. We compared it to an ICAM-2-FC fusion protein produced in NSO murine myeloma cells. These murine based mammalian expression systems were chosen on the basis that they yielded a bioactive form of ICAM-2. Biochemical analysis of ICAM-2FC protein was consistent with the expected molecular weight of the fusion protein (76 kD, Figure 33A) and purified human ICAM-2 displayed a molecular weight of 72-74 kD (Figure 33A). This size was similar to the 75 kD ICAM-2 purified from Jurkat T cells (data not shown).

The present inventors generated a FITC conjugated ICAM-2 (ICAM-2-FITC) to study LFA-1 receptor dynamics by flow cytometry and laser scanning confocal microscopy (LSCM) (*Reviewers please see Supplementary Figure 33C*). We tested for ligand binding of the LFA-1 receptor by monitoring the binding kinetics of ICAM2-FITC on single cells. Low binding was
5 observed in the first 150 seconds, whereupon there was a progressive increase until 750 seconds, and leveled thereafter (Figure 33B, middle panel). In contrast, an α -LFA-1 antibody displayed an initial spike in the first 50-100 seconds and equilibrated until 800 seconds (Figure 33B, bottom panel). Pre-activating LFA-1 by treatment with PMA (McDowall et al., 1998) showed an immediate binding of ICAM-2 (Figure 33C, top panel). The gradual ICAM-2 binding
10 after 150 seconds suggested an enhanced LFA-1 binding for its ICAM-2 ligand after some binding-induced event—a property not observed using the α -LFA-1 or upon PMA activated LFA-1 (Figure 33B). Binding of ICAM-2-FITC was not observed in trypsinized cells (data not shown) and was blocked by antibodies to LFA-1 (described below). Therefore, there appeared to be an increase in binding of the ICAM-2 ligand as a function of time, suggesting the presence
15 of an induced binding site on the target cells.

Analysis of the ICAM-2 binding population by flow cytometry showed a dependency on both the actin cytoskeleton and temperature. ICAM-2 adhesion was enhanced at 37°C vs. 4°C (Figure 33C). Pre-treatment with the actin depolymerizing agent cytochalasin D revealed two ICAM-2
20 binding populations at both 37°C and 4°C, contrasting with the binding phenomena observed for α -LFA-1 (Figure 33C). Saturation of ICAM-2-FITC was observed at 37°C more readily than at 4°C (data not shown, *Reviewers please see Supplementary Figure 33D*). Single cell binding affinity measurements for ICAM-2 were obtained by computing the percent ICAM-2-FITC bound per cell (Figure 33D). Curve fit analysis indicated a dissociation constant of $0.21 \pm 0.07 \mu\text{M}/10^4$
25 cells (Figure 33D). This value equates to 21.7 pM/cell, representing the first ligand binding measurements reported for ICAM-2 within the physiological context of cell surface LFA-1. Thus, quantitative single cell analysis of ICAM-2 ligand binding suggests strong binding at physiological temperatures.

30 Soluble ICAM-2 induces LFA-1 clustering and cytoskeleton polarization

The present inventors investigated if LFA engagement altered cytoskeletal structures and observed a reorganization of both the actin and microtubule cytoskeleton upon ICAM-2 stimulus (Figure 34A). The present inventors monitored the cytoskeletal architecture by flow
cytometry and observed a simultaneous change in the actin and microtubule organization upon
35 ICAM-2 binding (Figure 34B), an effect consistent with depolymerization. ICAM-2 treatment

induced a rapid clustering of LFA-1 within one minute, with multiple clustering events at five minutes (Figure 34C, left panel). Using the ICAM-2-FITC ligand to visualize the cell surface, indicated that the ICAM-2 ligand induced clustering of the LFA-1 receptor (Figure 34C, left panel). The clustering event showed some colocalization using a non-blocking $\beta 2$ integrin antibody (clone CTB104) (Figure 34C, right panel). Thus, we speculated that ICAM-2 binding to LFA-1 induced a signal that resulted in a reorganization of the LFA-1/ICAM-2 complex. We therefore decided to investigate this in relation to the observed changes in the actin/microtubule cytoskeleton.

The present inventors assessed LFA-1 receptor dynamics by multiparameter flow cytometry upon ICAM-2 binding to correlate LFA-1 activation and clustering. We utilized the doublet discriminator module on a FACSCalibur machine to distinguish between distributed and focalized fluorescence pulses (FFP) upon laser excitation of single cells. Incubation of ICAM-2 at 37°C vs. 4°C displayed a decrease in the FFP, an effect that was greatly enhanced upon cytochalasin D treatment (Figure 34D). ICAM-2-FITC surface binding was monitored by the fluorescence intensity and normalized against the time-of-flight (TOF) of the fluorescence pulse (FP). We interpreted the value of ICAM-2-FITC intensity per TOF as a quantitative assessment for LFA-1 clustering, as the TOF is proportional to the laser-excited cellular area. Computing this value as a function of time for an ICAM-2 stimulus (Figure 34E) is proportional to the increased clustering events observed by LSCM (see Figure 34C).

ICAM-2 adhesion induces a conformational change in LFA-1 that is regulated by the microtubule cytoskeleton.

Although the enhanced ICAM-2 adhesion and induced LFA-1 clustering is reflective of overall increased avidity for the ICAM-2 ligand, it does not necessarily reflect an LFA-1 activation state (high affinity state) (McDowall et al., 1998). Upon LFA-1 activation, a conformational change exposes an epitope that is recognized by the mAb24 antibody (Neeson et al., 2000). A mAb24-Alexa633 conjugate was used to assess the activation state of LFA-1 upon ICAM-2 stimulus by flow cytometry. Unstimulated cells did not display mAb24 binding, contrasting the induction observed with PMA treatment (Figure 34F). ICAM-2 stimulated cells displayed a bimodal population in active LFA-1, an effect that was attenuated by cytochalasin D (Figure 34F). Treatment with microtubule disrupting agents, nocodazole and taxol, resulted in full activation of LFA-1 upon ICAM-2 stimulus (Figure 34F). In contrast, disrupting the actin cytoskeleton via cytochalasin D diminished the ICAM-2 induced LFA-1 activation, although it enhanced LFA-1 receptor clustering and subsequent ICAM-2 binding (see Figure 34D). Therefore, the actin and microtubule cytoskeletal network differentially impact LFA-1 activity and avidity.

The present inventors monitored LFA-1 activation and clustering simultaneously as a function of ICAM-2 stimulus per time by flow cytometry. Correlating the mean fluorescence of mAb24 antibody with the LFA-1 clustering value revealed LFA-1 activation preceded LFA-1 clustering (Figure 34G) within 30 seconds there was a significant increase in binding of the mAb24 antibody but only a modest increase in clustering. However, after another 30 seconds up to 30 minutes the relative binding of mAb24 increased somewhat but there was a significant increase in the clustering value (Figure 34G). Thus, these results suggest that the ICAM-2 ligand induced activation of LFA-1 is followed by subsequent LFA-1 clustering.

The present inventors observed that treating cells with a PKC inhibitor, bisindolymaleimide I (BIM I), inhibited ICAM-2 induced LFA-1 activation as measured by using mAb24 binding (Figure 35A). ICAM-2 adhesion, as measured by the binding of ICAM-2-FITC, was not affected (Figure 35A). This suggested that the ligand induced receptor conformational change was dependent on intracellular kinases. Interestingly, ICAM-2 induced a calcium influx, a component necessary in PKC activation (data not shown). Thus, these observations suggest that the ICAM-2 ligand induced exposure of the mAb24 neoepitope triggers a PKC dependent intracellular signaling event. We decided to investigate the downstream signaling consequences of ICAM-2 binding to LFA-1.

ICAM-2 induces p44/42 MAPK activity through LFA-1

Flow cytometric based kinase profiling experiments were performed to identify a signaling pathway downstream of PKC activation upon ICAM-2 stimulus. Treatment with ICAM-2 induced both p44/42 MAPK phosphorylation and activation (Figure 35B-D). An ICAM-2 titration correlated with phosphorylation of p44/42 MAPK as determined by single cell flow cytometric analysis (Figure 35B, top panel), results congruent with kinase activity analysis (see Figure 35C). Titration of mAbs to α_L and β_2 integrins competed with ICAM-2 binding, and thus diminished the induced p44/42 MAPK phosphorylation (Figure 35B, bottom panel). This inhibition was not observed after pretreatment with mAbs to β_1 , β_3 , α_M , or α_X integrins (Figure 35C) indicating that the ICAM-2/LFA-1 interaction was mediating the p44/42 MAPK activation.

Activation of PKC, PYK2, and SYK are necessary for the ICAM-2/LFA-1 induction of p44/42 MAPK activity

The present inventors undertook flow cytometric based p44/42 MAPK kinase inhibition and activation profiling to identify necessary components for LFA-1 signaling. PKC inhibitor BIM I, cytoskeletal disrupting agents cytochalasin D, taxol, nocodazole, and sequestering of divalent

cations by EDTA diminished the ICAM-2 induced p44/42 MAPK signal (Figure 35D), suggesting that the ligand-induced events of LFA-1 are mechanically linked to signal transduction by the actin-microtubule cytoskeleton. To identify upstream kinases that were responsible for signal transmission from LFA-1 to p44/42 MAPK, a series of kinase inhibitors were applied and tested for their ability to abrogate the ICAM-2 induced p44/42 MAPK activity (Reviewers see Supplementary Table 1 and Supplementary Figure 34), whereas Herbimycin A and Emodin, inhibitors of src and p56lck had no effect. Tyrphostin A9 and piceatannol, specific inhibitors of proline-tyrosine kinase 2 (Pyk2) and Spleen-tyrosine kinase (Syk), respectively (Avdi et al., 2001; Fuortes et al., 1999) abrogated the ICAM-2 induced activation of p44/42 MAPK and its upstream activator Raf-1 (Figure 36A).

The present inventors tested whether Pyk2 and Syk interacted with the $\beta 2$ integrin. Pyk2 and Syk were phosphorylated and co-immunoprecipitated with $\beta 2$ integrin upon ICAM-2 treatment (Figure 36B), indicating Pyk2 and Syk translocated to the membrane (Reviewers also see Supplementary Figure 35). This was coincident with phosphorylation of Pyk2 and Syk upon ICAM-2 stimulus as a function of time (Figure 36B-C). Phosphorylation of PKC α/β_{II} and Pyk2 were detected at one minute, followed by Syk phosphorylation at 5 minutes (Figure 36C). We confirmed that Pyk2 and Syk activities were dependent on PKC activation (data not shown, Reviewers see Supplementary 36). Taken together with the above results, this suggested that the LFA-1 signaling mechanism imparted by ICAM-2 is at least initiated by PKC and relayed to the p44/42 MAPK pathway by Pyk2 and Syk.

LFA-1 is involved in effector-target cell adhesion and facilitates human cytotoxic T cell activation

Since LFA-1 is involved in adhesion between lymphocytes, a process that occurs at several immunological synapses, we were interested in investigating the molecular events identified for the ICAM-2/LFA-1 interaction in a physiological context. It has been suggested that a clustered topographic presentation of ICAM-2, independent of expression levels, is an effective target structure by which natural killer cells initiated cytotoxicity (Helander et al., 1996). We first applied a FACS based effector-target killing assay to quantitatively monitor target cell lysis of HL60 leukemic cells upon treatment with stimulated human PBMC at various effector : target cell ratios. Flow cytometric detection of target cell lysis has been reported to be more sensitive than the standard chromium release assays (Lecoeur et al., 2001). We labeled HL60 cells with the fluorescent dye CFSE and monitored the cell quantity by flow cytometry in standard effector-target cell based assays. Soluble ICAM-2 could initiate target cell lysis in the presence of IL-2 but not in the absence of IL-2 (Figure 37A). In IL-2 pre-activated cells, ICAM-1 and ICAM-3 did not initiate as potent a cytotoxic cell response in contrast to ICAM-2 (Figure 37B).

Since natural killer cells (NK) comprise a heterogeneous population, namely specific cytotoxic T lymphocytes (CTL, with C8⁺ subsets therein), NK cells (CD16⁺ and subsets therein), and CD4⁺ TH1 cells (Biron and Brossay, 2001), we determined if ICAM-2 was unique to a particular human NK cell subset. We utilized the multidimensional gating capability of flow cytometry to identify distinct cellular populations that were contributing to the cytolytic activity observed in human PBMC. We also monitored intracellular levels of perforin and granzyme-A by flow cytometry, two proteins that mediate target cell lysis by NK cells in these populations. We identified 6 distinct populations by CD8 and CD56 surface stains in human PBMC (Figure 38 panel I) and gated on these subsets for all subsequent intracellular functional assays (Figure 38A panels II-V). We performed effector-target cytotoxicity assays in the presence of ICAM-1, ICAM-2, and ICAM-3 soluble ligand and HL60 target cells. We did not observe significant changes in population subset frequencies post stimulation (Figure 38A, panel I.). The CD56⁺CD8^{low} population displayed no significant changes in intracellular perforin or granzyme-A upon stimulation with ICAM-1, -2, or -3 (Figure 38A, panel II). The CD56⁺CD8^{med} population displayed a slight increase (1.5-2 fold) in the frequency of the perforin negative population for ICAM-2 and ICAM-3 (21.5% ICAM-2 > 19.8% ICAM-3 > 13.7% ICAM-1) (Figure 38A panel III). The CD56⁺CD8^{high} population displayed a loss in both granzyme-A and perforin for ICAM-1, -2, -3 stimulations compared to unstimulated with a significant loss in the granzyme-A negative population for ICAM-2 (58.3%) compared to ICAM-1 (4.12%) or ICAM-3 (3.07%) (Figure 38A, panel IV). The CD56⁺CD8^{high} also displayed a loss of both granzyme-A and perforin by all ICAM stimulations (Figure 38A panel V). Since it was not possible to positively identify the subsets within the CD56⁺CD8⁺ population, they were omitted from analysis.

Quantifying the intracellular amounts of perforin and granzyme-A in the CD56CD8 subsets relative to unstimulated cells also identified similarities and differences for the ICAMs as evidenced below. ICAM-2 and ICAM-3 mediated loss of granzyme-A and perforin to a greater extent than ICAM-1 (Figures 38B-C). Additionally, in IL-2 pre-activated cells, differences were seen with the ICAM stimulations: ICAM-2 > ICAM-3 >> ICAM-1 displayed a loss of perforin, particularly in the CD56⁺CD8^{med/high} populations (Figure 38B). ICAM-2 and ICAM-3 also induced perforin loss in the CD56⁺CD8^{low}, however ICAM-2 required preactivation by IL-2 (Figure 38B). There were lower levels of granzyme-A detected for the CD8^{high} subsets (CD56⁺ or CD56⁻) for ICAM-2 > ICAM-3 > ICAM-1 > unstimulated (Figure 38C). In the presence of IL-2 pre-activation, all the ICAMs induced release of granzyme-A in the CD56⁺CD8^{high/med} populations, with a particular decrease by ICAM-2 (Figure 38C). No significant changes were seen in the CD56⁺CD8^{low} population for granzyme-A (Figure 38C). These differences were similar at

various effector-target cell ratios (50:1, 25:1, 12.5:1) (data not shown). Thus, similarities and difference exist for ICAM-1, -2, and -3 stimulation of cytolytic activity in CD56⁺CD8 subsets. All three ICAMs mediated perforin release in the CD56⁺CD8^{high} populations. ICAM-2 and ICAM-3 were most similar in mediating perforin/granzyme-A release in the CD56⁺CD8^{high} and
5 CD56⁺CD8^{med} populations.

We focused on the CD56⁺CD8⁺ cells (both the CD8^{med} and CD8^{high} subsets) and tested if inhibition of Syk, p44/42 MAPK or disruption of the cytoskeleton detrimentally affected effector-target (E:T) cell conjugation as measured by a flow cytometric conjugate formation assay
10 (Morgan et al., 2001). Disruption of cytoskeletal actin and microtubules enhanced E:T conjugate formation (Figure 39A) congruent with prior results that disruption by these agents enhanced LFA-1 activation. Inhibition of Syk by piceatannol inhibited conjugate formation whereas inhibiting p44/42 MAPK by PD98059 did not (Figure 39A). These results suggest that Syk activity is necessary for LFA-1 adhesion of effector-target cells and is consistent with a
15 report indicating that Syk/ZAP-70 are necessary for LFA-1 to LFA-1 activation on the same cell (Soede et al., 1999). p44/42 MAPK appeared to not be necessary for E:T conjugate formation. Monitoring active LFA-1 and intracellular activation of p44/42 depicted a time dependent correlation between these two markers in CD56⁺CD8⁺ cells as stimulated by ICAM-2 (Figure 39B).

20

Discussion

In this report it was observed that (1) ICAM-2 can induce LFA-1 clustering, activation, and cytoskeletal reorganization in the absence of exogenous activators such as cytokines or TCR signaling; (2) LFA-1 transmits a signal to the p44/42 MAPK pathway involving PKC, Pyk2, and
25 Syk upon ligand binding; and (3) LFA-1 receptor dynamics are mechanically coupled to signal transduction by both the actin and microtubule cytoskeleton network. The physiological outcome of these molecular events triggered perforin and granzyme A mediated CD56⁺ CD8⁺ T cell cytotoxicity that were mostly shared by ICAM-2 and ICAM-3 but not ICAM-1.

30 β 2 integrin signaling mechanisms vary depending on the system of study and are centered on adhesive roles in cell morphology and motility (Dib, 2000). β 2 integrin signaling has been shown to involve cytoskeletal reorganization via tyrosine phosphorylation of paxillin, vav, and GTPase activating proteins among others (Fuortes et al., 1994; Zheng et al., 1996). Studies focused on LFA-1 mediated leukocyte adhesion (CD11a/CD18) have shown a regulatory role
35 for PKC in LFA-1 avidity (Bleijis et al., 2001; Hedman and Lundgren, 1992) and have demonstrated that TCR signaling can activate LFA-1 (Peterson et al., 2001). It has also been shown that chemokines, in the absence of TCR signaling, can serve as activators of LFA-1

during lymphocyte/endothelial contact (Constantin et al., 2000). It has not been clear how LFA-1 integrin adhesion, clustering, and activation are coupled to intracellular signaling events, in the absence of external (chemokine) or internal (TCR or costimulatory molecule) stimulation.

- 5 A synthesized peptide of ICAM-2's first Ig domain (P1, amino acids 21-42) can induce LFA-1 mediated adhesion at high concentrations (62 μ M), which was comparable to a 48-fold lower ICAM-2 soluble protein concentration (1.3 μ M) in a bulk cellular adhesion assay (Kotovuori et al., 1999). However, P1 binding did not induce the active conformation of LFA-1 and did not induce calcium influx (Kotovuori et al., 1999), whereas full length ICAM-2 binding resulted in
- 10 active LFA-1 (see Figure 34D) and a calcium influx event (data not shown). The calculated ICAM-2 affinity of 217 ± 66 nM (per 10^4 cells) contrasts the 605 ± 55 nM k_D reported using BIAcore analysis of an engineered "active" locked I domain of LFA-1 (Shimaoka et al., 2001). The reported affinities for ICAM-2 binding here take advantage of single cell resolution within a physiological context, something not possible utilizing purified or genetically engineered LFA-1.
- 15 The differences observed for peptide vs. protein concentrations are likely attributed to impurities in the peptide synthesis and/or presence of carbohydrate moieties native to the endogenous ICAM-2, which comprise greater than 30 kD of its approximate 66 kD molecular weight and have been suggested to orient ICAM-2 binding to LFA-1 (Casasnovas et al., 1997; de Fougerolles et al., 1991).
- 20 We investigated the role of the actin and microtubule cytoskeleton in LFA-1 receptor activation and clustering as induced by the ICAM-2 ligand by multiparameter flow cytometry. Disruption of the actin cytoskeleton enhanced LFA-1 clustering and ICAM-2 binding, corroborating previous studies that suggested the actin cytoskeleton constrains LFA-1 mobility (Lub et al., 1997).
- 25 Interestingly, actin depolymerization abrogated the ICAM-2 induced LFA-1 activation. In contrast, disruption of the microtubules by both nocodazole and taxol enhanced LFA-1 activation as determined by exposure of the neo-epitope recognized by the mAb24. Recently, it has been reported that depolymerization of microtubules increases the lateral mobility of $\beta 2$ integrins in macrophage cell lines (Zhou et al., 2001); therefore its conceivable that the
- 30 microtubules regulate the conformational change upon ligand binding necessary for exposure of the LFA-1 activation epitope. These observations suggest the actin-microtubule cytoskeleton regulates both the high-avidity and high affinity state of LFA-1 upon ligand binding. We observed that LFA-1 signal transduction was abrogated in the presence of all cytoskeletal disrupting agents tested (cytochalasin D, nocodazole, and taxol) indicating that the LFA-1
- 35 receptor is linked to signal transduction machinery by the cytoskeleton. Thus, the mechanistic

uncoupling of the high avidity and high affinity states of LFA-1 suggests that intracellular events that regulate/mediate these two states exist at the LFA-1 integrin-cytoskeletal juncture and relay the LFA-1 receptor dynamics to intracellular signaling proteins upon ligand binding.

5 Several chemical inhibition screens were designed to identify the proteins involved in the LFA-1 to p44/42 MAPK signaling event. Both Pyk2 and Syk were identified to be necessary for activation of the p44/42 MAPK pathway and were dependent on PKC activity upon ICAM-2 binding. Phosphorylation of Pyk2 has been associated with homotypic adhesion mediated by an LFA-1/ICAM-1 interaction in B cells (McDonald et al., 2000). In addition, Pyk2 activation has
10 been shown to be necessary for p44/42 MAPK activity in other model systems (Barsacchi et al., 1999; Lev et al., 1995). Syk is a tyrosine kinase essential in α III β 3 signaling (Saci et al., 2000), and links Fc ϵ RI signaling to the ras/MAPK pathway (Jabril-Cuenod et al., 1996). Inhibition or ablation of Syk, either by pharmacological means (via inhibition by piceatannol), biochemical means (dominant negative Syk), or genetic means (Syk^{-/-} mice) inhibits natural
15 cytotoxicity (Brumbaugh et al., 1997; Colucci et al., 1999). Thus LFA-1 activation signaling to Syk, a kinase that has been shown to be important for NK cell function, provides a biochemical link between surface integrin activation and effector cell function.

The present inventors demonstrated that both Pyk2 and Syk are necessary in ICAM-2 induced
20 LFA-1 signaling to Raf-1, the upstream kinase in the p44/42 MAPK (RAF/MEK/ERK) cascade. Inhibition of p44/42 MAPK did not prevent the occurrence of CD56⁺CD8⁺ cell conjugation. By immunofluorescence analysis, it has been shown that treatment of the NK leukemic cell line YT with the p44/42 MAPK inhibitor PD98059 inhibits perforin redistribution to the site of effector-target cell contact (Wei et al., 1998). In addition, the p44/42 MAPK pathway has been shown to
25 be important in the regulation of cytotoxicity in natural killer cells (Jiang et al., 2000). Thus, the p44/42 MAPK pathway, here demonstrated to become active upon LFA-1/ICAM-2 binding, has been shown to be connected to at least perforin granule exocytosis. Thus, the LFA-1 signaling pathway as elicited by ICAM-2 contains signaling junctures that map to both the effector-target cell adhesion event and activation of cytolytic machinery in the human CD56⁺CD8⁺ cytotoxic T
30 cell population. These results provide direct evidence for a functional consequence of LFA-1 integrin adhesion with cytolytic signaling mechanisms.

We also observed that ICAM-2 was similar to ICAM-3 in mediating cytolytic activity as evidenced by release of perforin and granzyme-A in effector-cell conjugation, effects of which contrasted ICAM-1 (see Figure 38). We have previously observed similarities between ICAM-
35 2 and ICAM-3 intracellular signaling mechanism that also differed from that of ICAM-1 (Perez et al., 2002). However, the results do not exclude the possibility of ICAM-2 stimulating other yet to

be identified cytotoxic capable subsets, as high cytolytic activity was observed in bulk PBMC (see Figure 37).

Prior investigations into cytotoxic T cells have established that blocking the LFA-1/ICAM interactions inhibits effector-target cell adhesion and therefore concluded that it also blocks cytolytic activity in NK cells (Donskov et al., 1996; Krensky et al., 1984; Matsumoto et al., 2000). Functional studies of NK cells from LFA-1^{-/-} mice have demonstrated that LFA-1 adhesion is necessary for IL-2 activated NK killing (Matsumoto et al., 2000) and also that LFA-1^{-/-} CD8⁺ T cells are defective for T cell activation and effector function (Shier et al., 1999). Interestingly, NK cell cytotoxicity is defective in NK cells from LAD patients (Shibuya et al., 1999). It has only recently been shown that the directed killing of cytotoxic T lymphocytes involves polarization of the microtubule-organizing center (MTOC) towards LFA-1 at the CTL-target site (Kuhn and Poenie, 2002), an indication that LFA-1 may possess a functional role other than strictly adhesion.

In conclusion we find that ICAM-2, as an LFA-1 ligand, can mediate activation and clustering of the LFA-1 receptor—an event that in turn polarizes the microtubule and actin cytoskeleton and activates the p44/42 MAPK pathway. These events were found to be necessary for effector-target cell binding of CD56⁺CD8⁺ T cells, and perforin/granzyme A mediated cytolytic activity. This effect was shared by ICAM-3. The mechanisms governing LFA-1 receptor dynamics and intracellular signaling reported here suggest LFA-1 signaling functionally contributes in CD56⁺CD8⁺ cytolytic activity in addition to possessing an adhesive role upon which other molecular interactions occur. Improper localization of the MTOC has been shown to inhibit exocytosis of lytic granules in CD8⁺ tumor infiltrating T cells, thereby ablating perforin mediated cytolytic activity necessary for a CTL response in murine tumor models (Radoja et al., 2001b). Ironically, defective CD8⁺ tumor infiltrating T cells can effectively mediate cell killing in vitro (Radoja et al., 2001a), suggesting tumor mediated inhibitory mechanisms exist within the tumor microenvironment. The production of soluble ICAMs (1 and 3) has been observed in sera from cancer and autoimmunity patients, though analysis has not been extended to ICAM-2 (Bloom et al., 2002). Only one report has indicated that elevated levels of soluble ICAM-2 were present in leukemia patients and decreased upon chemotherapy (Mustjoki et al., 2001). The etiology of these observations is unknown. In the context of the work presented here, it is plausible to speculate that either dysregulation of surface ICAM-2 or secretion of soluble ICAM-2 can prematurely trigger or block CD56⁺CD8⁺ cytolytic activity at the effector-target site and permit tumor escape from T cell lysis. Other, specific roles, of ICAM-2 in its interaction with other

integrin ligands could lead to a better understanding of events that promulgate from the effector:target cell interface

Materials and Methods

5 Immunological and Chemical Reagents

mAbs to $\beta 1$, $\beta 2$, $\beta 3$, $\beta 4$, $\beta 5$, $\beta 6$, $\alpha 1$, $\alpha 4$, $\alpha 5$, α_L , LFA-1, Pyk2, Syk, Mac-1, ICAM-1, and ICAM-3 (PharMingen). CD3, CD4, CD8, CD19, CD56, CD45 direct conjugates (FITC/PE/PERCP/APC/Biotin), granzyme-A-FITC (PharMingen). Perforin-CY5 and CD8-CY5PE (gift from the Herzenberg Laboratory, Stanford University). ICAM-2 mAb and ICAM-2-FITC (IC2/2 Research Diagnostics). Anti-phospho PYK2(Y402), anti-phospho-p44/42 (pT185Py187) (Biosource). Anti-phospho PKC α/β (Thr638), anti-phospho-Syk(Tyr525/526), anti-phosphoRaf1(Ser259) (Cell Signaling Technologies). Protein and chemical reagents used: fluorescein isothiocyanate (FITC) (Pierce), Alexa fluor dye series 488, 546, 568, 633, taxol-alex546, phalloidin-alex633, and CFSE (Molecular Probes). Tyrphostin A9 and 18, SB203580, piceatannol, bisindolylmaleimide I and II, herbimycin A (Calbiochem). Emodin, genistein, DMSO, PMA, PHA, staurosporine, ionomycin, propidium iodide, cytochalisin D (Sigma). Protein A/G agarose (SCBT). Recombinant human IL-2 (Roche), recombinant human ICAM-1-FC, ICAM2-FC, ICAM3-FC (Genzyme). Secondary antibodies to mouse and rabbit IgG (Santa Cruz). Mock treatments consisted of mouse IgG (for antibodies), 1% BSA (for proteins), or 0.001% DMSO vehicle (for chemicals).

Cell Culture

NIH3T3 cells were maintained in DMEM, 10% DCS, 1% PSQ (Duelbecco Modified Eagle Media, 10% Donor calf serum, 1% penicillin-streptomycin (1000 units/ml and 2mM L-glutamine PSQ). Jurkat T-cells were maintained in RPMI-1640, 10% FCS, 1% PSQ at 1×10^5 cells/ml and serum starved 12 hours for all functional assays. Cells were maintained at 5% CO₂ /37°C humidified incubator. Human peripheral blood monocytes were obtained by Ficoll-plaque density centrifugation (Amersham Pharmacia, Uppsala, Sweden) of whole blood from healthy donors (Stanford Blood Bank) and depleted for adherent cells. Magnetically activated cell sorting was used to negatively isolate naïve CD8⁺ T cells for studies as indicated (Dynal, Oslo, Norway).

Soluble ICAM-2 generation and synthesis of ICAM-2-FITC and ICAM2-Beads

Full length ICAM2 cDNA was obtained from Jurkat cells and cloned into retroviral vector PBM-Z-IN at the BamHI/Sal1 site as described (Perez et al., 2002). Human ICAM-2 was overexpressed in NIH3T3 cells by retroviral infection and harvested by immunoaffinity chromatography. ICAM-2 was affinity purified using a two step lysing procedure and

subsequent purification on an anti-ICAM-2 solid support. Cells were lysed in buffer A (20 mM Tris pH 7.5, 150 mM NaCl 1 mM EDTA 1 mM EGTA, 0.1% NP40, 2.5 mM Na₂PO₄, 1 mM β-glycerolphosphate, 1 mM Na₃VO₄, 1 μg/ml Leupeptin, 1 mM PMSF, protease inhibitor cocktail tablet (Boehringer Mannheim) for 5 min 4°C, and subsequently permeabilized with 50% v/v with
5 buffer B (Buffer A plus 1% Triton-X-100) for 30 min 4°C. Supernatant was harvested by centrifugation (14,000 RPM, 5 min, 4°C). An Anti-ICAM-2 pAb to the C-terminal (4 mgs, Santa Cruz) was conjugated to an Affi-Gel Hz activated support (Biorad) as suggested by manufacturer. This support couples Ig molecules via the FC region, resulting in higher antigen binding capacity. Batch lysate of harvested supernatant was performed (4°C, for 2 hrs), and
10 washed 4 times in buffer C (0.1% Tween-20, PBS pH 7.4). ICAM-2 protein was eluted by 4.5 M NaCl (in Tris pH 6.8), dialyzed overnight (in PBS pH 7.4, 0.001% azide, 0.01% glycerol, 4°C), concentrated using size exclusion spin chromatography and stabilized using 0.01% glycerol. Anti-ICAM-2 solid support was re-equilibrated in buffer C, stored in 0.001% thimerosal and re-used up to 3 times. Purity was >98% as assessed by coomassie gel. Size exclusion
15 chromatography removed higher molecular weight aggregates and were not observed on purified ICAM-2 by native gel electrophoresis. 20 mgs were purified by this method and used for this study. ICAM-2-FITC synthesis was achieved by chemical conjugation to NHS-Fluorescein (Pierce) and unreactive dye was removed by gel filtration. ICAM-2-FITC probe did not integrate into trypsinized Jurkat cells or bind when blocked by LFA-1 antibody clones
20 TS1/22 or TS1/18 (Developmental Hybridoma Studies Bank) or unlabeled ICAM-2 protein as determined by flow cytometry. ICAM-2-FITC binding was not blocked by β2 integrin clone CT104 (Santa Cruz). Purified ICAM-2 was comparable to human recombinant ICAM-2FC fusion protein purified from NSO murine myeloma cells (Genzyme). ICAM-1FC and ICAM-3FC were also purified from NSO cells (Genzyme). Proteins were spun at 14,000 RPM, 5 min prior to use. 1 mg of ICAM-2 protein was conjugated to 2X10⁸ epoxy activated beads as suggested
25 by manufacturer (Dynal). 4X10⁵ beads containing a total of 2 μg ICAM-2 protein were used as indicated. Gel imaging was performed on a VersaDoc machine (Biorad) and analyzed using Quantity One quantitation software (Biorad).

30 Flow Cytometry

Intracellular and extracellular staining was performed as described (Perez and Nolan, 2002). Intracellular probes for active kinases were made by conjugating phospho-specific antibodies to the Alexa Fluor dye series as described and used in phospho-protein optimized conditions (Perez and Nolan, 2002). Kinetic analyses was performed by direct application of fixation buffer
35 in time synchronized 96-wells maintained at 37°C. Intracellular actin and microtubule staining

was performed using phalloidin-Alexa633 and taxol-Alexa546 dyes (Molecular Probes).

Adhesion and clustering assays were performed using ICAM-2-FITC as described in text. LFA-1 activation was assessed by either mAb24-Alexa633 or mAb24-Alexa546 conjugate, surface stained at 37°C. Flow cytometry data are representative of 3 independent experiments of 10⁶ cells/sample. 10-50,000 events were collected and manually calibrated on a FACSCalibur machine. Data plotted in bar graph format is expressed as geometric mean-fluorescence intensity (MFI) and normalized for isotype controls. Log ratios are defined as the MFI of stimulus to the MFI of unstimulated cells. Data was analyzed using Flowjo software (Treestar).

10 **Single Cell ICAM-2 binding measurements**

Percentage of ICAM-2-FITC binding was expressed as $100 \times ((\text{MFI}_{\text{exp}} - \text{MFI}_{\text{ctl}}) / (\text{MFI}_{\text{final}} - \text{MFI}_{\text{ctl}}))$, where MFI_{exp} equals the mean fluorescent intensity of experimental concentration, MFI_{ctl} equals mean fluorescent intensity of unstained cells, $\text{MFI}_{\text{final}}$ equals mean fluorescent intensity of final concentration that saturated binding. The samples were incubated with final concentrations as indicated in Figure for 30 min at 37°C in 50 µL staining media (def RPMI, 4%FCS), washed 1X (500 µL, PBS pH 7.4, containing 1mM EDTA), and resuspended in 100 µL (1% paraformaldehyde). Dilution factor of staining conditions and molecular weight of 72.1 kD was used in determining molar concentrations. The staining buffer contained 2.4 mM calcium and 2 mM magnesium. The data were fit to the equation $V = V_{\text{max}}[S] / (K_m + [S])$ where V is the percent bound, [S] is the ICAM-2-FITC concentration, and K_m is the Michaelis-Menten binding constant using Kaleidagraph software.

Laser Scanning Confocal microscopy

Jurkat cells were treated as indicated and adhered to poly-L-lysine (Sigma) coated sterilized coverslips (1 mg/ml, 30 min) by mild centrifugation (1000 RPM, 10 min), washed twice in phosphate buffered saline pH 7.4 (PBS) and fixed in 2.7% paraformaldehyde (in PBS). Cells were permeabilized (5 min, 0.1% Triton-X-100 in PBS), washed twice in PBS, blocked in 4% FCS, and subjected to antibody or intracellular staining as indicated. Stained coverslips were mounted and visualized using a Zeiss laser scanning confocal microscope 510.

30

Immunoprecipitations, Immunoblotting and Kinase Assays

Cell extracts were prepared by washing 2x10⁶ cells (treated as indicated) in ice cold PBS and harvesting in lysis buffer (20 mM Tris pH 7.5, 150 mM NaCl 1 mM EDTA 1 mM EGTA, 1% Triton X-100, 2.5 mM Na₂PO₄, 1 mM β-glycerolphosphate, 1 mM Na₃VO₄, 1 µg/ml Leupeptin, 1 mM PMSF, protease inhibitor cocktail tablet (Boehringer Mannheim). Extracts were centrifuged 14,000 RPM (5 min, 4°C) and 10-20 µg (BCA protein assay (Pierce)) were immunoblotted using standard procedures. Immunoprecipitations (IP) were pre-cleared with protein A/G plus-

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agarose beads, incubated with primary ab (1h), protein A/G plus-agarose beads (1h) and washed 4X with lysis buffer. Blots were incubated with the indicated antibodies and developed using ECL (Amersham). Immunoblots stripped and reprobed (as indicated) were done by incubating with stripping buffer (62.5 mM Tris, pH 6.8, 10% SDS, 1% β -mercaptoethanol) (30 min, 55°C). MAPK activity was detected by a p44/42 MAPK kinase kit as suggested by manufacturer (Cell Signaling Technologies).

Cytolytic activity, perforin release assays, and conjugate formation assays

Target cell lysis was measured by flow cytometric based detection of CFSE labeled HL60 cells. HL60 cells were labeled with 1 μ g of CFSE (30 min, 37°C). Targets were washed twice and mock treated, IL-2 activated (100 U/ml), CD3/CD28 activated (1 μ g/ml), or treated with ICAM2 beads or soluble ICAM-1, -2, or 3 (10 μ g/ml, 30 min, 37°C) before plating at 10^4 target cells/well of a 96-well round bottom plate. CTLs were added at 50:1, 25:1, and 12.5:1 E:T ratio, and incubated at 37°C for 4 hrs. Cells were then processed for multiparameter flow cytometry and intracellular perforin stain. Percent specific lysis was calculated by the following equation: %specific lysis = 100 - 100 x (experimental HL60 count/total control HL60 count). HL60 counts were detectable by the CFSE fluorescence. Percent perforin was calculated by the following equation: % perforin = 100 x [(experimental perforin MFI – isotype mAb MFI)/(total perforin MFI – isotype mAb MFI)]. MFI refers to mean fluorescent intensity of flow cytometric based intracellular detection. Cell conjugates were determined by flow cytometry as described (Morgan et al., 2001). Chemical inhibition was done at 10 μ M of indicated compound (30 min, 37°C) prior to stimulation as indicated. All experiments were performed in triplicate.

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5

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We claim:

1. A method of detecting the activation state of at least a first and a second activatable protein in single cells, said method comprising the steps of:
 - 5 a) providing a population of cells comprising said first and said second activatable proteins;
 - b) contacting said population of cells with a plurality of activation state-specific antibodies,wherein said plurality of activation state-specific antibodies comprise:
 - 10 i) at least one first activation state-specific antibody that is capable of binding to a corresponding isoform of said first activatable protein in said population of cells; and
 - ii) at least one second activation state-specific antibody that is capable of binding to a corresponding isoform of said second
 - 15 activatable protein in said population of cells; and
 - c) using flow cytometry to detect said binding of said first and second activation state-specific antibodies in single cells of said population of cells, wherein said binding of said first activation state-specific antibody is indicative of a specific activation state of said first activatable protein, and said binding of said second
 - 20 activation state-specific antibody is indicative of a specific activation state of said second activatable protein.
2. The method according to claim 1, wherein said first activatable protein is a kinase.
- 25 3. The method according to claim 1, wherein said first activatable protein is a caspase.
4. The method according to claim 1, wherein said first activatable protein is a first kinase and said second activatable protein is a second kinase.
- 30 5. The method according to Claim 4, wherein said isoform of said first kinase is a first phosphorylated kinase, and said isoform of said second kinase is a second phosphorylated kinase.
6. The method according to claim 5, wherein said first activation site-specific antibody binds to

said first phosphorylated kinase, and said second activation site-specific antibody binds said second phosphorylated kinase.

5 7. The method according to claim 1, wherein said first activatable protein is a first caspase and said second activatable protein is a second caspase.

8. The method according to claim 7, wherein said isoform of said first caspase is a cleaved product of a first pro-caspase, and said isoform of said second caspase is a cleaved product of a second pro-caspase.

10

9. The method according to claim 8, wherein said plurality of activation site-specific antibodies comprise a first activation site-specific antibody that binds to said isoform of said first caspase, and a second activation site-specific antibody that binds to said isoform of said second caspase.

15

10. A method of detecting the activation state of at least a first activatable protein in single cells, said method comprising the steps of:

a) providing a population of cells comprising at least said first activatable protein;

b) contacting said population of cells with a plurality of substrates;

20 wherein said plurality of substrates comprise at least a first substrate that is capable of being modified by a corresponding isoform of said first activatable protein in said population of cells; and

c) using flow cytometry to detect the modification of said first substrate in single cells of said population of cells, wherein said modification is indicative of

25 a specific activation state of said first activatable protein.

11. The method according to claim 10, wherein said population of cells further comprises a second activatable protein, wherein said plurality of substrates further comprise a second substrate that is capable of being modified by a corresponding isoform of said second activatable protein in said population of cells and, wherein step c) further comprises using said flow cytometry to detect the modification of said second substrate in single cells of said population of cells, wherein said modification of said second substrate is indicative of a specific activation state of said second activatable protein.

35 12. A method of detecting a protein activation state profile of single cells based on the activation state of at least a first activatable protein in said cells, said method comprising the

steps of:

- a) providing a population of cells comprising at least said first activatable protein;
- b) contacting said population of cells with a plurality of substrates;

wherein said plurality of substrates comprise at least a first substrate that is capable of being modified by a corresponding isoform of said first activatable protein in said population of cells;

- c) contacting said population of cells with a plurality of activation state-specific antibodies,

wherein said activation state-specific antibodies comprise at least one first activation state-specific antibody that is capable of binding to a corresponding isoform of said first activatable protein in said population of cells; and

- d) using flow cytometry to simultaneously detect:

- i) said binding of said first activation state-specific antibody in single cells of said population of cells,

wherein said binding of said first activation state-specific antibody is indicative of a specific activation, state of said first activatable protein; and

- ii) the modification of said first substrate said single cells,

wherein said modification is indicative of said specific activation state of said first activatable protein.

13. The method according to claim 12, wherein said population of cells further comprises a second activatable protein, wherein said plurality of substrates further comprises a second substrate that is capable of being modified by a corresponding isoform of said second activatable protein in said population of cells, and step d) further comprises using said flow cytometry to detect the modification of said second substrate in said single cells, wherein said modification is indicative of a specific activation state of said said second activatable protein.

14. The method according to claim 13, wherein said plurality of activation state-specific antibodies further comprises a second activation state-specific antibody that is capable of binding to a corresponding isoform of said second activatable protein in said population of cells, and wherein step c) further comprises using said flow cytometry to detect said binding of said second activation state-specific antibody said single cells of said population of cells, wherein said binding of said second activation state-specific antibody is indicative of said specific activation state of said second activatable protein.

15. A method of screening for a bioactive agent capable of modulating the activity of at least a

first activatable protein in cells, said method comprising:

- a) providing a population of cells, each of said cells comprising at least said a first
activatable protein, a second activatable protein, and a third activatable protein,
wherein said first activatable protein can activate said second activatable protein thereby forming
5 a specific isoform of said second activatable protein (isoform-2), and
wherein said second activatable protein can activate said third activatable protein thereby forming
a specific isoform of said third activatable protein (isoform-3);
- b) contacting said cells with a second activation state-specific antibody, a third
activation state-specific antibody, and a candidate bioactive agent,
10 wherein said second activation state-specific antibody is capable of binding to said isoform-2,
and
wherein said third activation state-specific antibody is capable of binding to said isoform-3;
- c) using fluorescent activated cell sorting (FACS) to sort single cells of said population
of cells based on the presence of said isoform-2 and said isoform-3; and
15 d) determining the ratio of said isoform-2 to said isoform-3 in said single cells in the
presence and absence of said candidate bioactive agent,
wherein a difference in the ratio of said isoform-2 to said isoform-3 in the presence and
absence of said candidate bioactive agent is indicative of the ability of said candidate bioactive
agent to modify said activity of said first activatable protein.

20

16. The method according to claim 15, wherein said first activatable protein is activated by an
activating agent.

17. The method according to claim 15, wherein said first activatable protein is a caspase.

25

18. The method according to claim 15, wherein said first activatable protein is a kinase.

19. The method according to claim 18, wherein said kinase is PI3K, wherein said second
activatable protein is PIP2, wherein said third activatable protein is PIP3, wherein said isoform-
2 is PIP 4,5 bisphosphate and, wherein said isoform-3 is PIP 3,4,5.

30

20. The method according to claim 19, wherein said PI3K is activated by a growth factor or by
activation of a cell surface receptor.

21. The method according to claim 20, wherein said first activatable protein is ICAM-2, wherein
said activity is apoptosis, second activatable protein is PIP2, wherein said third activatable
protein is PIP3, wherein said isoform-2 is PIP 4,5 bisphosphate and, wherein said isoform-3 is

35

PIP 3,4,5.

22. The method according to claim 21, wherein step a) further comprises clustering said ICAM-2 with ICAM-3.

5

23. The method according to any of claims 1-9 and 12-14, wherein said plurality of activation state-specific antibodies comprise at least one antibody selected from a group of antibodies consisting of: anti-AKT-phospho-Ser473, anti-AKT phospho-Thr308, anti-p44/42 MAPK phospho-Thr202/Tyr204, anti-TYK2 phospho-Tyr1054/1055, anti-p38 MAPK phospho-Thr180/Tyr182, anti-JNK/SAPK phospho-Thr183/Tyr185, anti-phospho-tyrosine, anti-phospho-threonine, anti-PIP2, and anti-PIP3.

10

24. The method according to any of claims 1-22, wherein step a) further comprises contacting said population of cells with an agent that induces the activation of at least said first activatable protein.

15

25. The method according to any of claims 1-9, 11, and 13-22, wherein step a) further comprises contacting said population of cells with an agent that induces the activation of at least one of said first and said second activatable proteins.

20

26. The method according to any of claims 15-22, wherein step a) further comprises contacting said population of cells with an agent that induces the activation of at least one of said first, said second, and said third activatable proteins.

25

27. The method according to any of claims 1-9, 11, and 13-22, wherein said method further comprises sorting said single cells based on said activation state of said first activatable protein, and said activation state of said second activatable protein.

30

28. The method according to any of claims 1-9, and 14-22, wherein said first activation state-specific antibody comprises a first label, wherein said second activation state-specific antibody comprises a second fluorescent label and, wherein said sorting is by fluorescent activated cell sorting (FACS).

35

29. The method according to any of claims 1-9 and 14-22, wherein said first activation state-specific antibody comprises a first FRET label, wherein said second activation state-specific

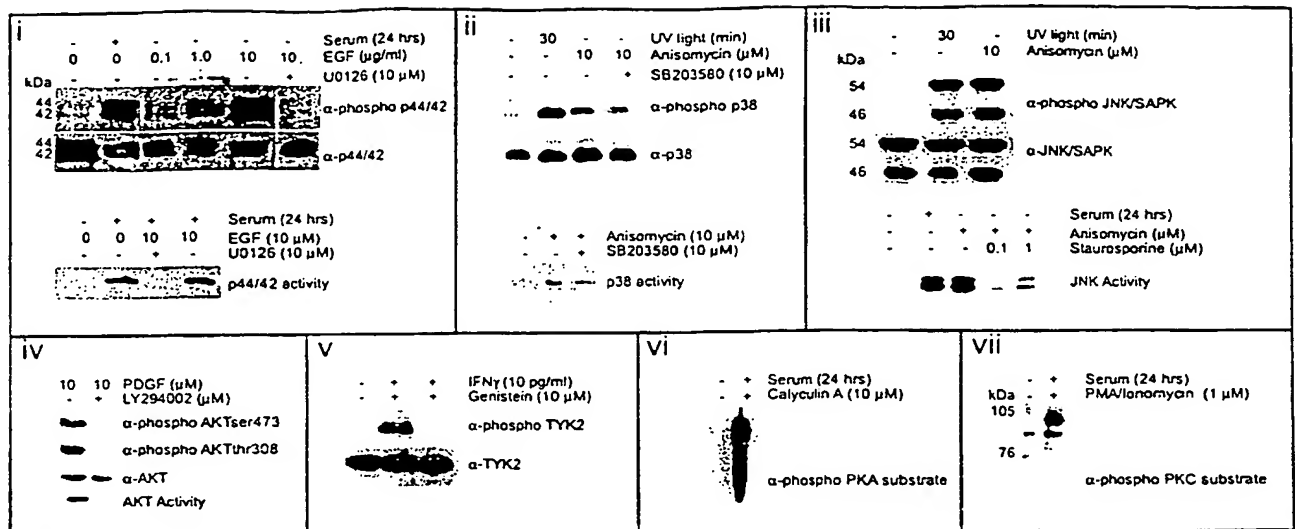
antibody comprises a second FRET label and, wherein said sorting is by fluorescent activated cell sorting (FACS).

5 30. The method according to any of claims 1-9 and 14-22, wherein said first activation state-specific antibody comprises a FRET label, wherein said second activation state-specific antibody comprises a label and, wherein said sorting is by fluorescent activated cell sorting (FACS).

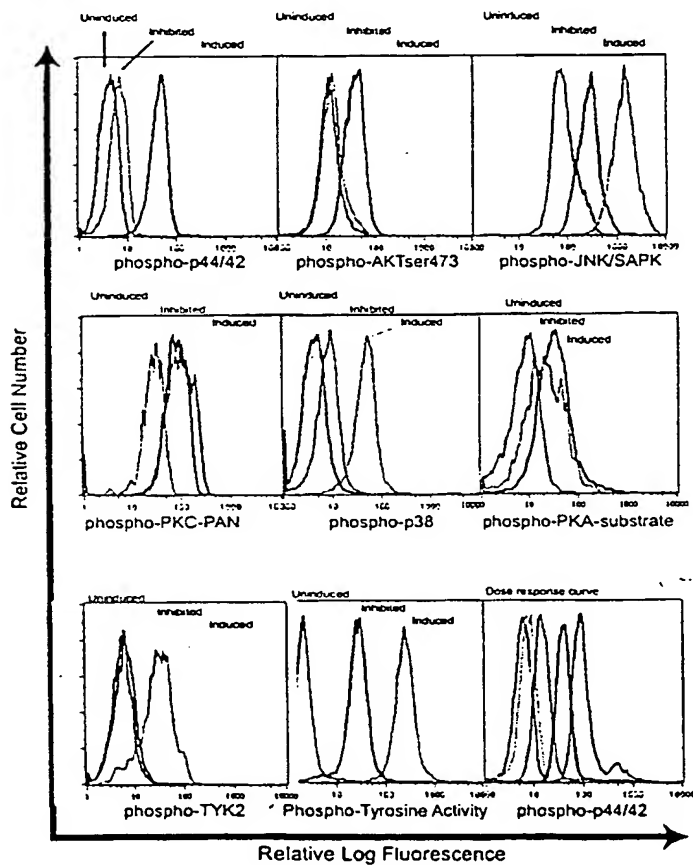
10 31. The method according to any of claim 1-22, wherein step a) further comprises fixing said cells.

32. The method according to any of claims 1-22, wherein said cells are mammalian cells.

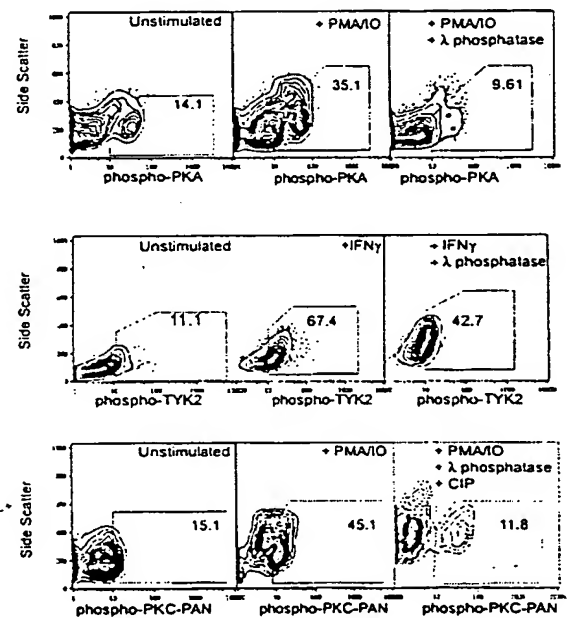
A.



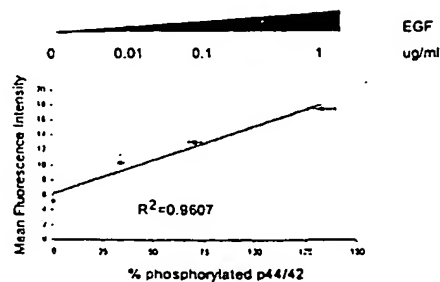
B.



C.



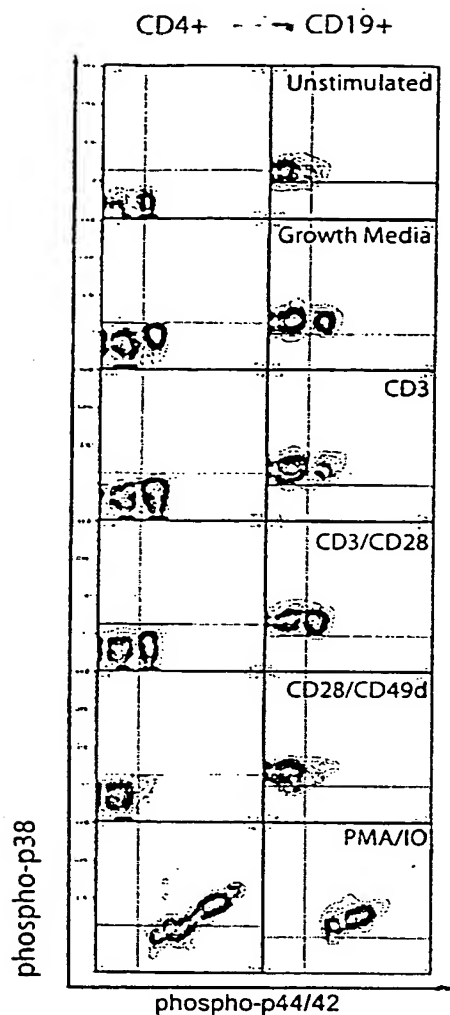
D.



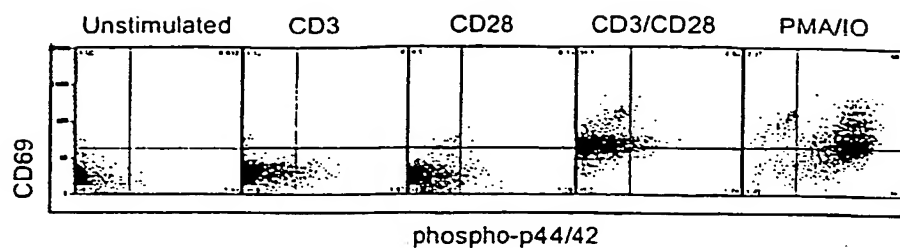
Phospho-specific antibodies correlate with kinase activity.

Figure 1

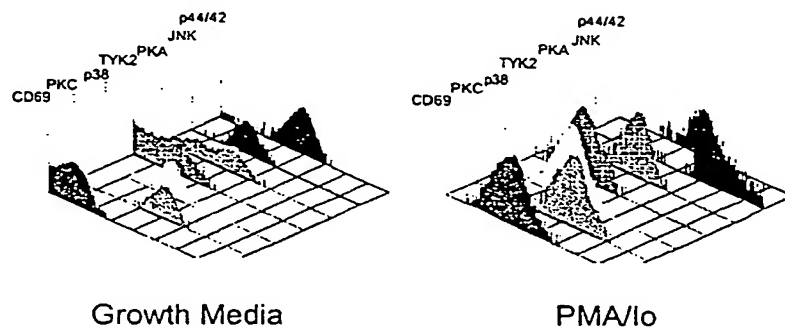
A.



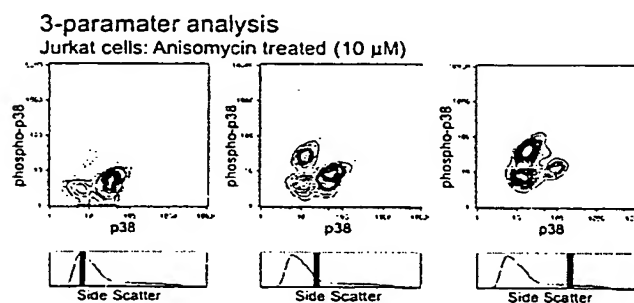
B.



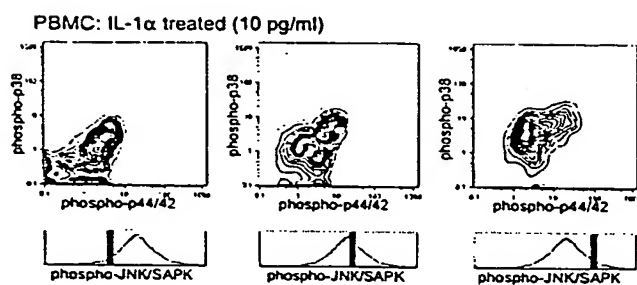
C.



D.



E.



F.

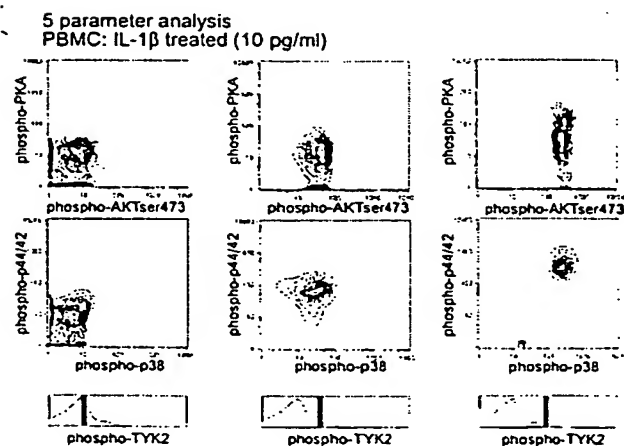


FIGURE 2

Multi-dimensional analysis provides insights into signaling networks as induced by differing stimuli.

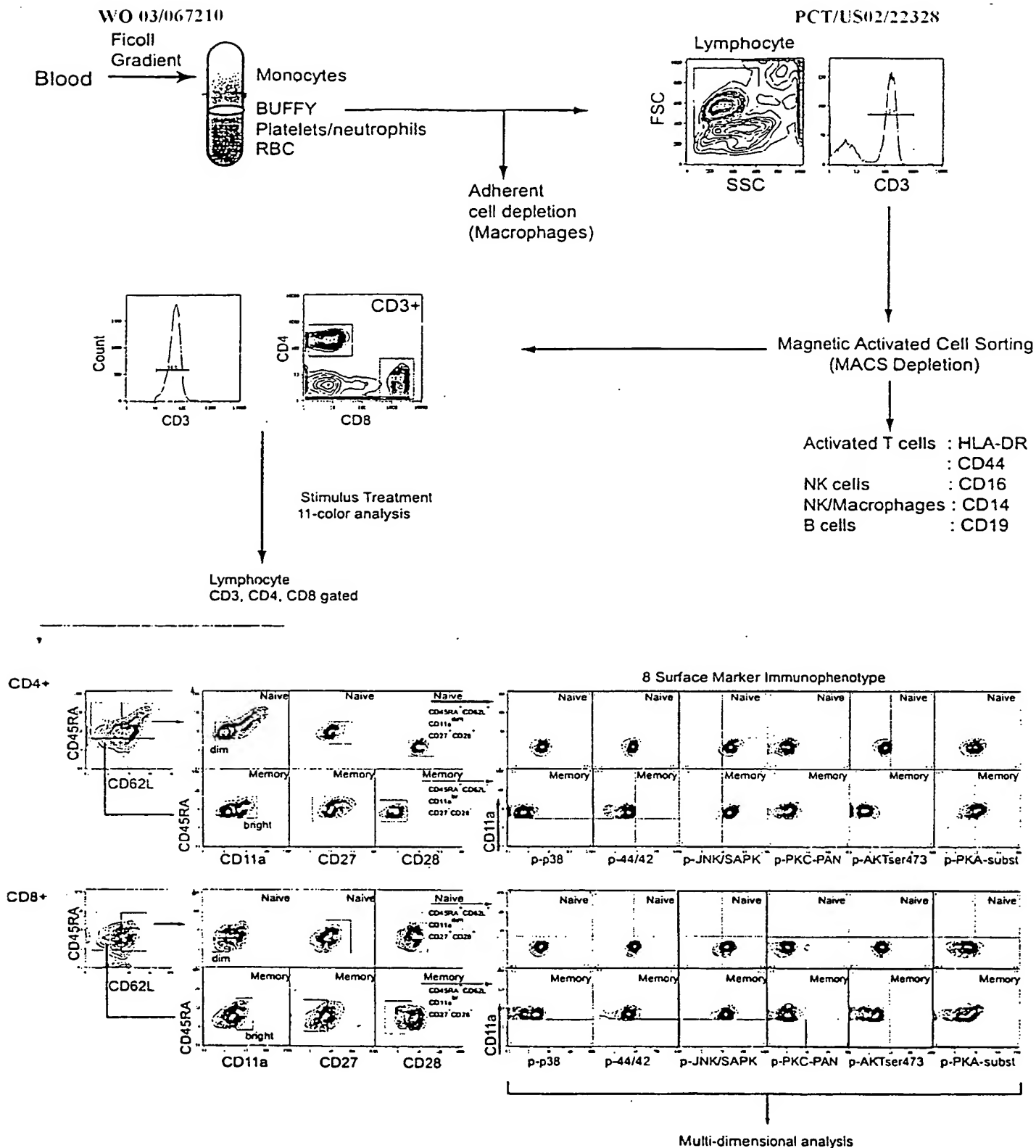


FIGURE 3

Flow diagram of resting T cell isolation, preparation, and analysis by polychromatic flow cytometry (11-color, 13 parameter).

CD4⁺ Naive: CD3⁺CD4⁺CD8⁻CD45⁺CD62L⁺CD11a^{dim}CD27⁺CD28⁺
 Memory: CD3⁺CD4⁺CD8⁻CD45⁺CD62L⁻CD11a^{br}CD27⁻CD28⁻

CD8⁺ Naive: CD3⁺CD4⁻CD8⁺CD45⁺CD62L⁺CD11a^{dim}CD27⁺CD28⁺
 Memory: CD3⁺CD4⁻CD8⁺CD45⁺CD62L⁻CD11a^{br}CD27⁻CD28⁻

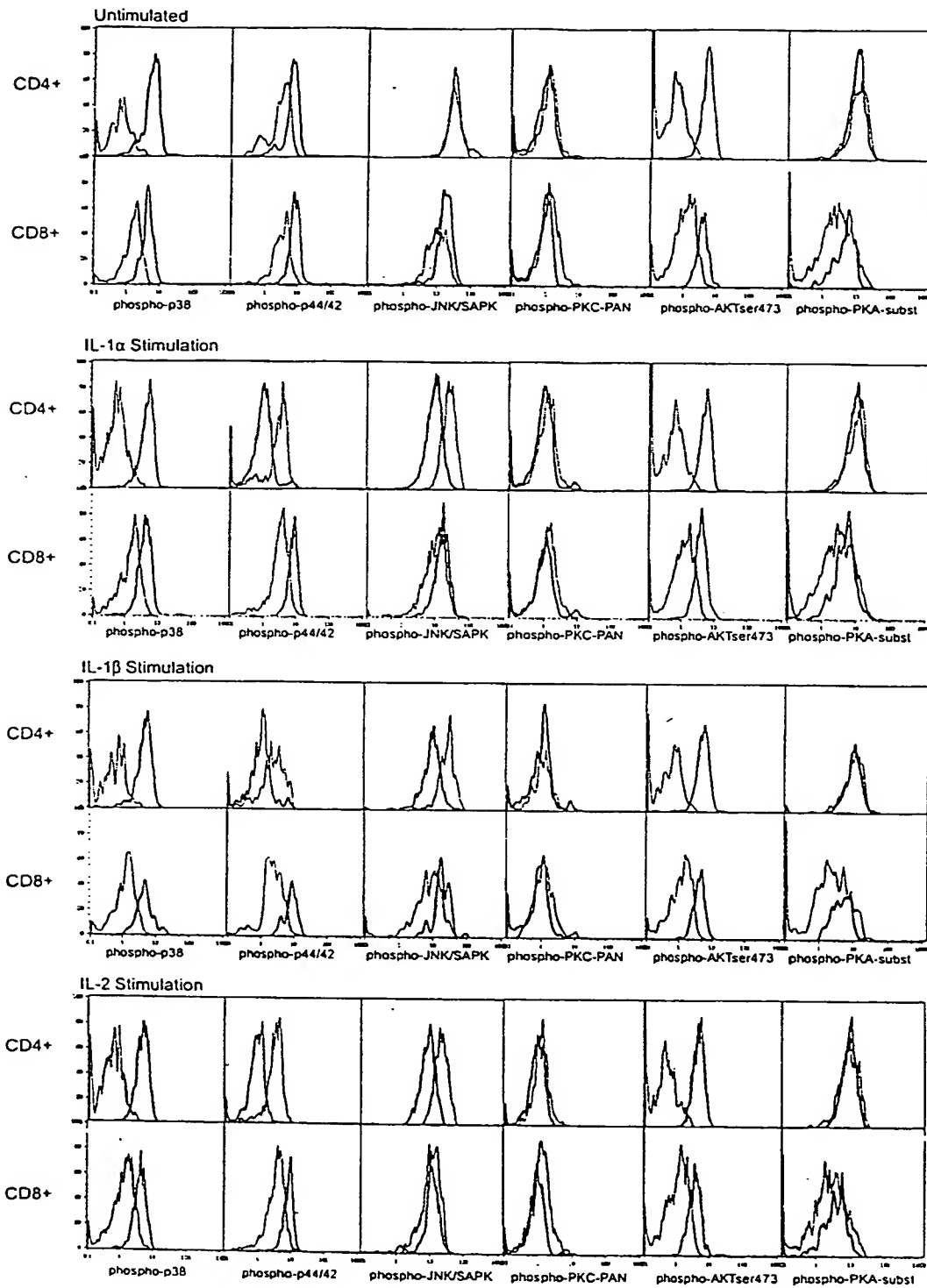


FIGURE 4

Signaling profiles are readily discernible in particular lymphocyte subsets using polychromatic flow cytometry.

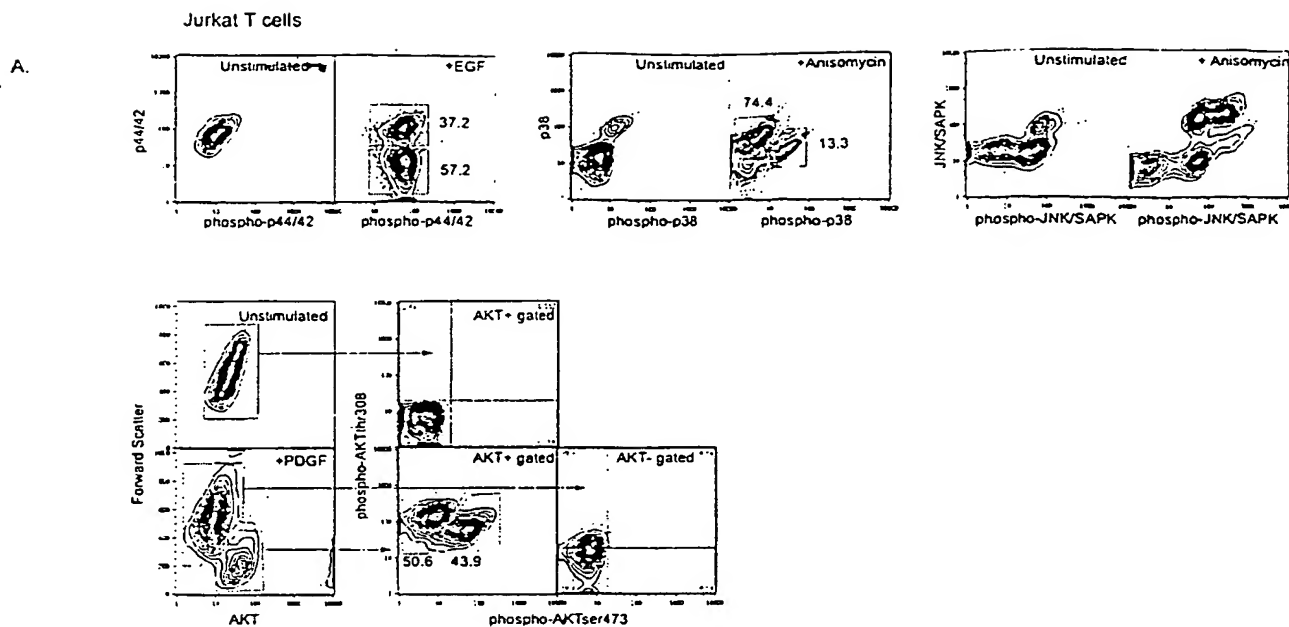


FIGURE 5

Monitoring intensities of both active and non-active kinases by flow cytometry.

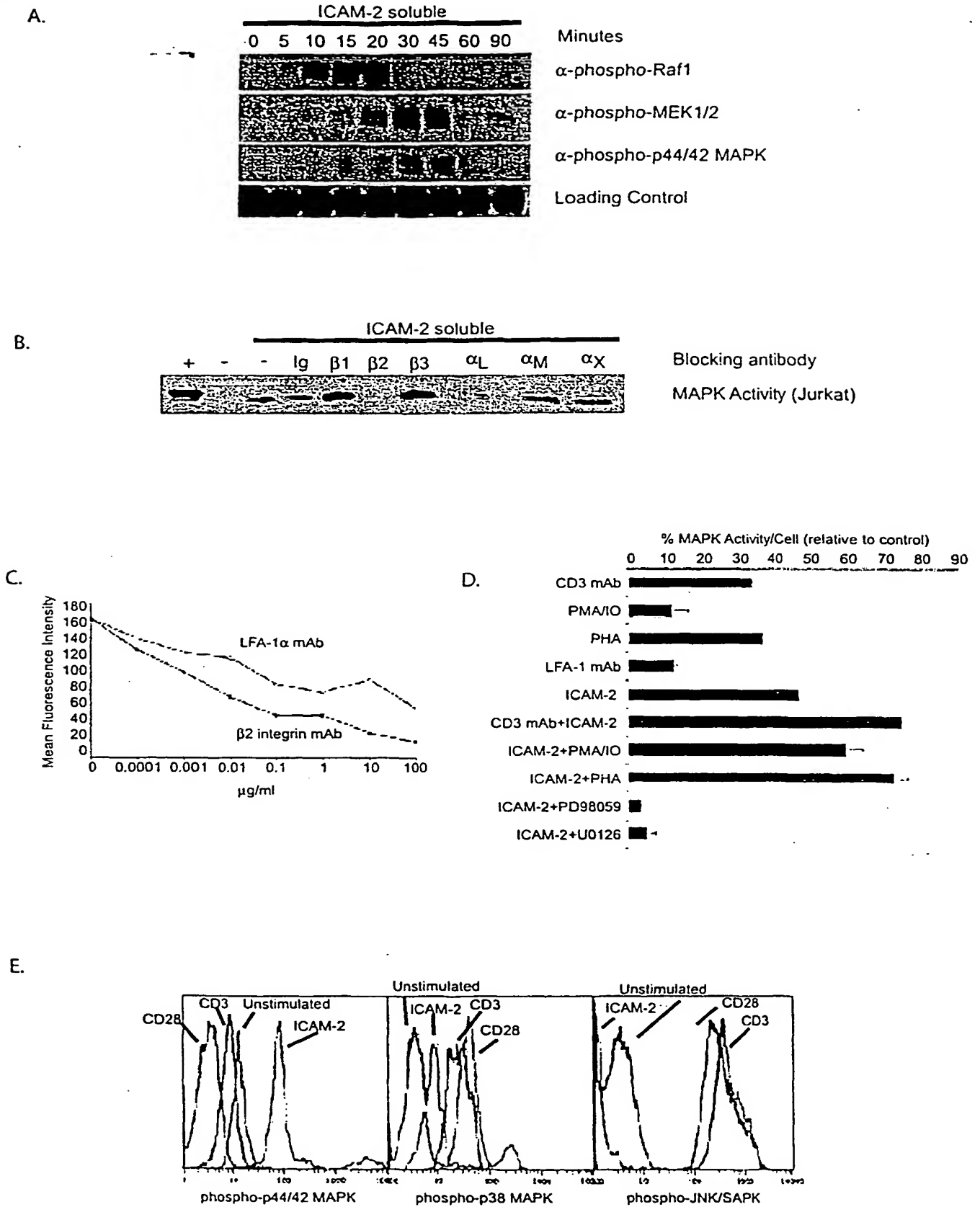
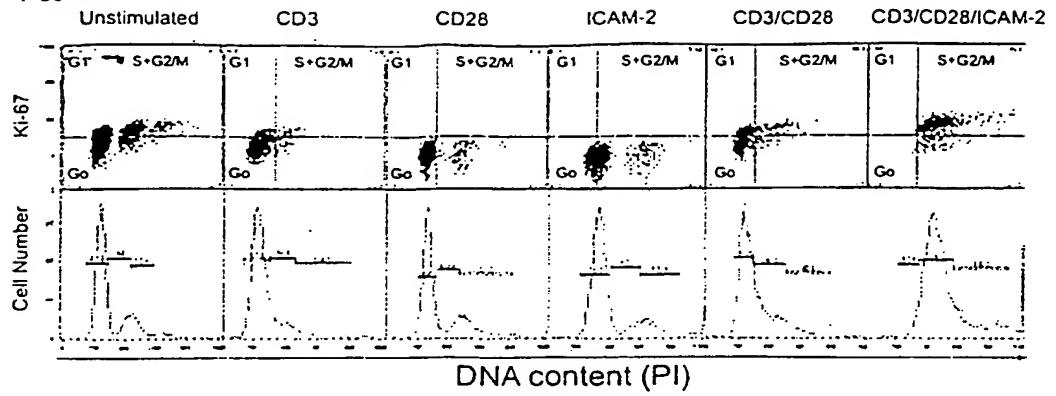


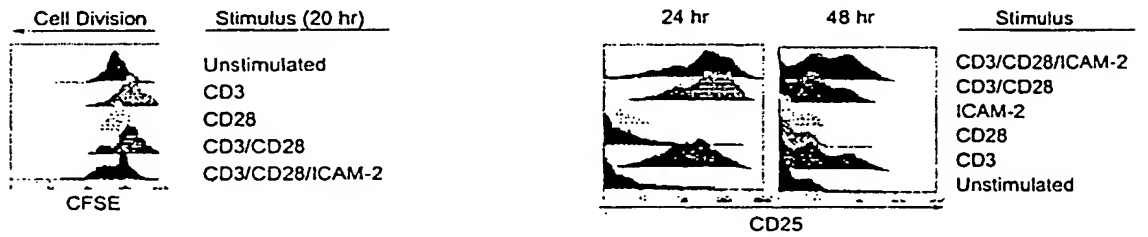
FIGURE 6

Figure 2. CD4+ T cells

A.



B.



C.

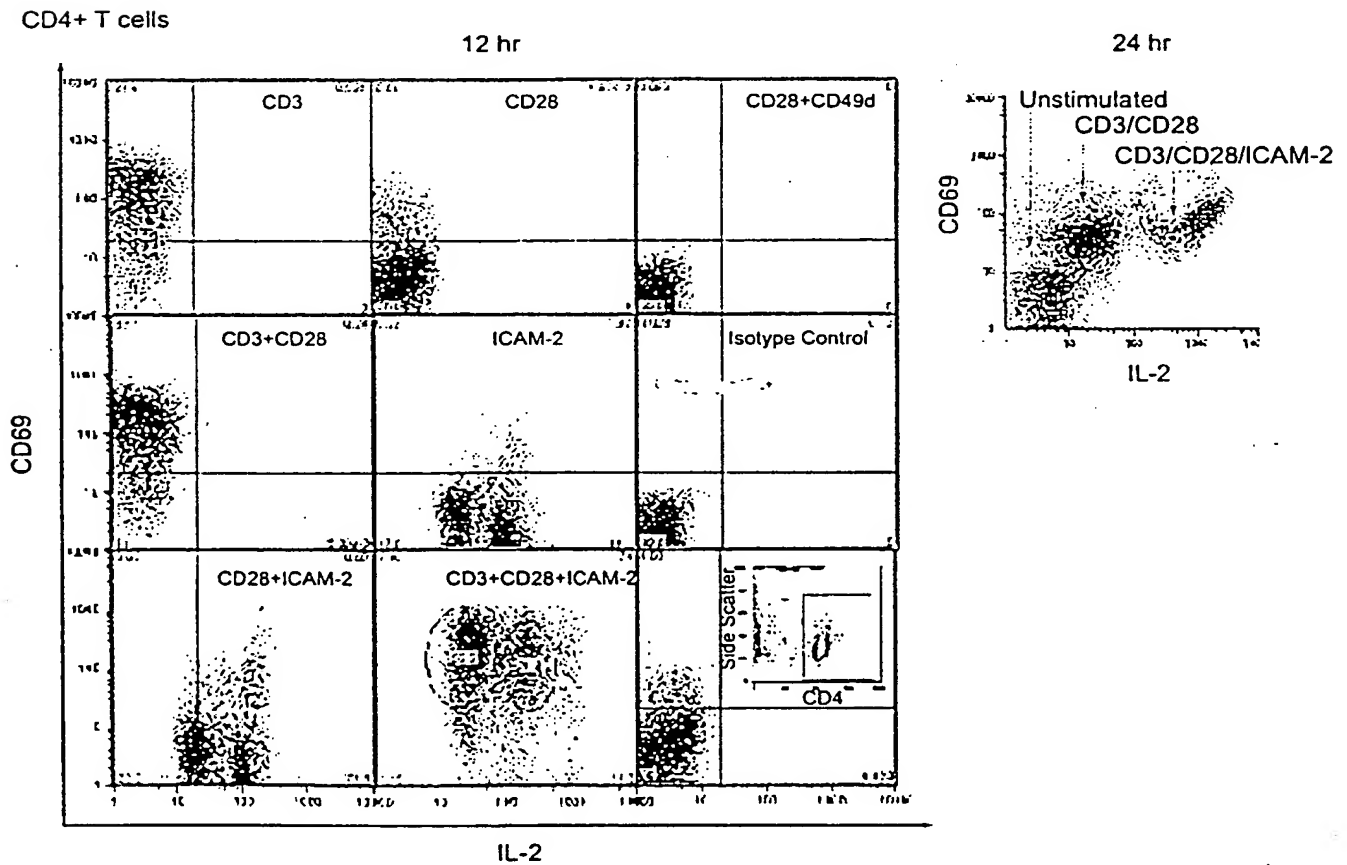
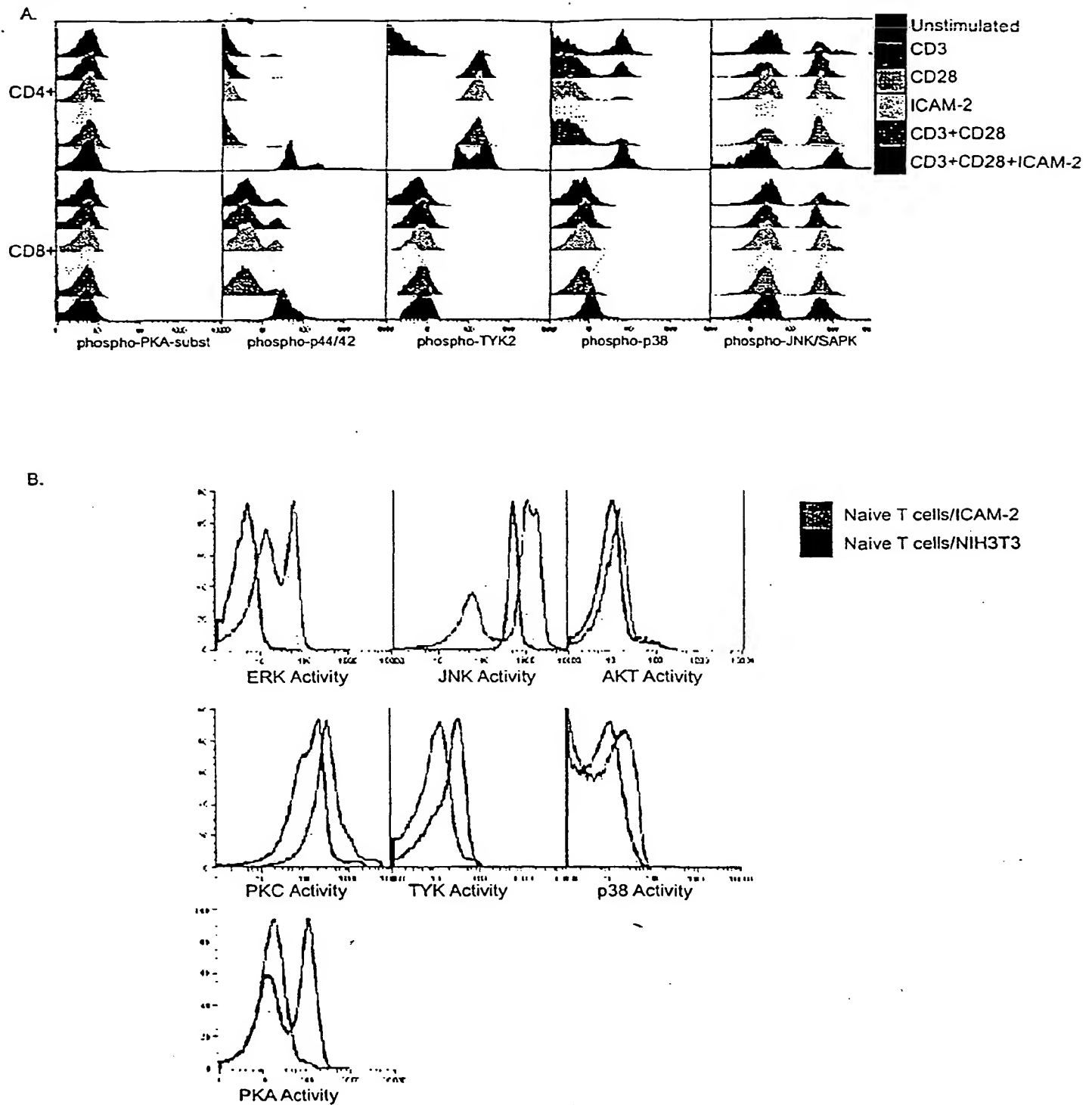


FIGURE 7

Figure 3.

**FIGURE 8**

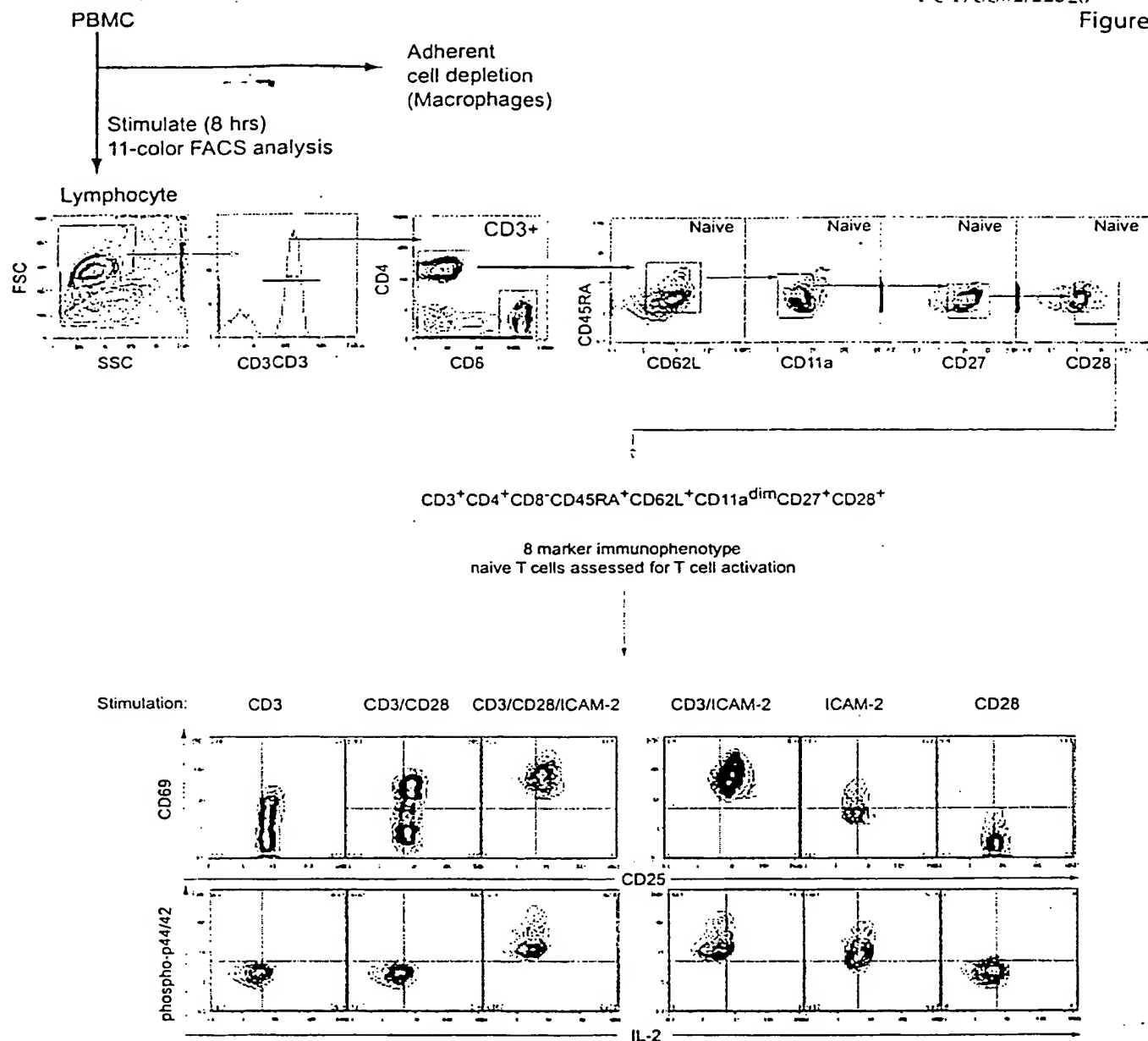
**FIGURE 9**

Figure 5.

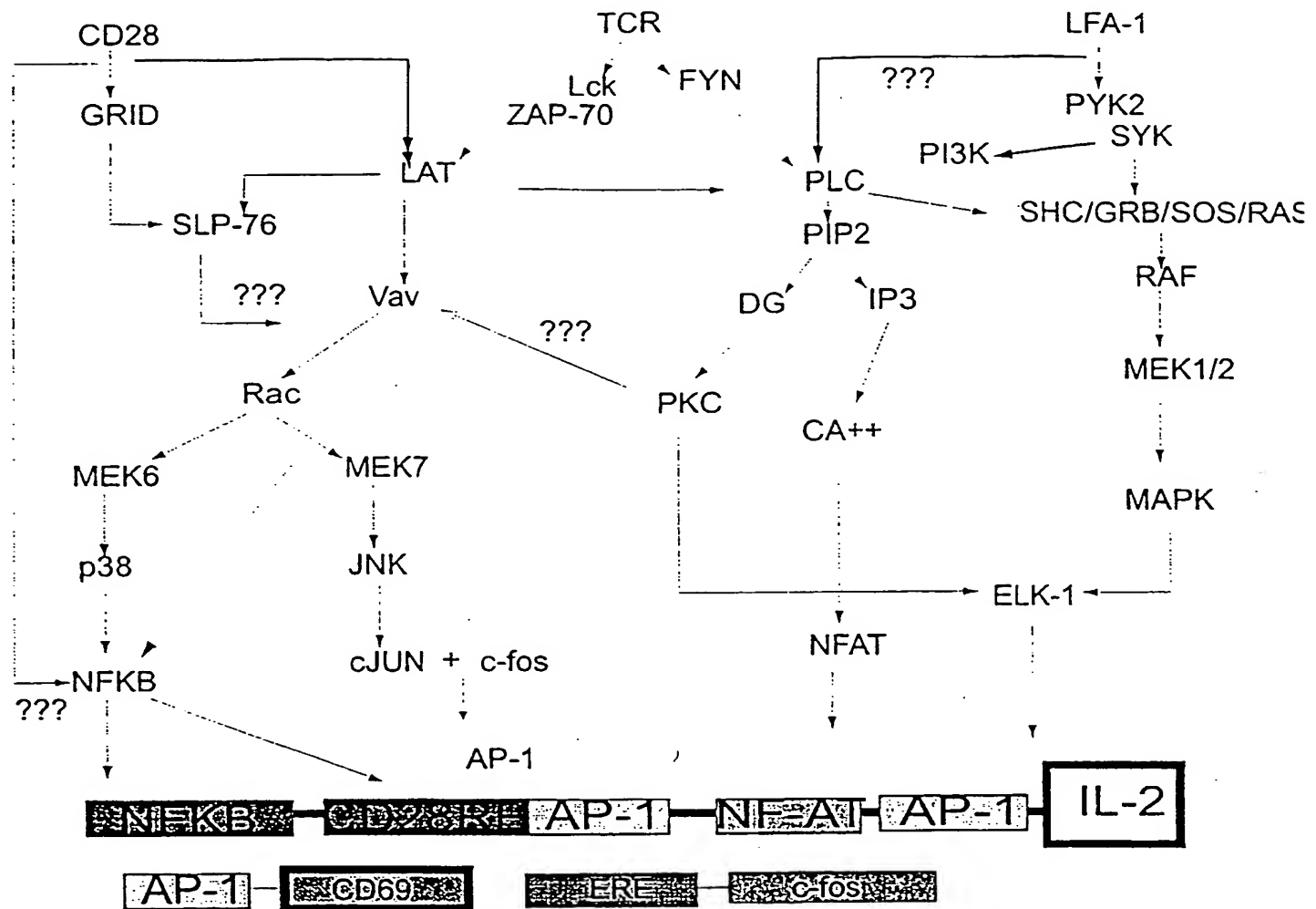
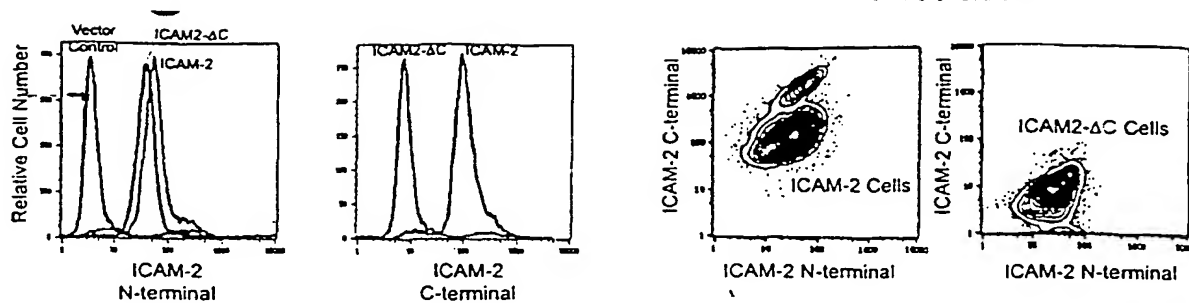
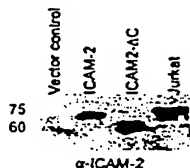


FIGURE 10

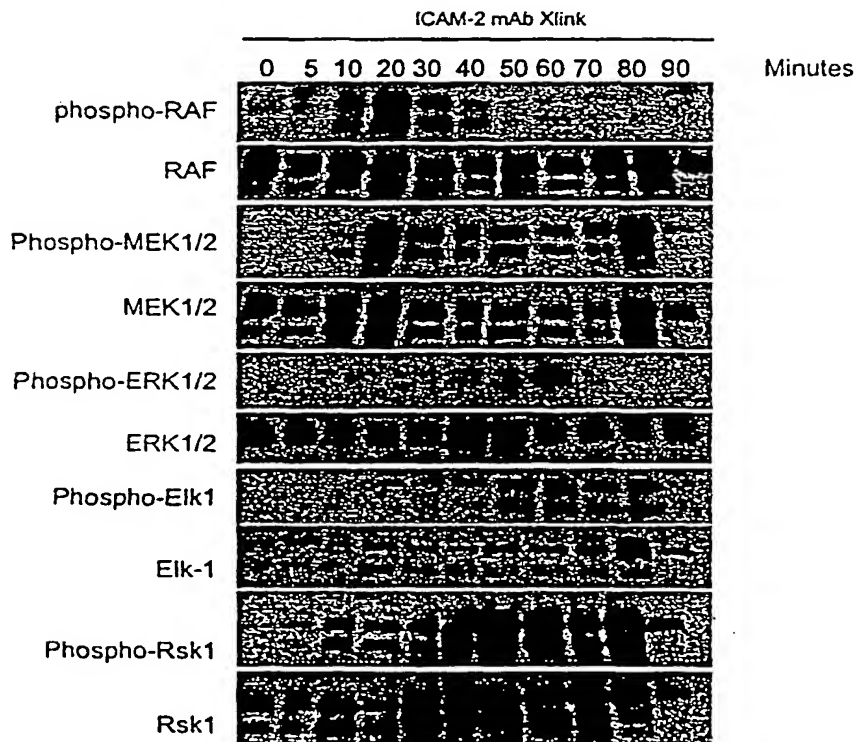
Figure 1.
A.



B.



C.



D.

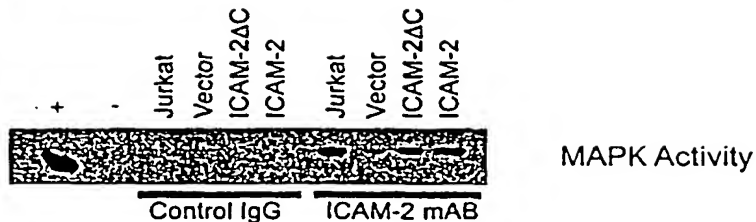
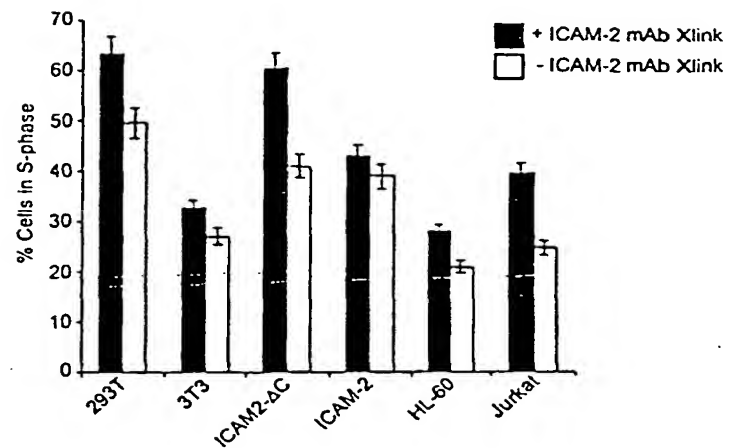


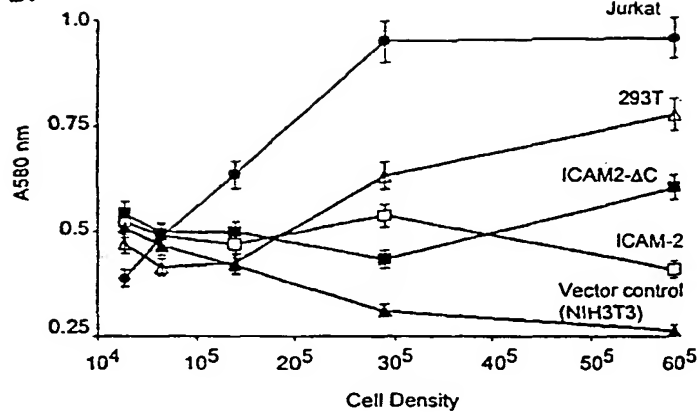
FIGURE 13

Figure 2.

A



B.



C.

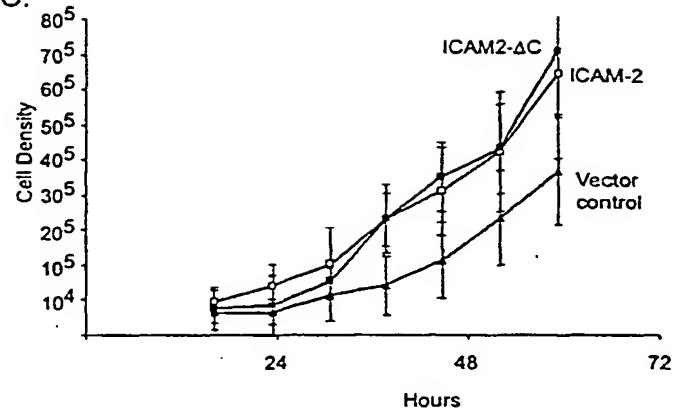


FIGURE 14

Figure 3.

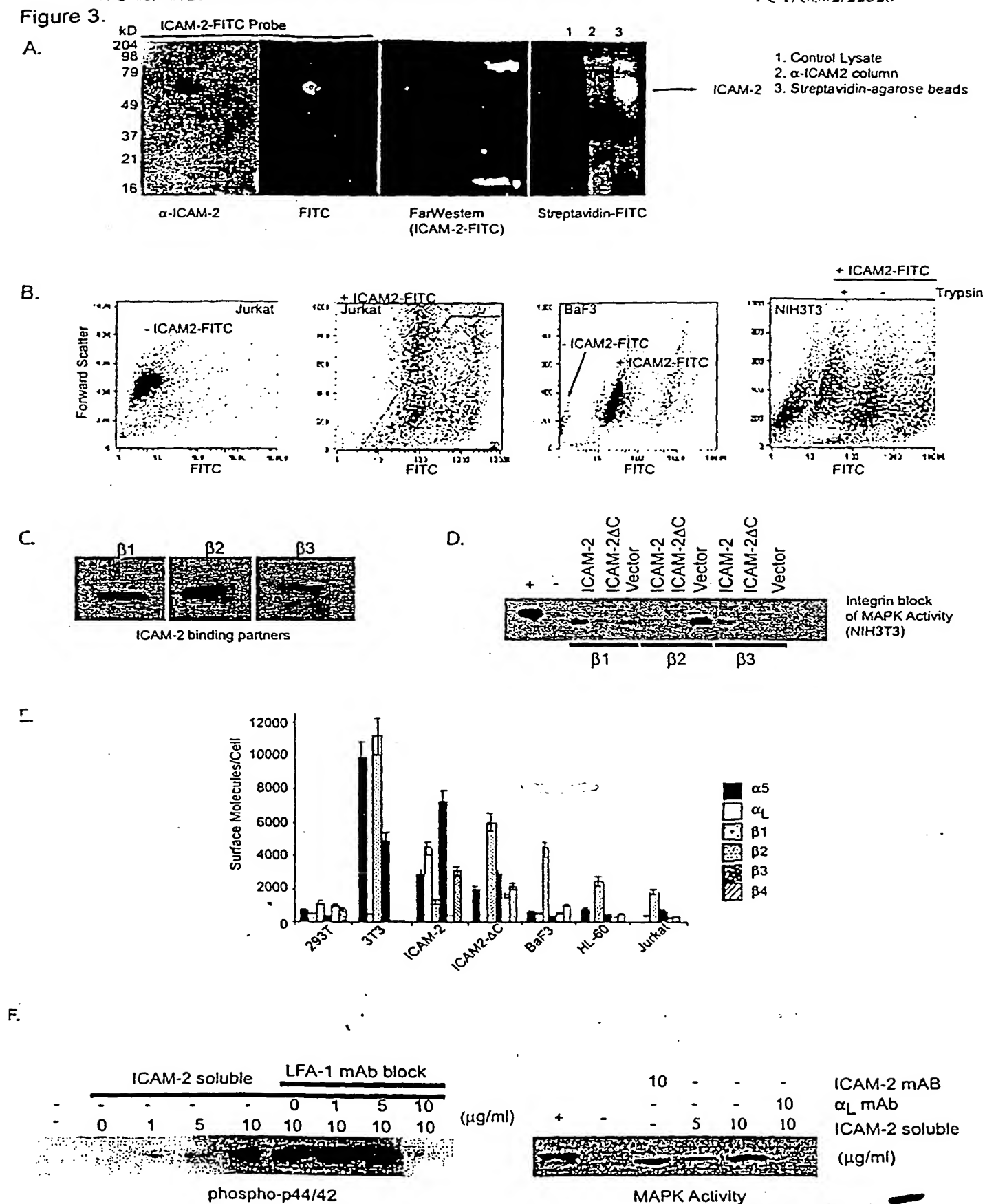
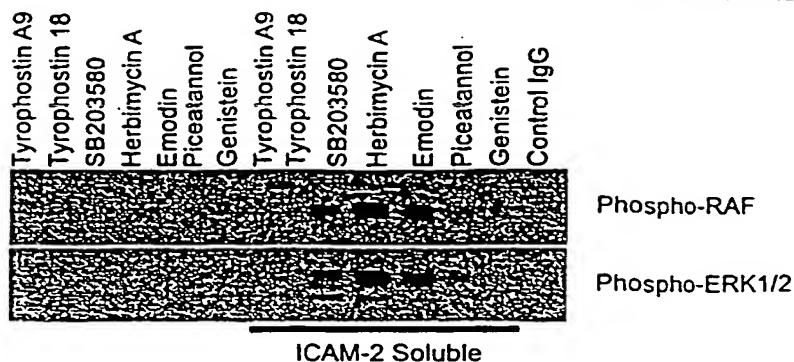


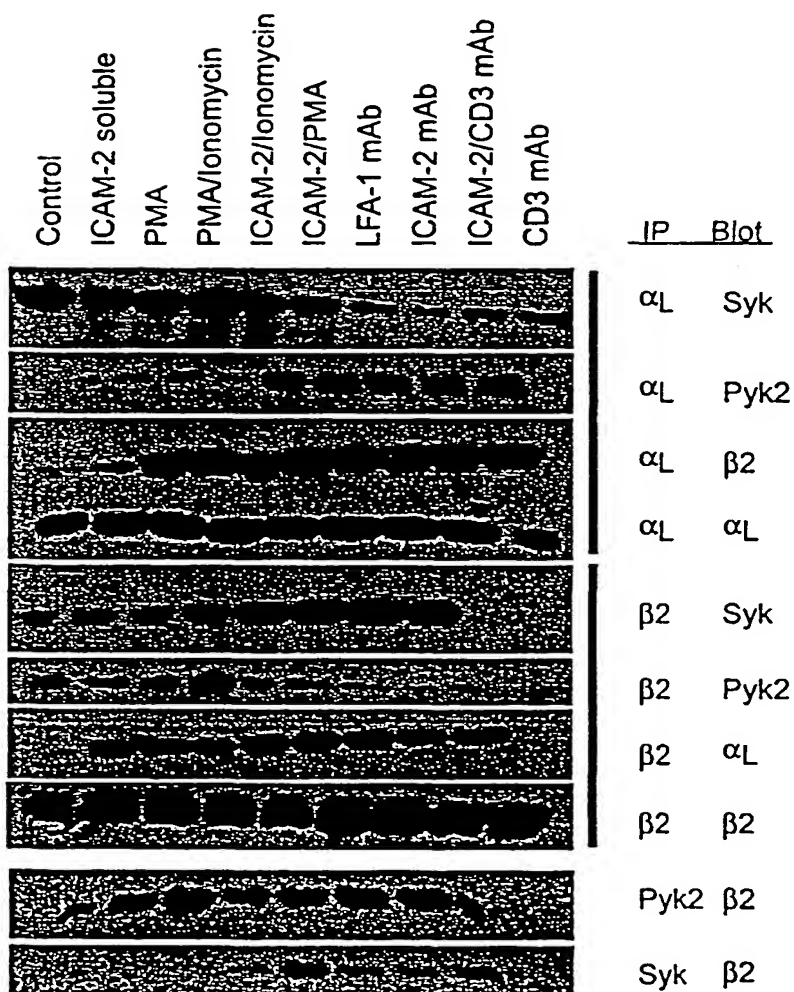
FIGURE 15

Figure 4.

A.



B.



C.



IP: Anti-phosphotyrosine
Blot: PYK2

FIGURE 16

A.

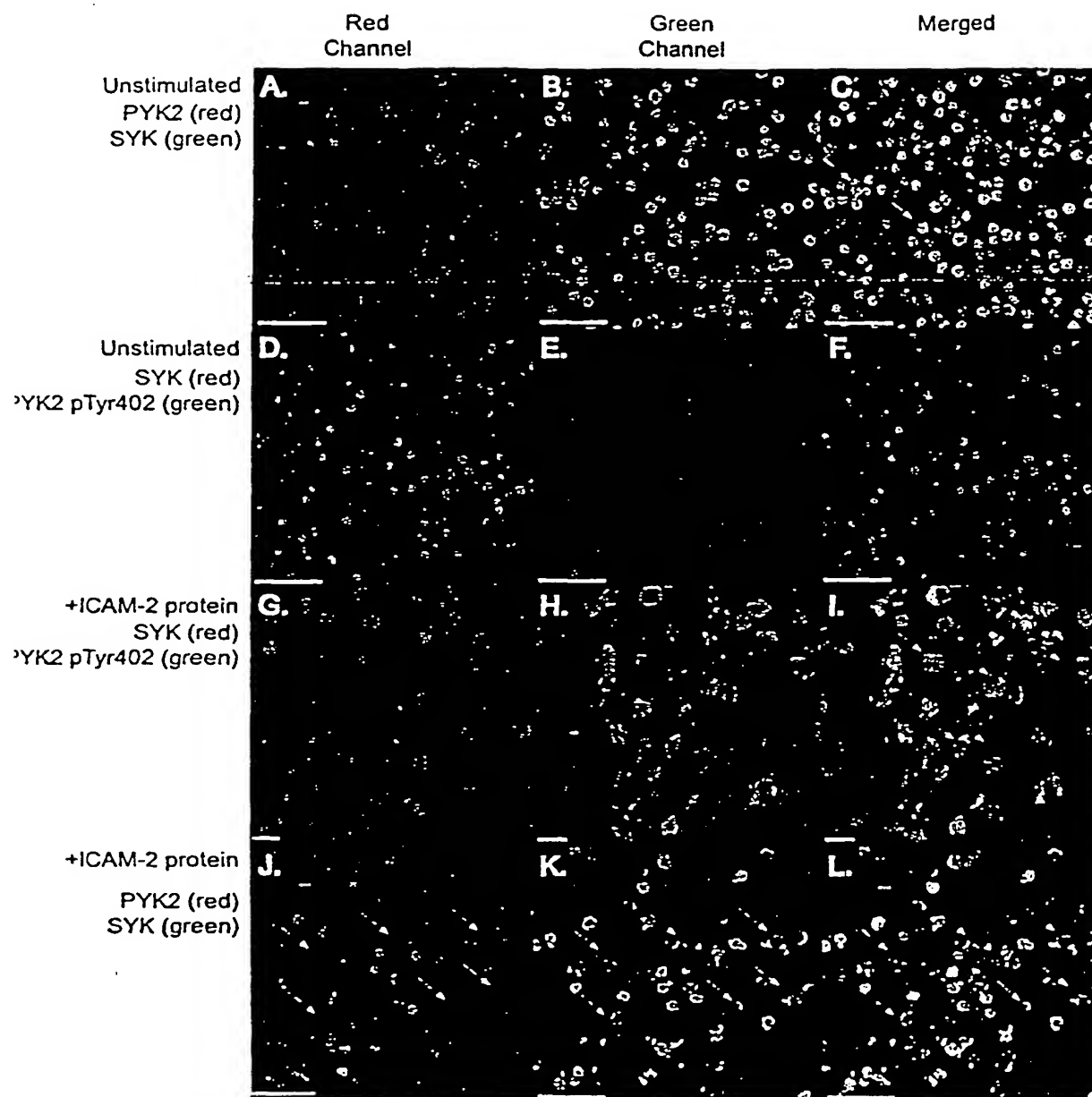
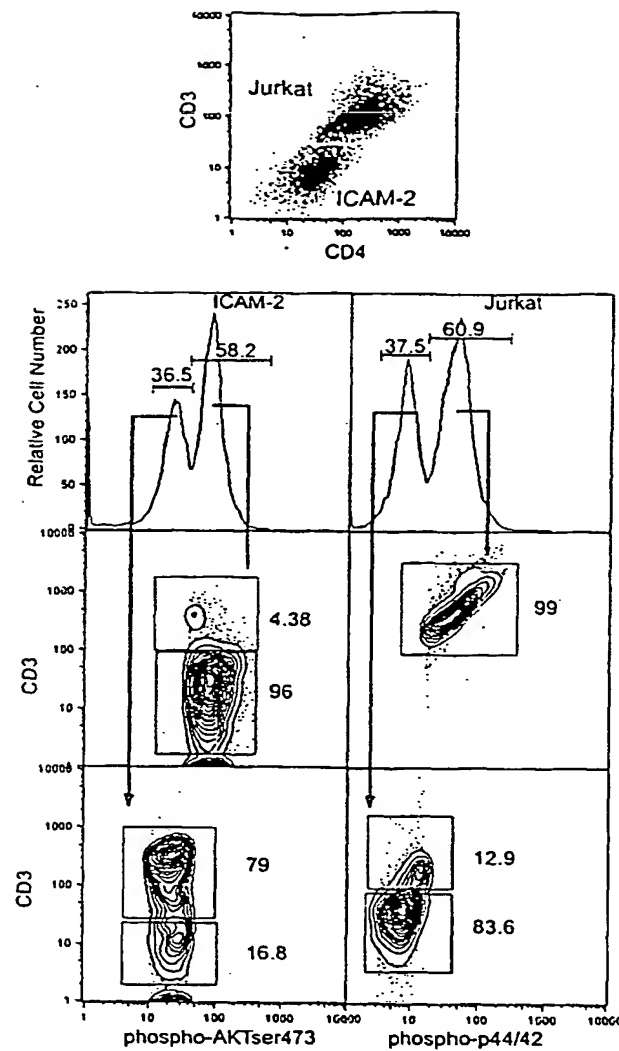


FIGURE 17

A.



B.

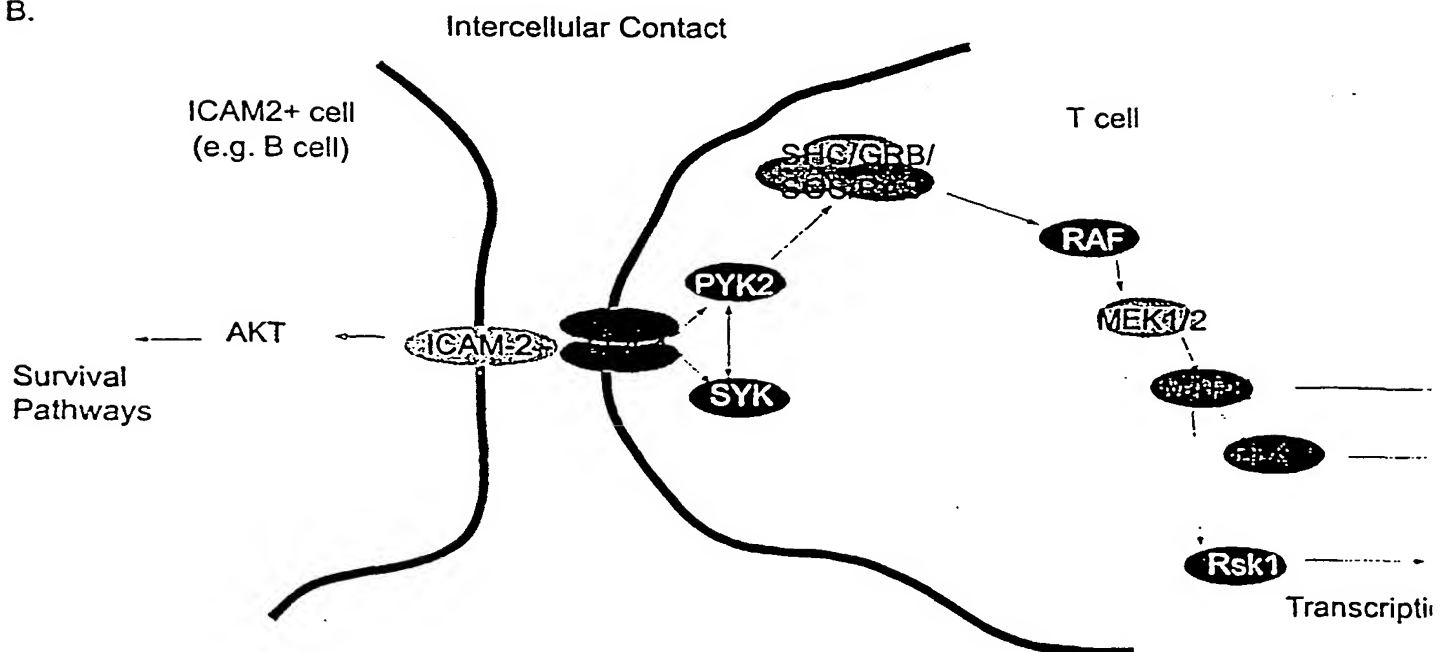
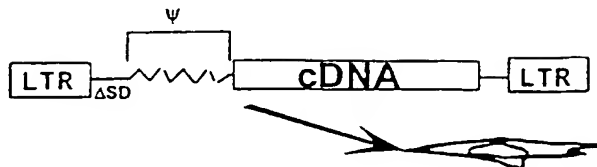


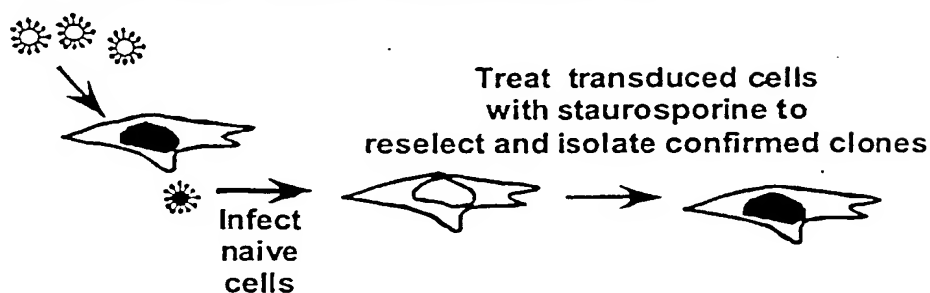
FIGURE 18

A.

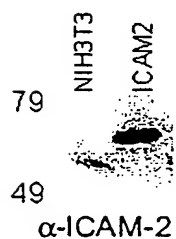
1. Infect 3T3 cells with cDNA library



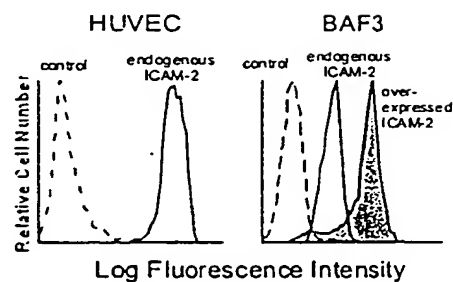
2. Treat with staurosporine for 24 hours. Grow out survivors for one week. Repeat x 3.
3. Infect surviving cells with MMLV to rescue proviruses as infectious particles.



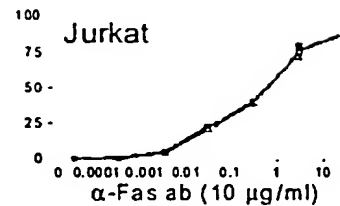
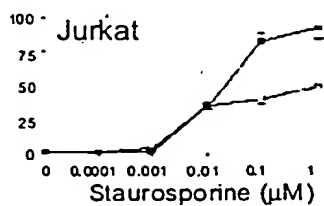
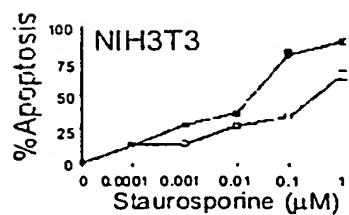
C.



D.

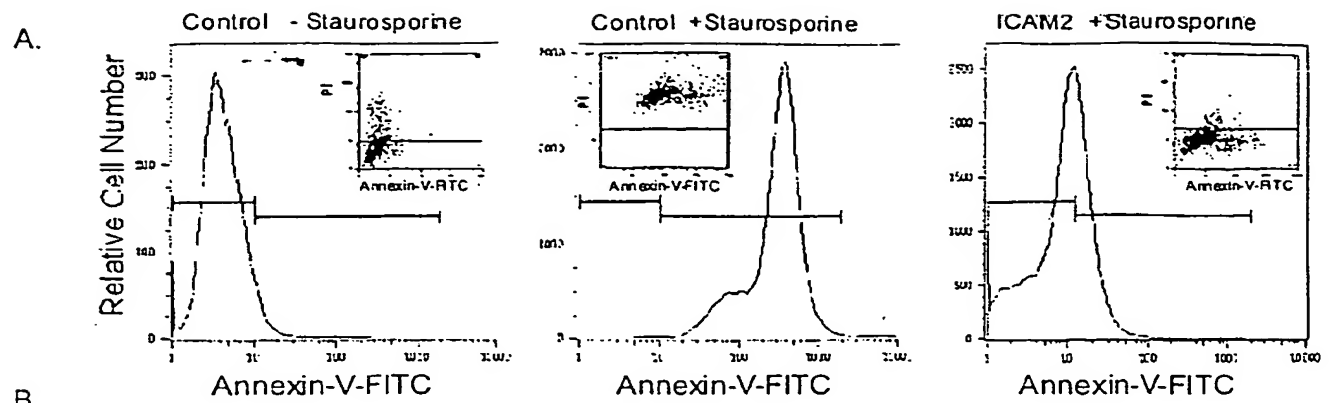


E.

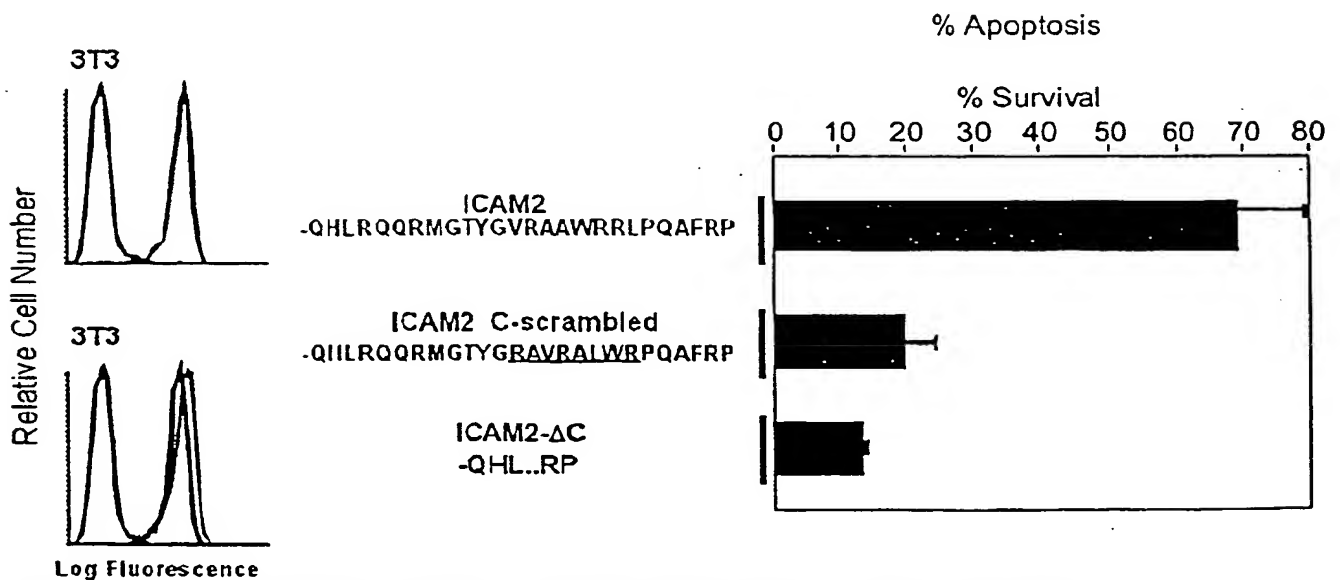


Retroviral library screen for anti-apoptotic molecules to staurosporine induced apoptosis identifies ICAM-2.

FIGURE 19



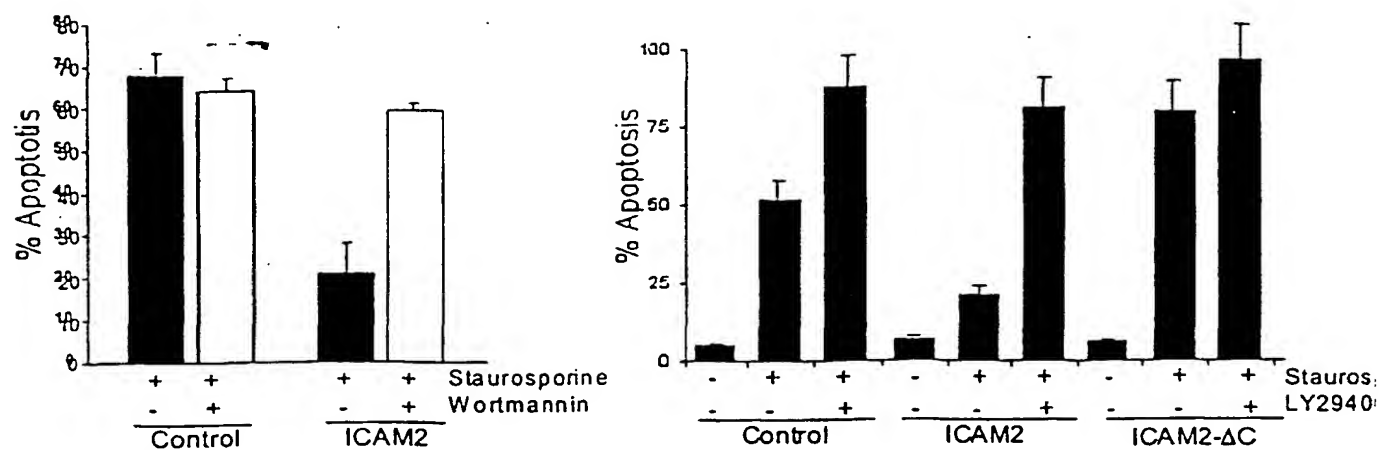
Cell Type/Infected virus	Apoptotic Inducer
Jurkat / ICAM2	α -FAS
Jurkat / Vector control	α -FAS
Jurkat / ICAM2	Staurosporine
Jurkat / Vector control	Staurosporine
NIH3T3 / Vector control	Staurosporine
NIH3T3 / ICAM2	Staurosporine
NIH3T3 / Bcl2	Staurosporine
70Z/3 / Vector control	Staurosporine
70Z/3 / ICAM2	Staurosporine
BAF3 / Vector control	IL-3 withdrawal
BAF3 / ICAM2	IL-3 withdrawal
BAF3 / Bcl2	IL-3 withdrawal
BAF3 / Vector	α -FAS
BAF3 / ICAM2	α -FAS
BAF3 / ICAM2 Δ N	α -FAS



Overexpression of ICAM-2 results in a broadly acting anti-apoptotic phenotype.

FIGURE 20

A.



B.



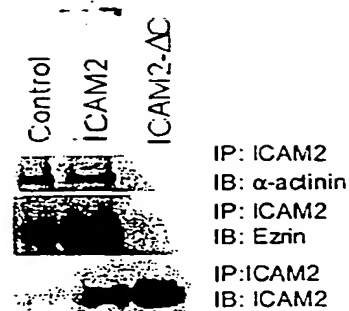
C.



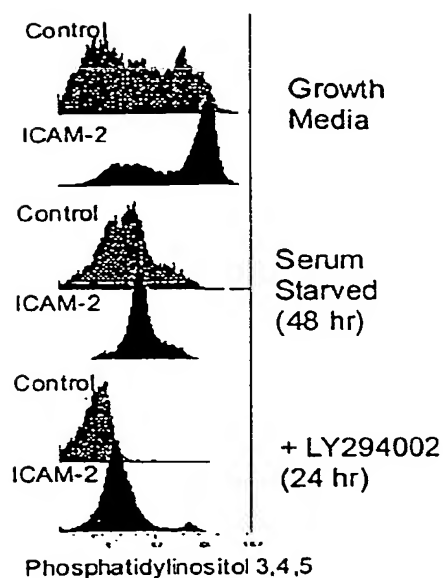
PI3 Kinase is necessary for ICAM-2's anti-apoptotic effect

FIGURE 21

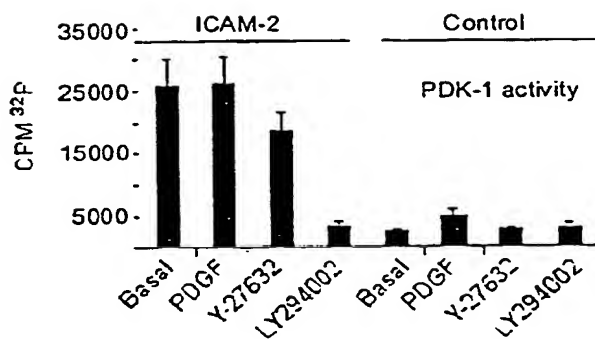
A.



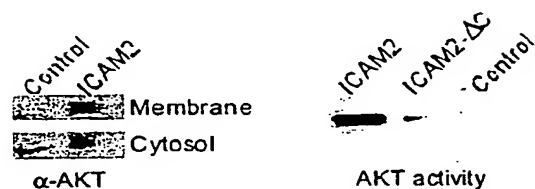
B.



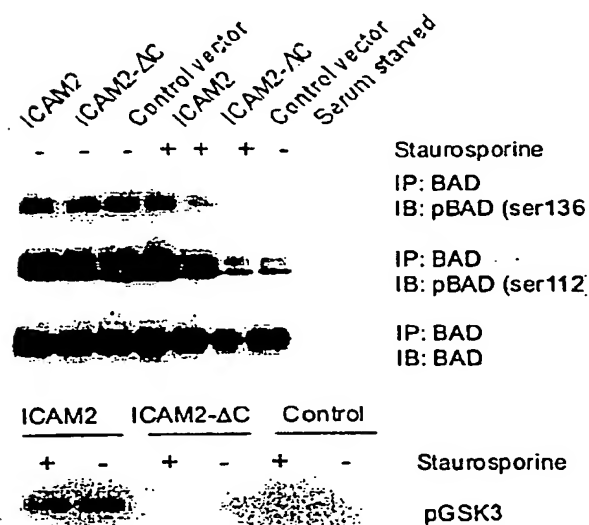
C.



D.

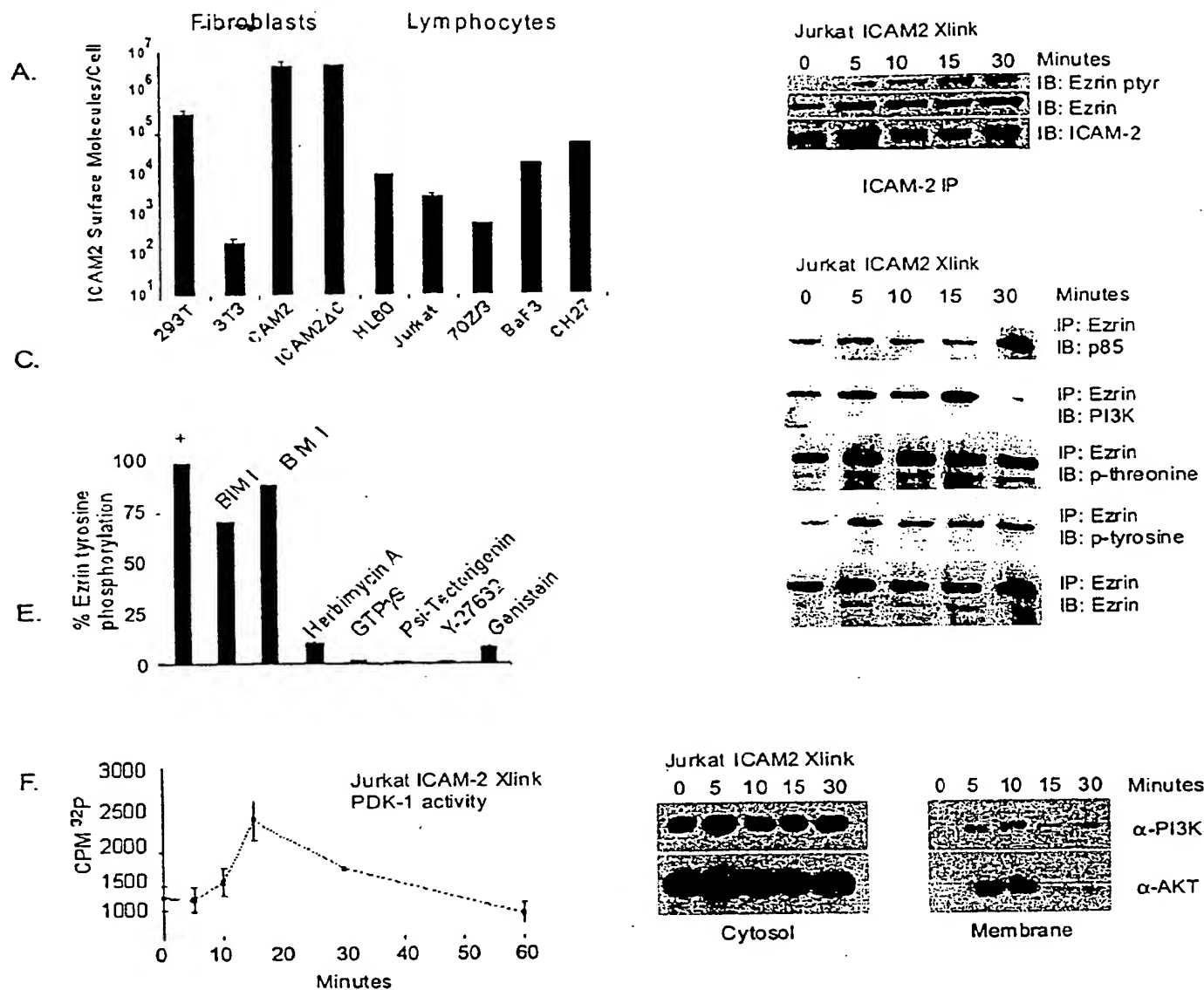


F.



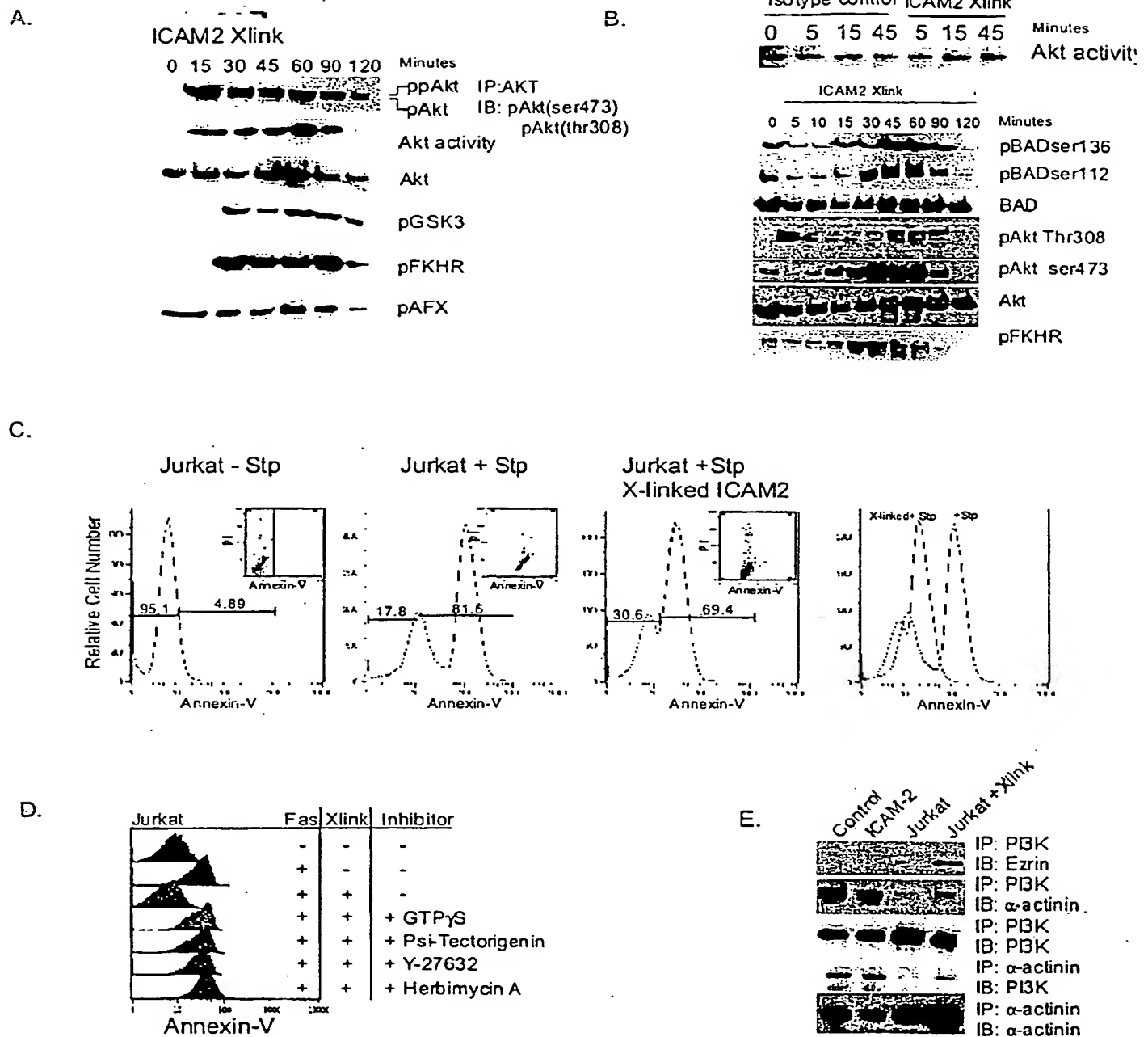
ICAM-2s signal results in phosphatidylinositol 3,4,5 production, PDK-1 and AKT activation

FIGURE 22



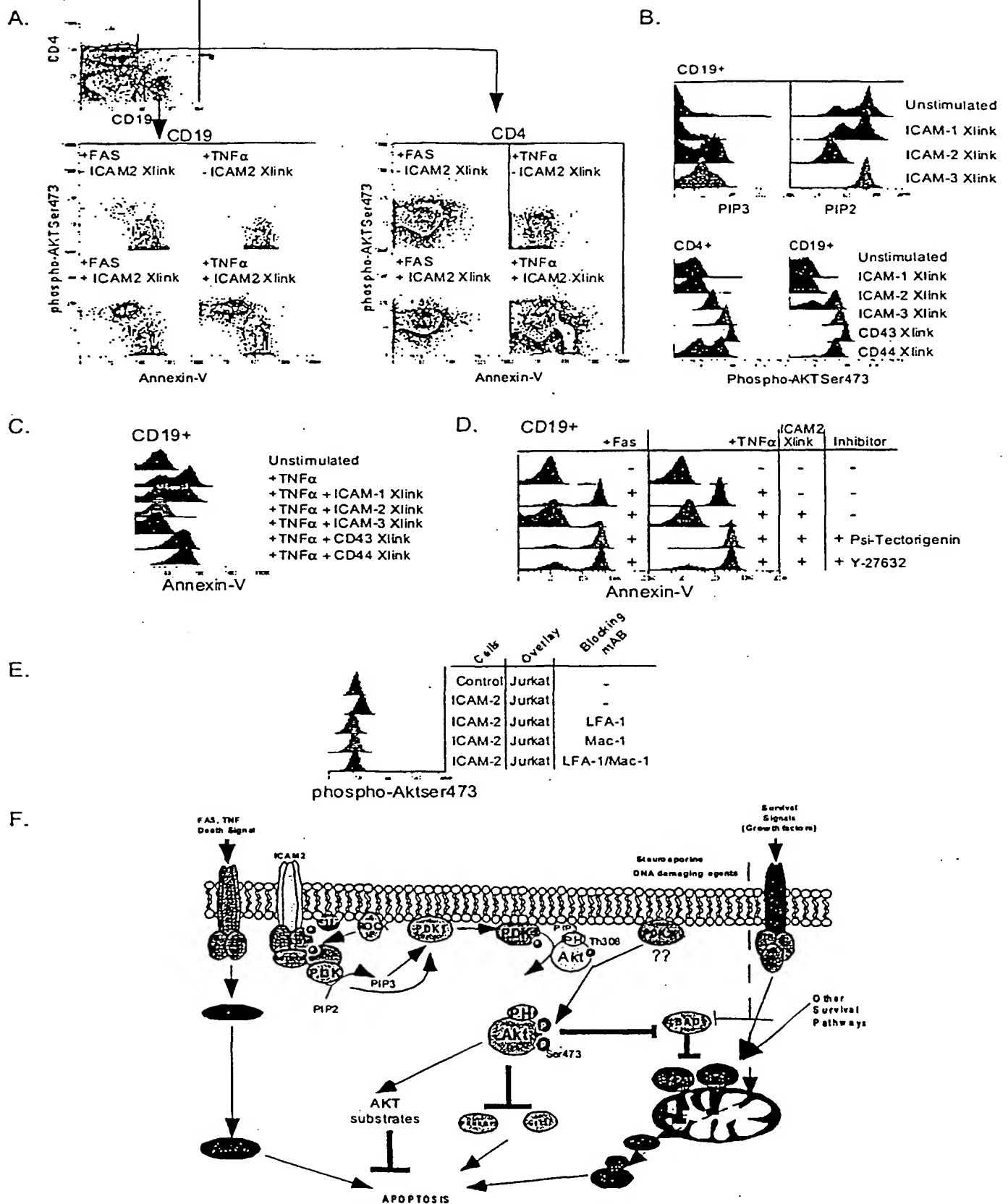
Initiation of endogenous ICAM-2 induces tyrosine phosphorylation of ezrin recruits

FIGURE 23



Ligation of endogenous ICAM-2 induces AKT activation in lymphoid cells conferring

FIGURE 24

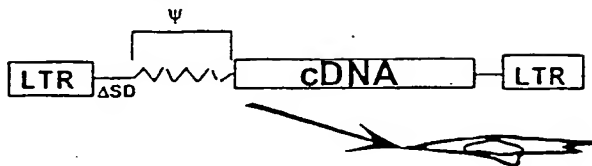


Resistance to TNF α and FAS induced apoptosis in primary human CD4 $^{+}$ and CD19 $^{+}$ cells is mediated by active AKT following ICAM-2 crosslinking.

FIGURE 25

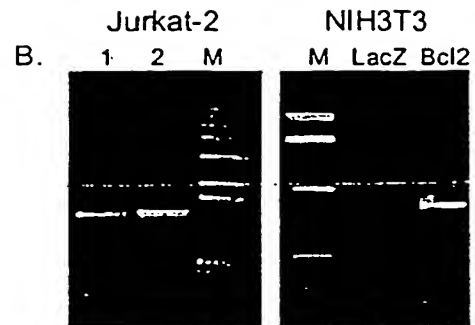
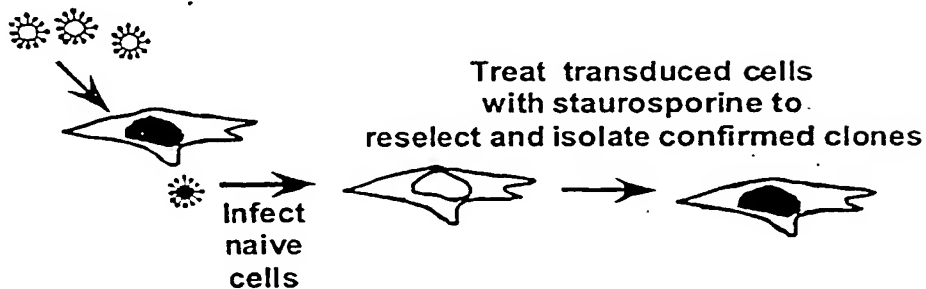
A.

1. Infect 3T3 cells with cDNA library



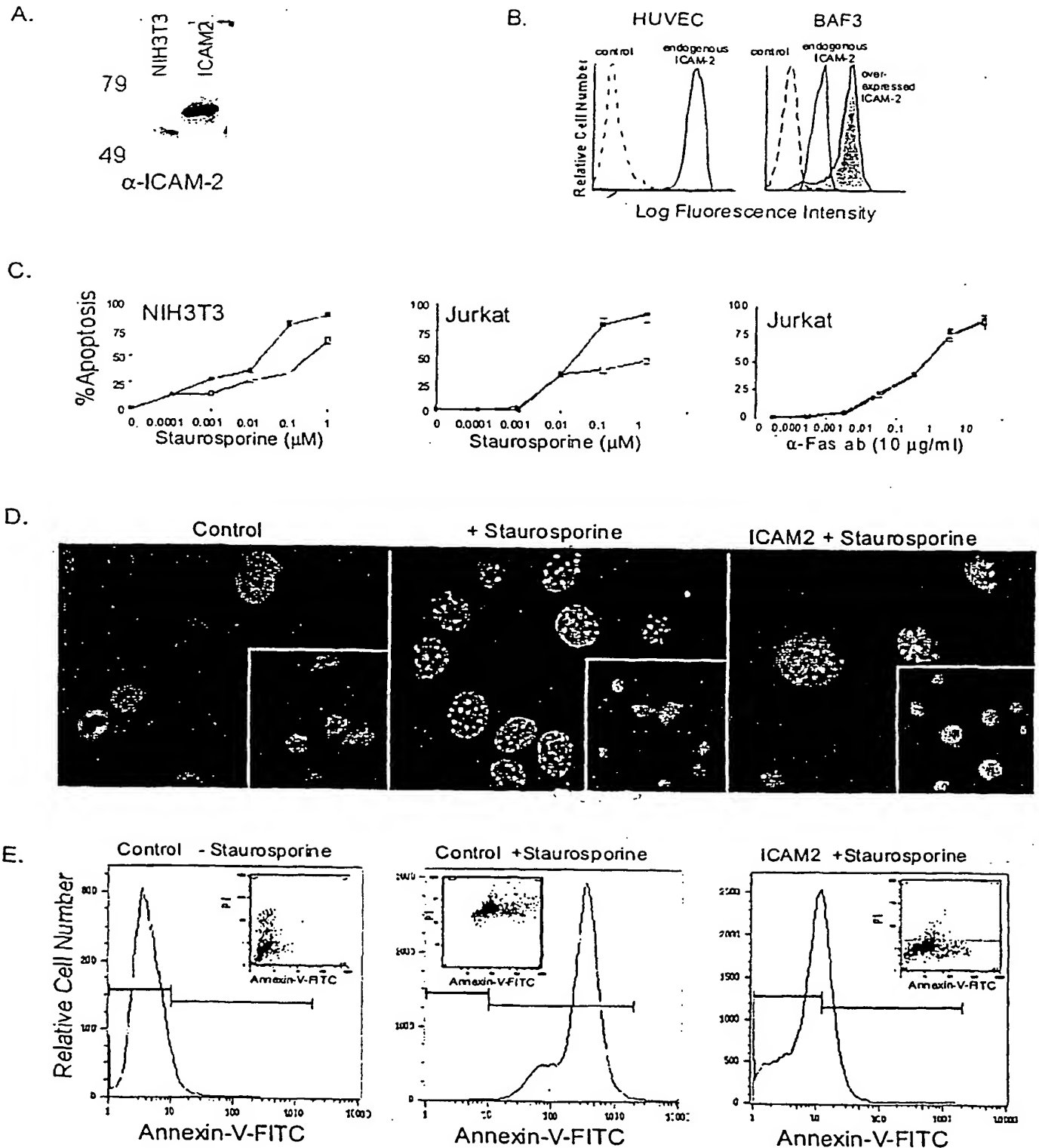
2. Treat with staurosporine for 24 hours. Grow out survivors for one week. Repeat x 3.

3. Infect surviving cells with MMLV to rescue proviruses as infectious particles.



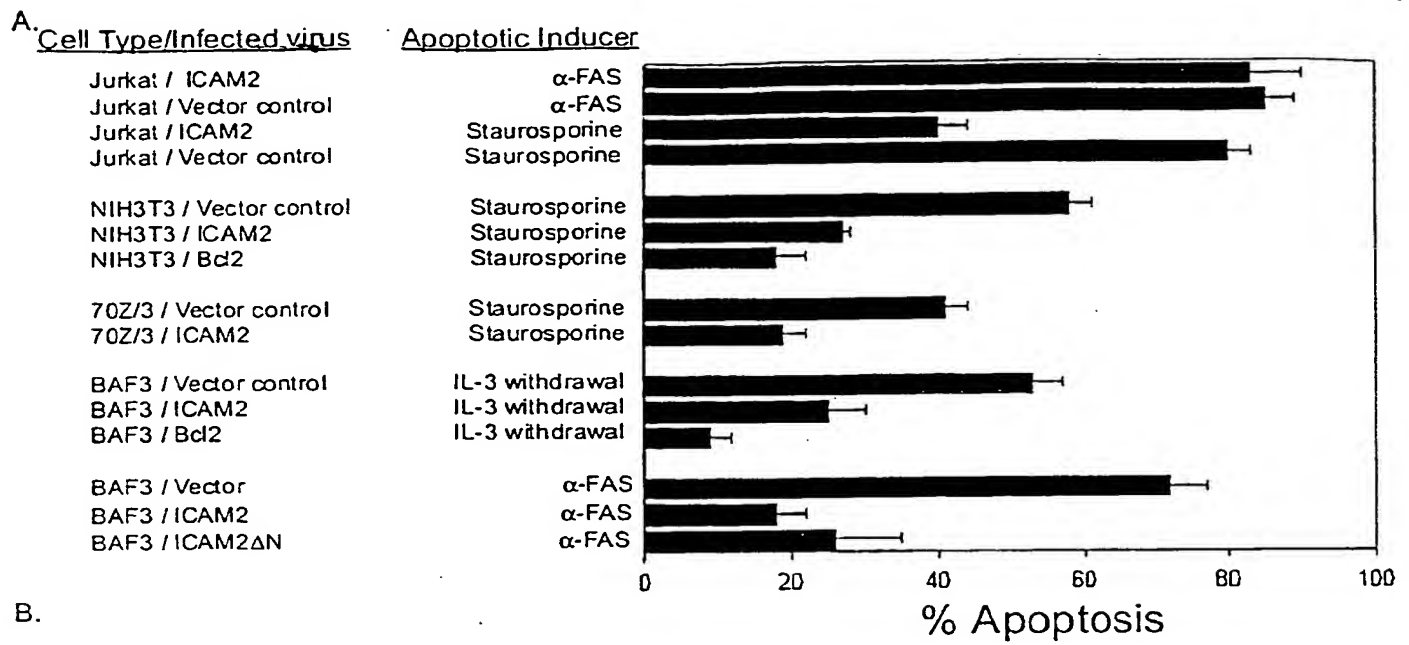
Retroviral library screen for anti-apoptotic molecules to staurosporine induced apoptosis.

FIGURE 26

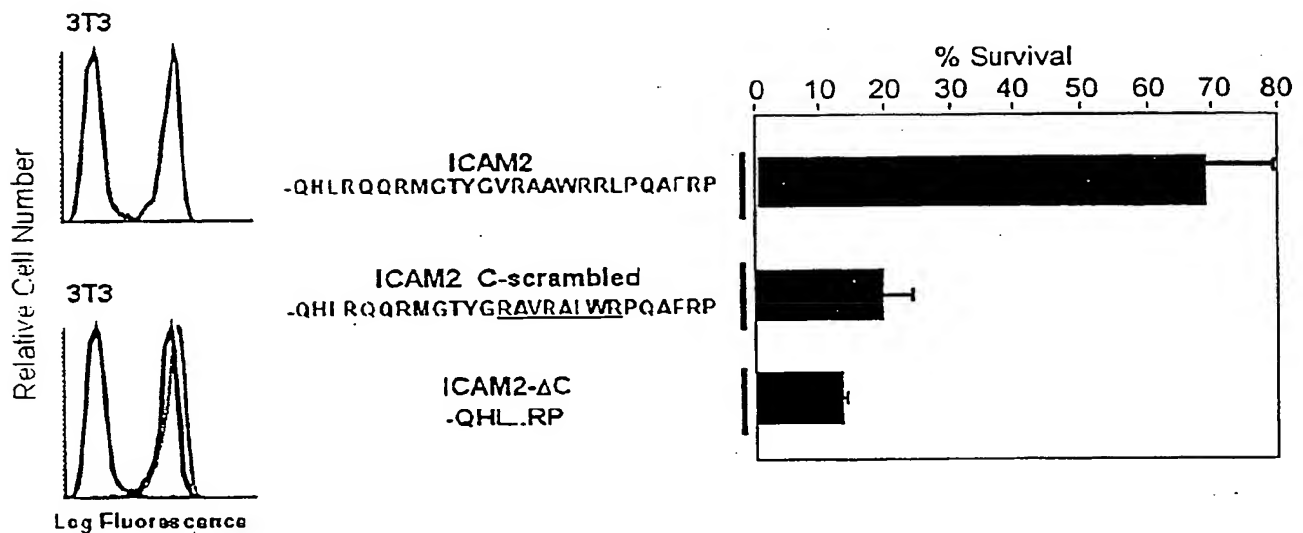


Overexpression of ICAM-2 inhibits staurosporine induced apoptosis.

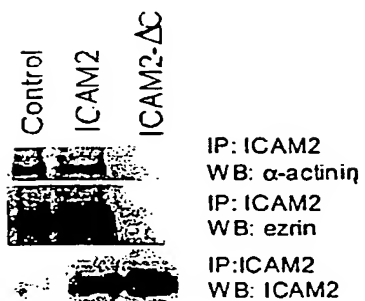
FIGURE 27



B.

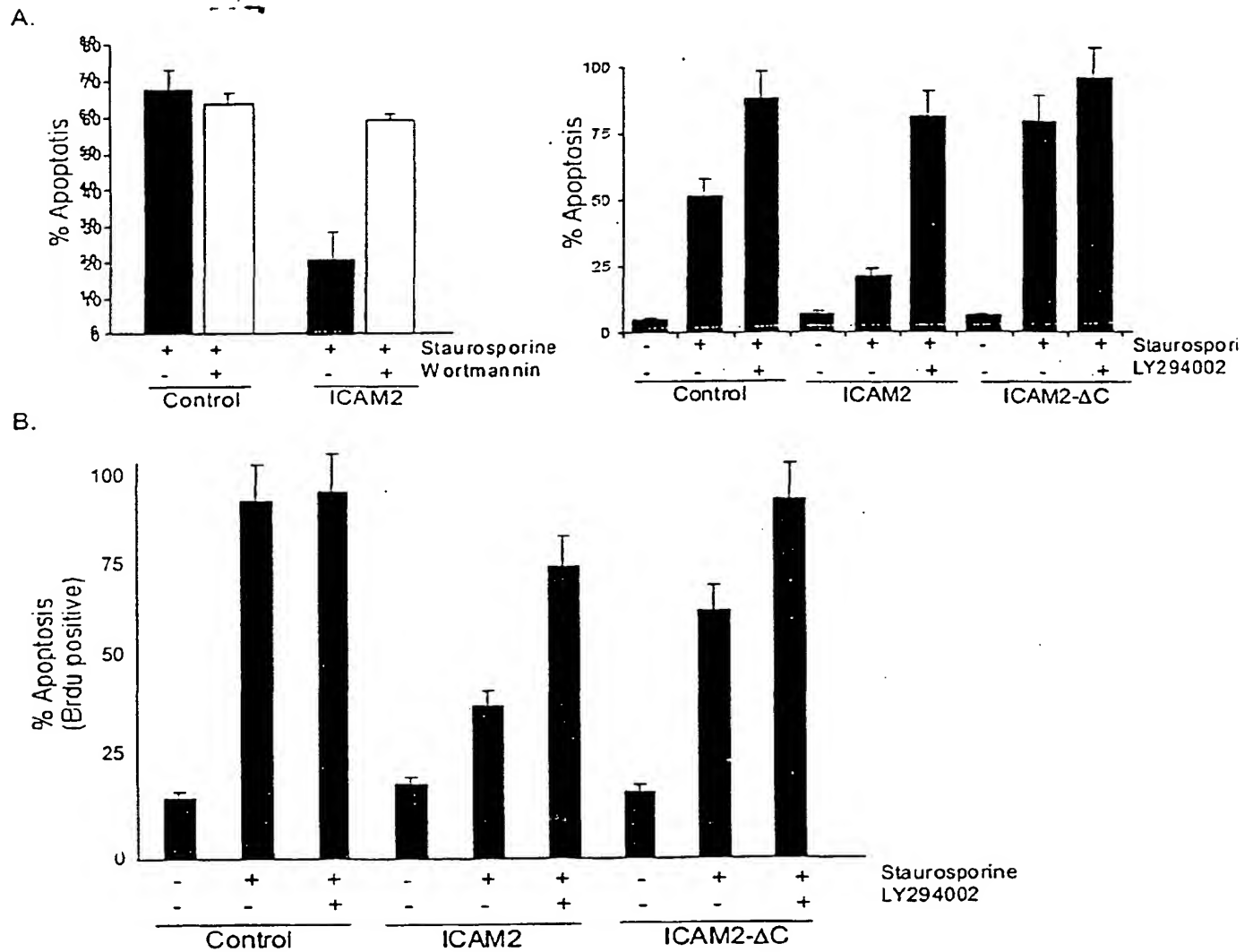


C.



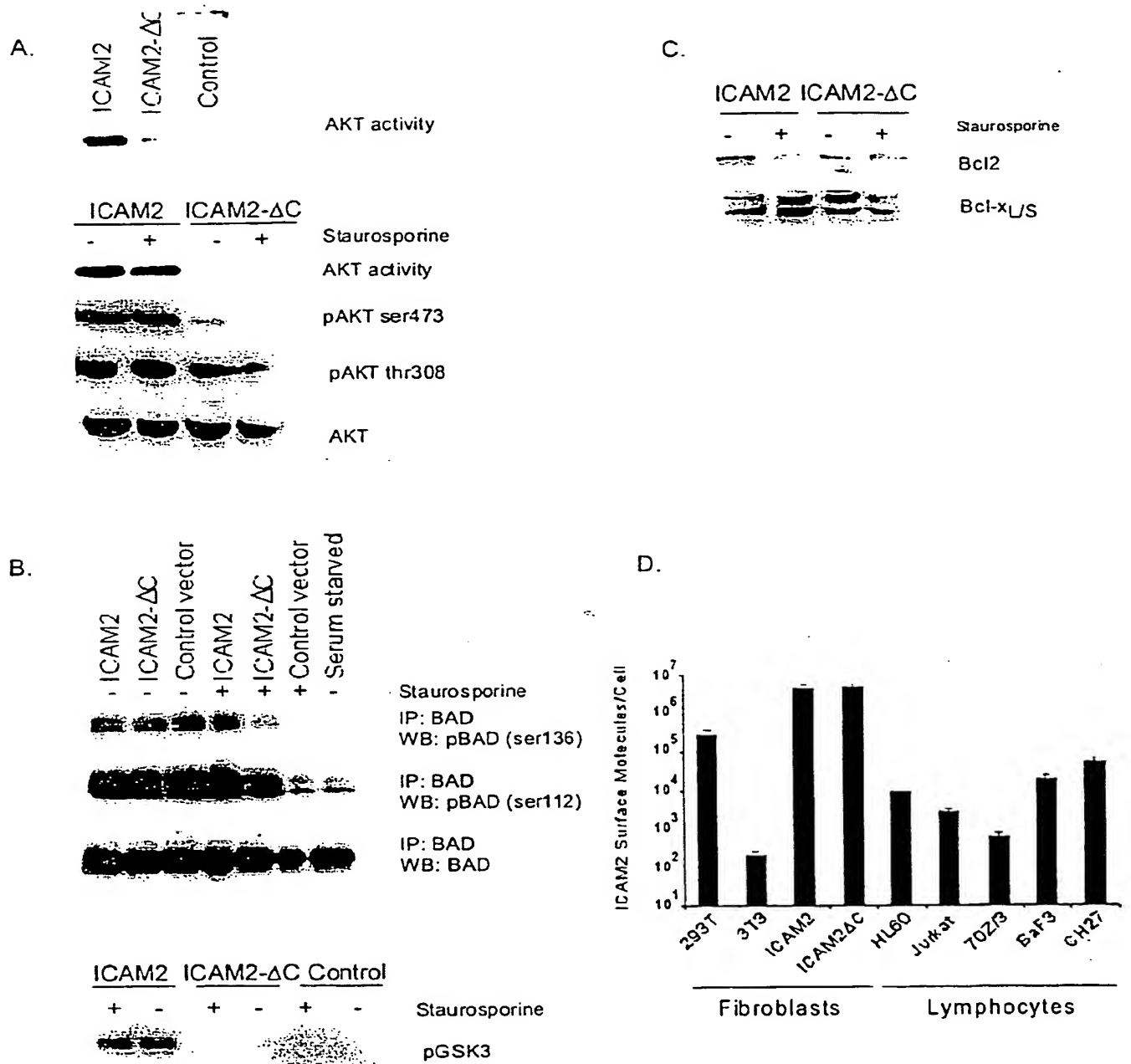
ICAM-2 mediates an anti-apoptotic effect.

FIGURE 28



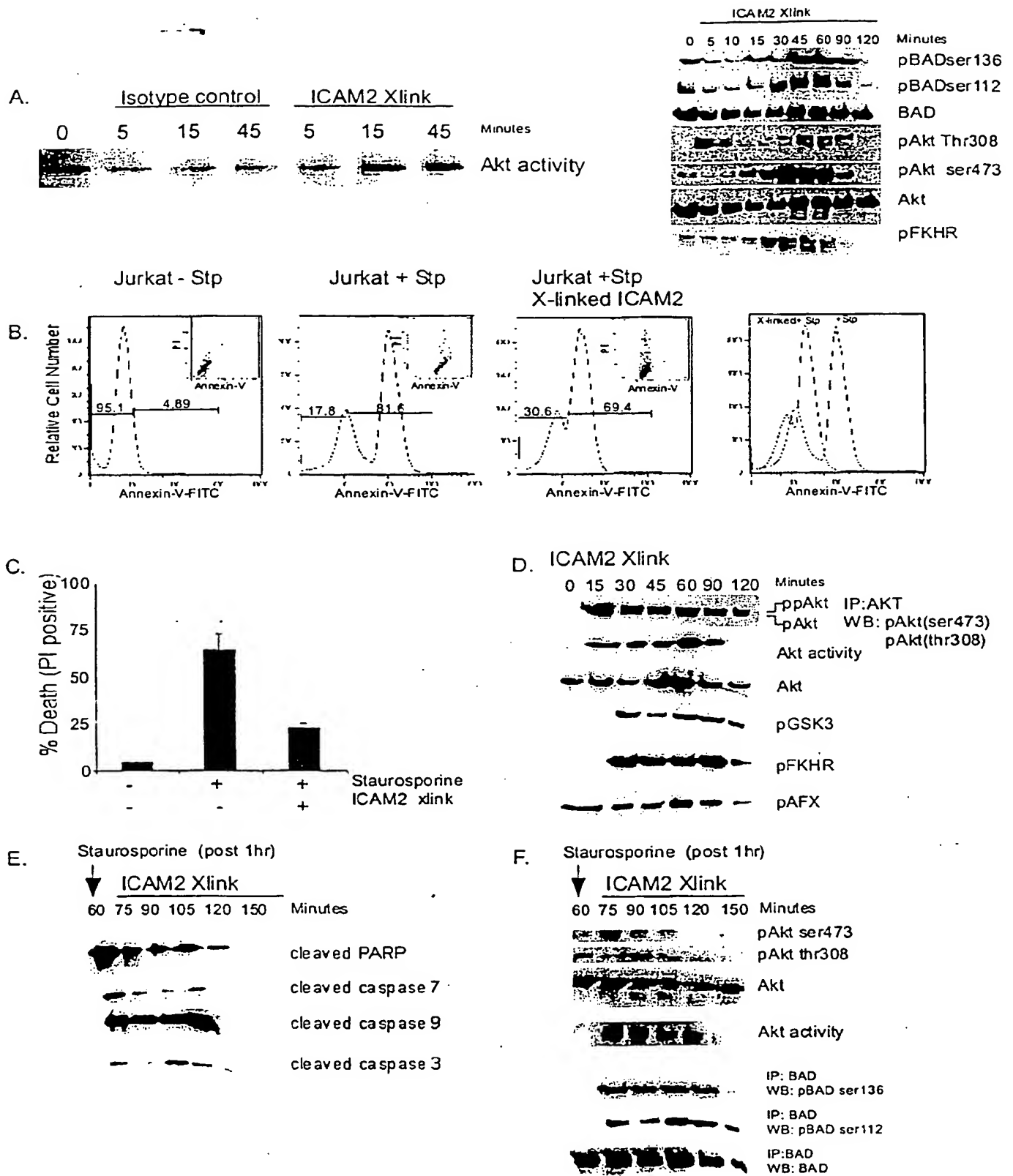
Pharmacological inhibitors of PI3K abrogate ICAM-2's anti-apoptotic effect.

FIGURE 29



ICAM-2 induces AKT kinase activity and phosphorylation of BAD and GSK3.

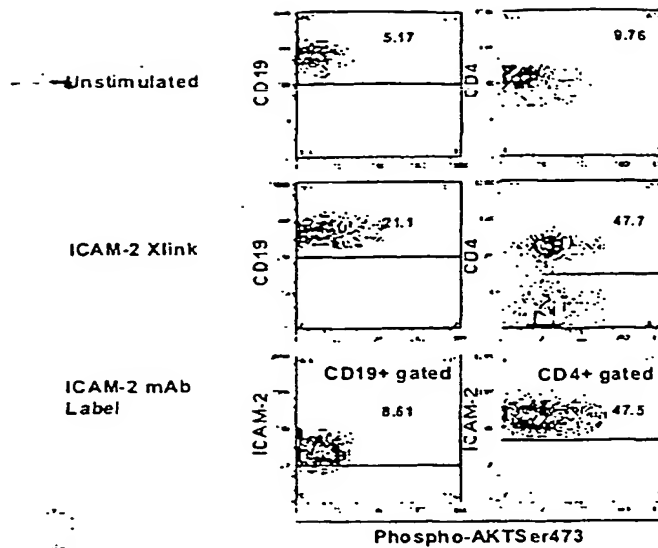
FIGURE 30



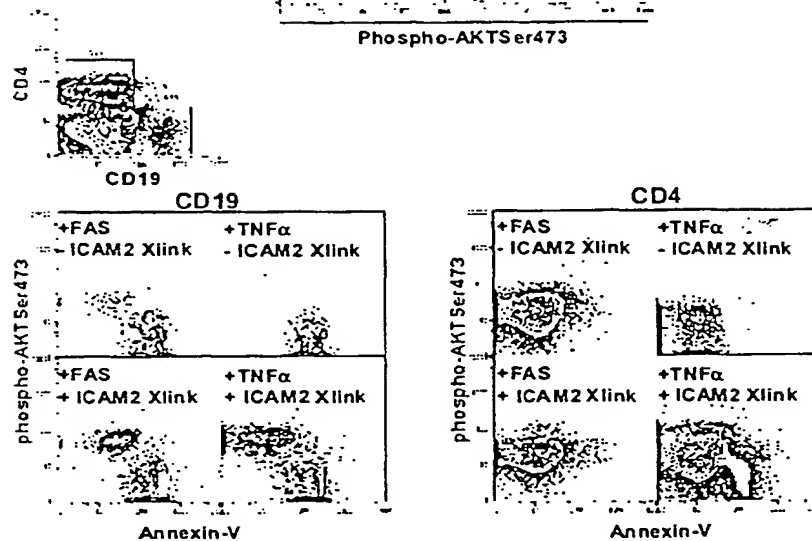
Cross-linking of endogenous ICAM-2 induces AKT activation in BaF3 B-cells

FIGURE 31

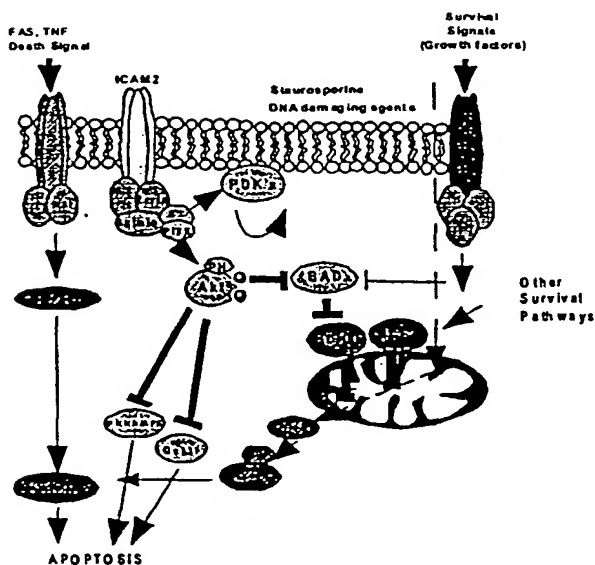
A.



B.



C.



Resistance to TNFα and FAS induced apoptosis in primary human CD4+ and CD19+ cells is mediated by active AKT following ICAM-2 crosslinking.

FIGURE 32

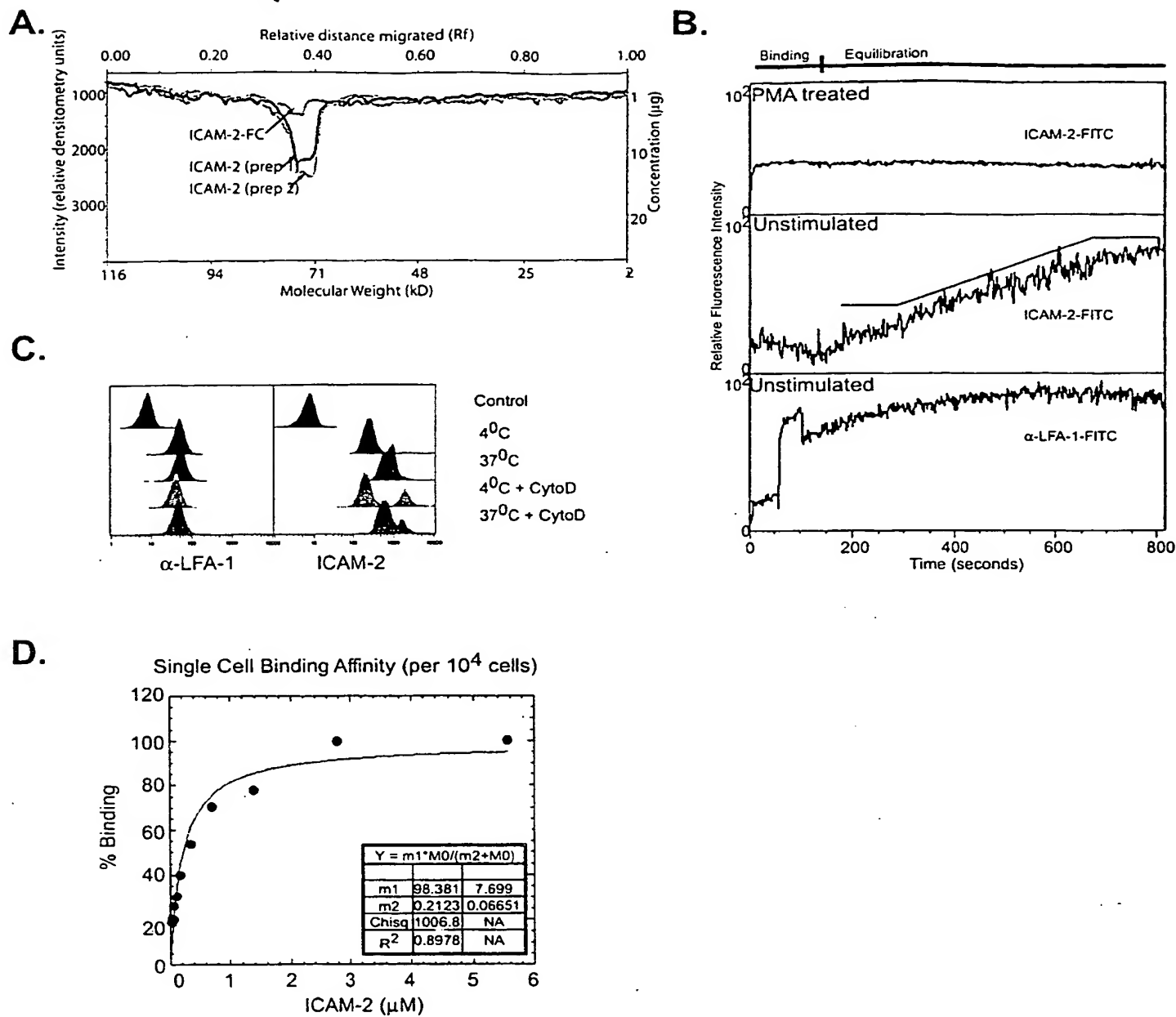


Figure 33

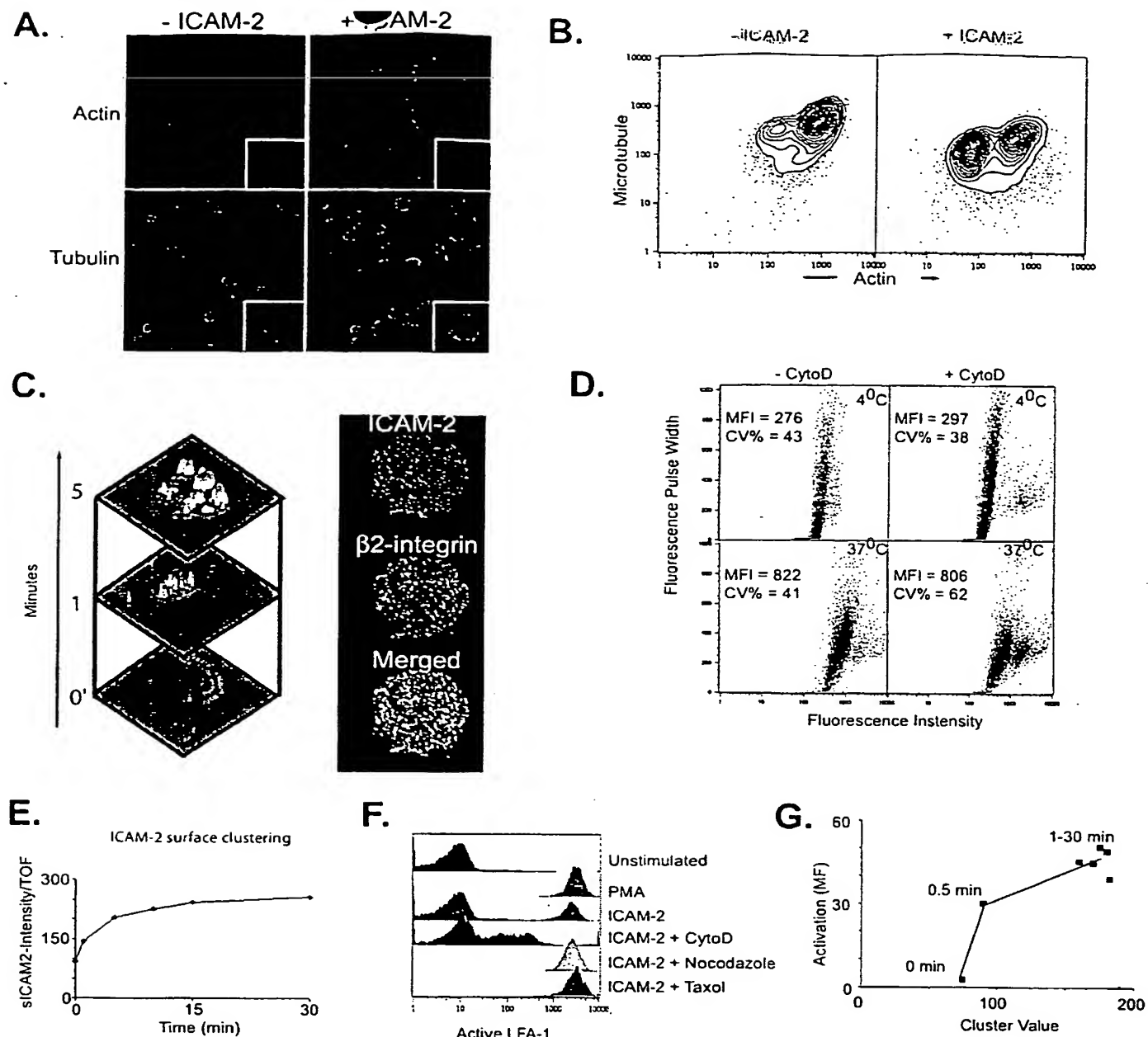
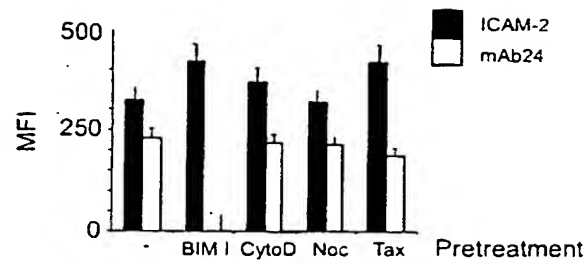
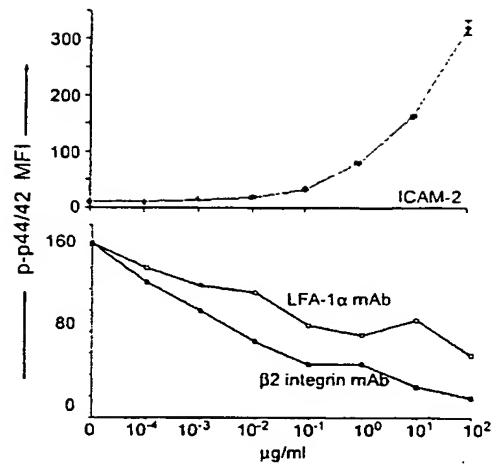


Figure 34

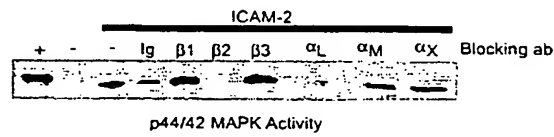
A.



B.



C.



D.

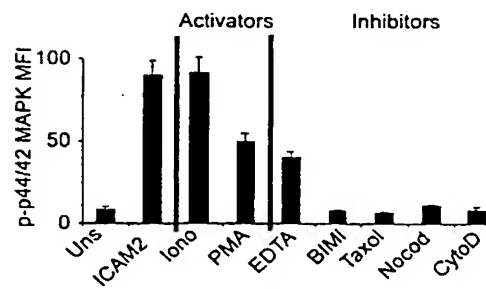
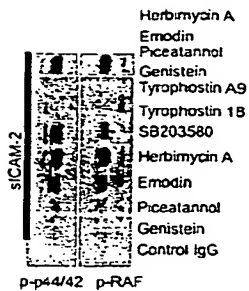
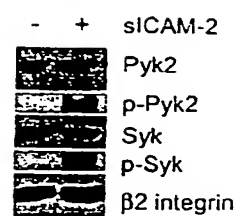


Figure 35

A.



B.



IP: β2 integrin

C.

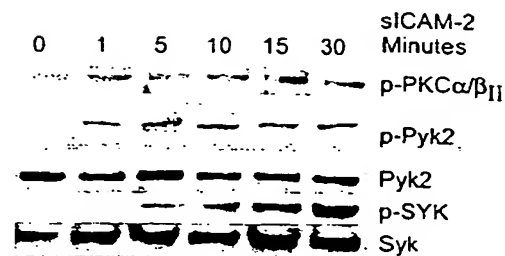
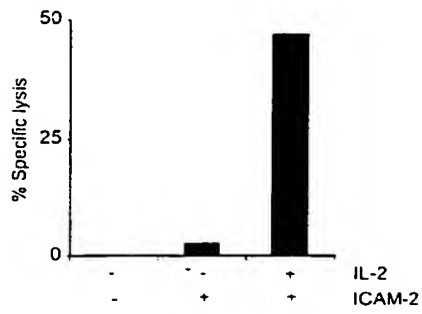


Figure 36

A.



B.

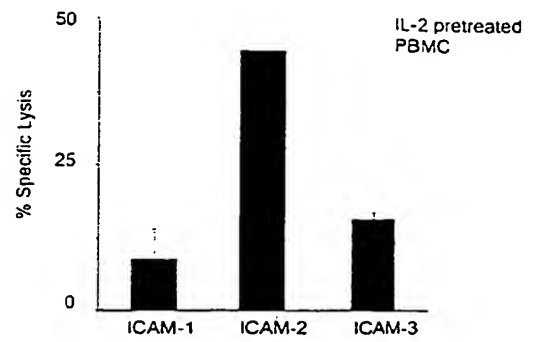
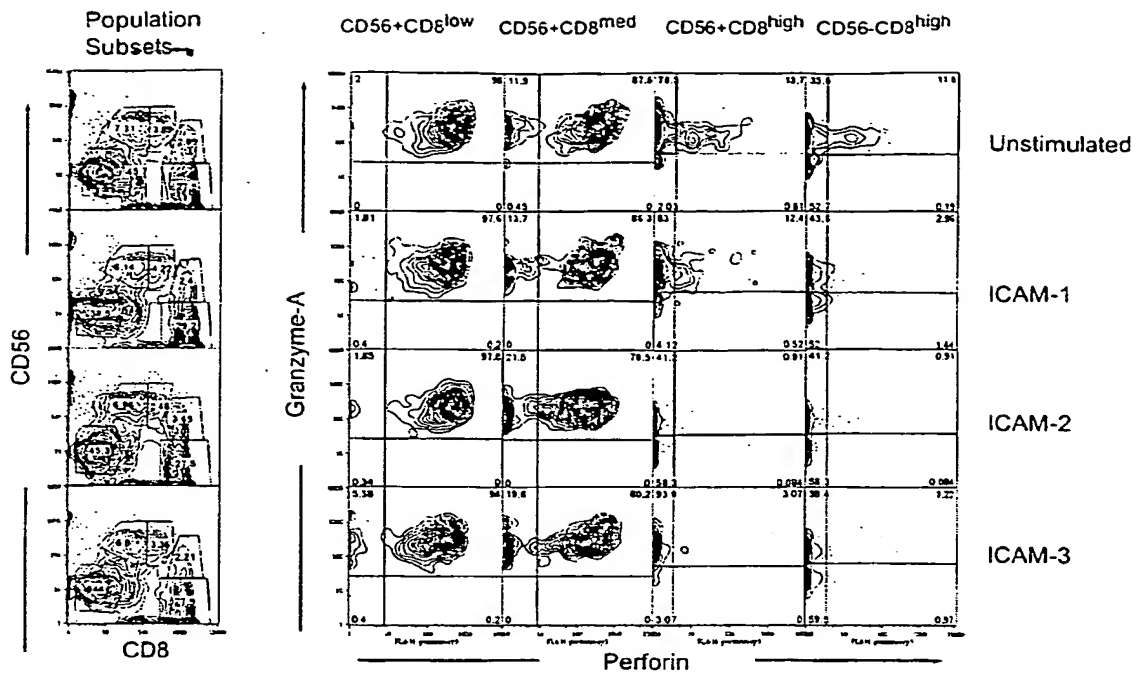
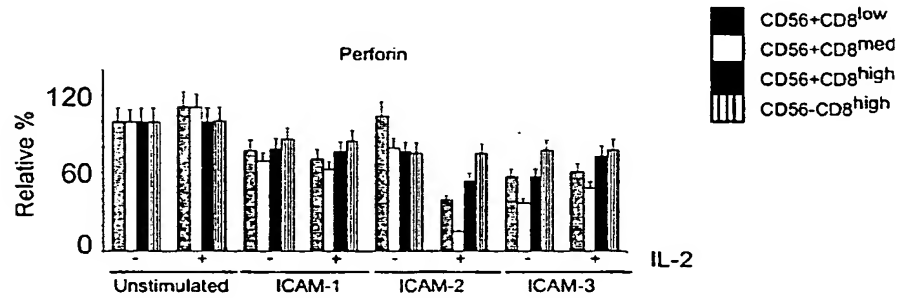


Figure 37

A.



B.



C.

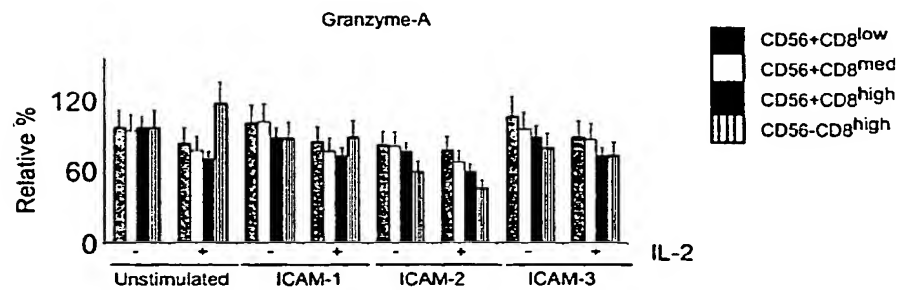
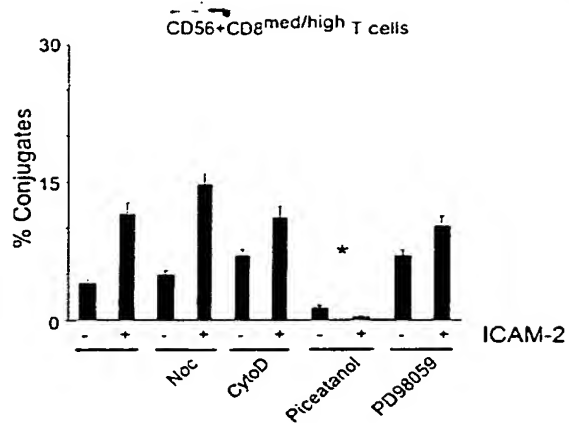


Figure 38

A.



B.

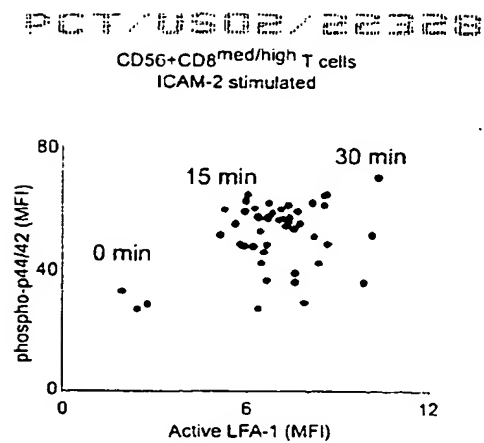
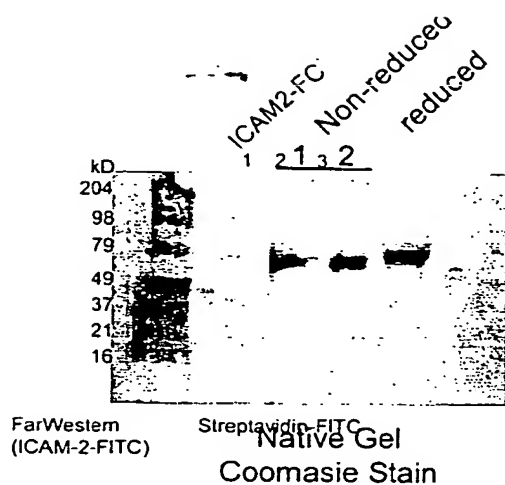
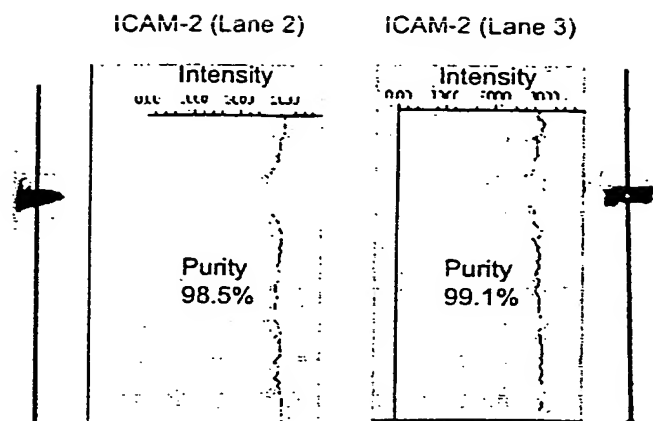


Figure 39

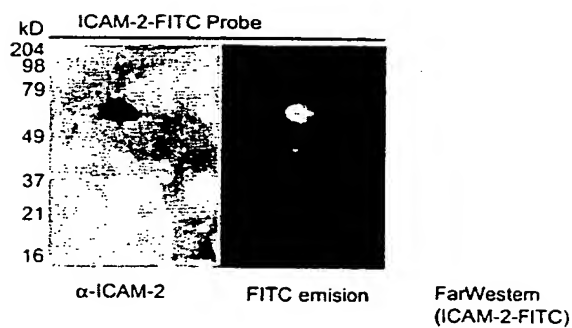
A.



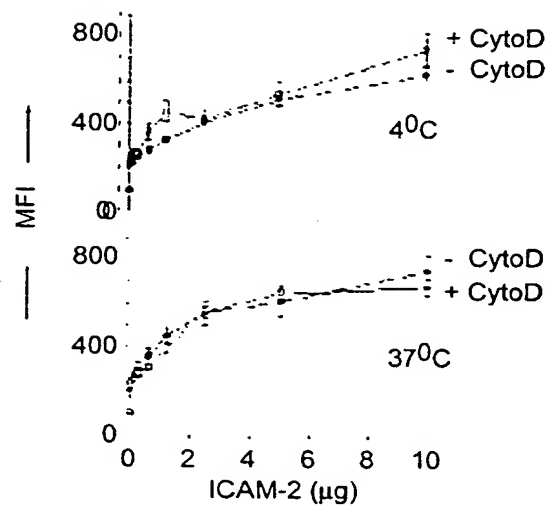
B.



C.



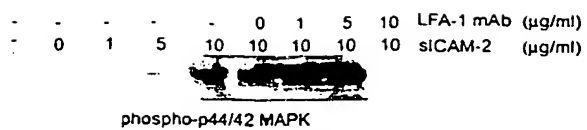
D.



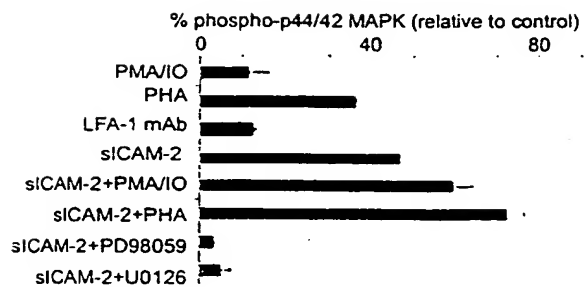
Supplementary Figure 33

Supplementary Figure

A.

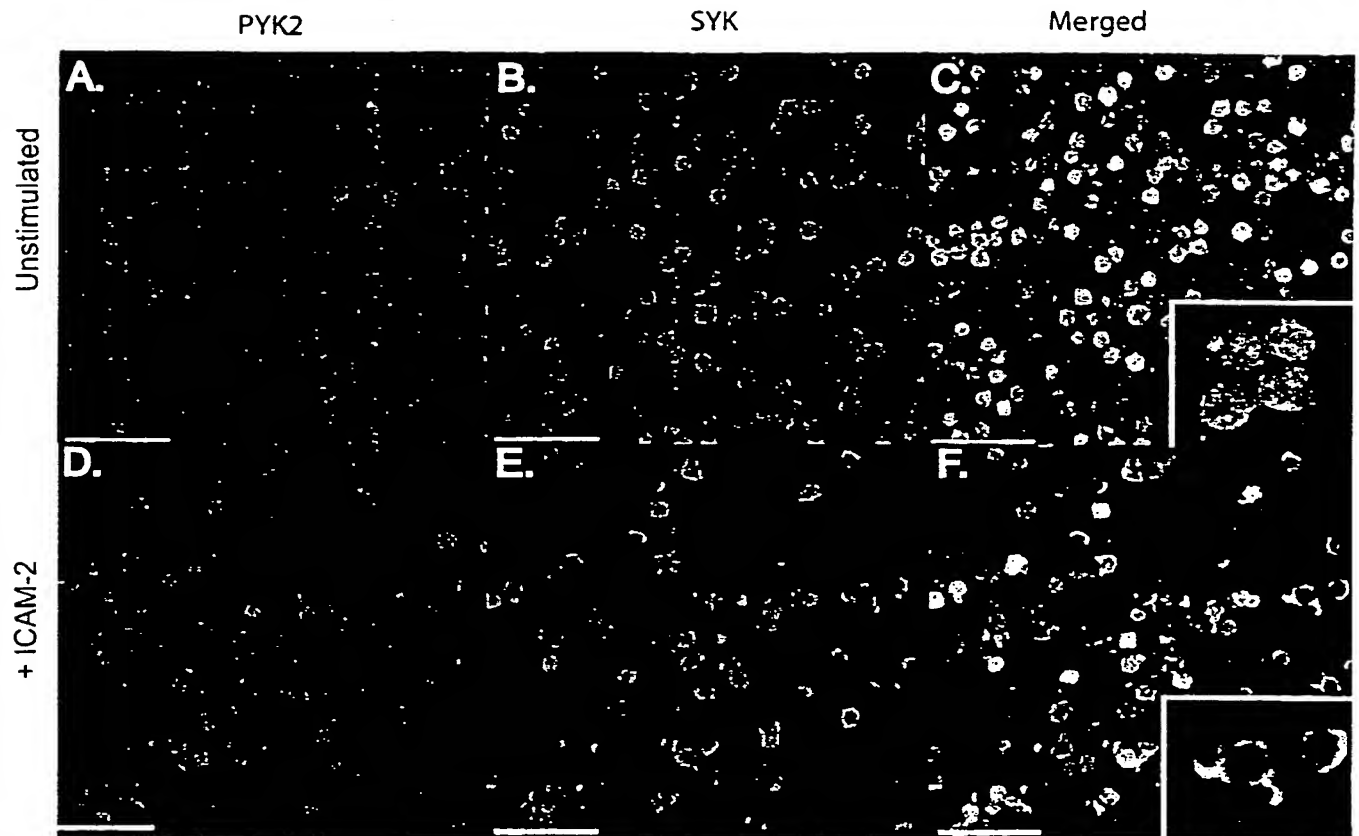


B.



Supplementary Figure 34

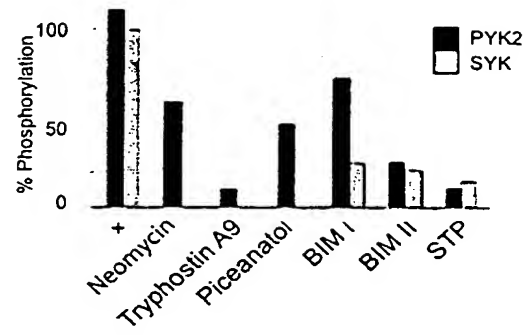
Supplementary Figure



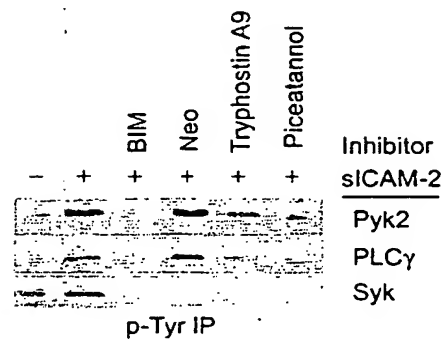
Supplementary Figure 35

A.

Supplementary Figure



B.



Supplementary Figure 36

Table 1: 4, 10, and 11-color FACS staining combinations

405 nm			488 nm excitation					595 nm excitation			
CasBI	CasYel		FITC	PE	CY5 PE	Cy5.5 PE	CY7 PE	Alexa 594	APC	Cy5.5 APC	Cy7 APC
1			p-probe	p-probe				p-probe	p-probe		
2			p-probe	p-probe				CD19	CD4		
3			p-probe	p-probe				CD25	CD69		
4	CD11a	CD8	p-probe	p-probe	CD28	CD4	CD62L	CD27	p-probe	CD4	CD45RA
5	CD11a	CD8	p-probe	p-probe	CD28	CD4	CD62L	p-probe	p-probe	CD4	CD45RA
6	CD11a	CD8	p-probe	p-probe	CD19	CD45RA	CD62L	p-probe	p-probe	CD4	CD16

488 nm excitation			595 nm excitation			
Alexa 488	Alexa 546	Alexa 568	Alexa 594	Alexa 633	Alexa 660	Alexa 680
p-p44/42		p44/42				p-PKA-Substrate
p-PKC-PAN	JNK	p-AKTser473	p-TYK2	p-JNK		p38
AKT	p-p38				p-AKT thr308	
p-Tyrosine					p-threonine	

Table 2 Kinase inducing and inhibitory agents

Kinase	Inducing Agent	Inhibitory Agent	Inhibitory agent specificity
p44/42 MAPK	EGF (10 µg/ml)	U0126 (10 µM)	MEK1/2 upstream kinase of p44/42 MAPK
AKT	PDGF (10 µg/ml)	LY294002 (10 µM)	PI3K upstream kinase of AKT
p38 MAPK	Anisomycin (10 µM)	SB203580 (10 µM)	p38 inhibitor, binds active site
TYK2	IFNγ (10 pg/ml)	Genistein (10 µM)	General tyrosine phosphorylation inhibitor
JNK	Anisomycin (10 µM)	Staurosporine (0.1 µM)	Broad acting kinase inhibitor (specific JNK inhibitor not available)
PKC	PMA/Ionomycin (1 µM)	Bisindoylmaleimide II (10 µM)	PKC inhibitor
PKA	PMA/Ionomycin (1 µM)	Calyculin A (10 µg/ml)	General phosphatase inhibitor for PP2A
Phospho Tyrosine	PMA/Ionomycin (1 µM)	Genistein (10 µM)	General tyrosine phosphorylation inhibitor

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